Check for updates

Received: 7 February 2021 | Revised: 23 March 2021 | Accepted: 28 March 2021

DOI: 10.1111/all.14840

Allergy EUROPEAN JOURNAL OF ALLERGY AND CLINICAL IMMUNOLOGY EAACI WILEY

REVIEW ARTICLE

Vaccines and allergic reactions: The past, the current COVID-19 pandemic, and future perspectives

Vanitha Sampath[1] | Grace Rabinowitz[1] | Mihir Shah[1] | Surabhi Jain[1] | Zuzana Diamant[2,3,4,5] | Milos Jesenak[6] | Ronald Rabin[7] | Stefan Vieths[8] | Ioana Agache[9] | Mübeccel Akdis[10] | Domingo Barber[11,12] | Heimo Breiteneder[13] | Sharon Chinthrajah[1,14] | Tomas Chivato[15] | William Collins[1,16] | Thomas Eiwegger[17,18,19] | Katharine Fast[1] | Wytske Fokkens[20] | Robyn E. O'Hehir[21] | Markus Ollert[22,23] | Liam O'Mahony[24] | Oscar Palomares[25] | Oliver Pfaar[26] | Carmen Riggioni[27,28] | Mohamed H Shamji[29,30] | Milena Sokolowska[10] | Maria Jose Torres[31] | Claudia Traidl-Hoffmann[32,33] | Menno van Zelm[34,35] | De Yun Wang[36] | Luo Zhang[37] | Cezmi A. Akdis[10] | Kari C. Nadeau[1,14]

[1]Sean N. Parker Center for Allergy and Asthma Research at Stanford University, Stanford, CA, USA

[2]Departmentt of Microbiology Immunology & Transplantation, KU Leuven, Catholic University of Leuven, Leuven, Belgium

[3]Department of Respiratory Medicine & Allergology, Institute for Clinical Science, Skane University Hospital, Lund University, Lund, Sweden

[4]Department of Clinical Pharmacy &Pharmacology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands

[5]Department of Respiratory Medicine, First Faculty of Medicine, Charles University and Thomayer Hospital, Prague, Czech Republic

[6]Department of Pediatrics and Department of Clinical Immunology and Allergology, Jessenius Faculty of Medicine in Martin, Center for Vaccination in Special Situations, University Hospital in Martin, Comenius University in Bratislava, Bratislava, Slovakia

[7]Office of Vaccines Research and Review, Center for Biologics Evaluation and Research, US Food and Drug Administration, Silver Spring, MD, USA

[8]Paul-Ehrlich-Institut, Federal Institute for Vaccines and Biomedicines, Langen, Germany

[9]Transylvania University, Brasov, Romania

[10]Swiss Institute of Allergy and Asthma Research (SIAF), University Zurich, Zurich, Switzerland

[11]Departamento de CienciasMédicasBásicas, Facultad de Medicina, Instituto de Medicina Molecular Aplicada (IMMA), Universidad San Pablo-CEU, CEU Universities, Madrid, España

[12]Instituto de Salud Carlos III, RETIC ARADYAL, Madrid, Spain

[13]Division of Medical Biotechnology, Department of Pathophysiology and Allergy Research, Center for Pathophysiology, Infectiology and Immunology, Medical University of Vienna, Vienna, Austria

[14]Department of Medicine, Division of Pulmonary and Critical Care Medicine, Stanford University, Stanford, CA, USA

[15]School of Medicine, University CEU San Pablo, Madrid, Spain

[16]Department of Medicine, Division of Hospital Medicine, Stanford University, Stanford, CA, USA

[17]Division of Immunology and Allergy, Food Allergy and Anaphylaxis Program, The Hospital for Sick Children, Toronto, ON, Canada

[18]Translational Medicine, Research Institute, The Hospital for Sick Children, Toronto, ON, Canada

[19]Department of Immunology, University of Toronto, Toronto, ON, Canada

[20]Department of Otorhinolaryngology, Amsterdam University Medical Centers, Amsterdam, The Netherlands

[21]Department of Allergy, Immunology and Respiratory Medicine, Central Clinical School, Monash University, and Alfred Health, Melbourne, Vic, Australia

[22]Department of Infection and Immunity, Luxembourg Institute of Health, Esch-sur-Alzette, Luxembourg

Sampath, Rabinowitz and Shah are co-first authors.

Akdis and Nadeau are co-senior authors.

© 2021 European Academy of Allergy and Clinical Immunology and John Wiley & Sons Ltd.

[23]Department of Dermatology and Allergy Center, Odense Research Center for Anaphylaxis, Odense University Hospital, University of Southern Denmark, Odense, Denmark

[24]Department of Medicine and School of Microbiology, APC Microbiome Ireland, University College Cork, Cork, Ireland

[25]Department of Biochemistry and Molecular Biology, School of Chemistry, Complutense University, Madrid, Spain

[26]Department of Otorhinolaryngology, Head and Neck Surgery, Section of Rhinology and Allergy, University Hospital Marburg, Philipps-Universität Marburg, Marburg, Germany

[27]Department of Paediatrics, Allergy and Clinical Immunology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore

[28]Institut de Recerca Sant Joan de Déu, Barcelona, Spain

[29]Department of National Heart and Lung Institute, Immunomodulation and Tolerance Group, Allergy and Clinical Immunology, Imperial College London, London, UK

[30]Centre in Allergic Mechanisms of Asthma, London, UK

[31]Allergy Unit, Malaga Regional University Hospital-UMA-ARADyAL, Málaga, Spain

[32]Department of Environmental Medicine, Medical Faculty, University Augsburg, Augsburg, Germany

[33]Institute of Environmental Medicine, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany

[34]Department of Immunology and Pathology, Monash University, Melbourne, VIC, Australia

[35]Allergy, Asthma and Clinical Immunology, Alfred Health, Melbourne, VIC, Australia

[36]Department of Otolaryngology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore

[37]Department of Otolaryngology Head and Neck Surgery, Beijing TongRen Hospital, Capital Medical University, Beijing, China

Correspondence
Kari C. Nadeau, Sean N. Parker Center for Allergy and Asthma Research at Stanford University, Biomedical Innovations Building (BMI), 240 Pasteur Drive, Room#1755, Palo Alto, CA 94304, USA.
Email: knadeau@stanford.edu

Abstract

Vaccines are essential public health tools with a favorable safety profile and prophylactic effectiveness that have historically played significant roles in reducing infectious disease burden in populations, when the majority of individuals are vaccinated. The COVID-19 vaccines are expected to have similar positive impacts on health across the globe. While serious allergic reactions to vaccines are rare, their underlying mechanisms and implications for clinical management should be considered to provide individuals with the safest care possible. In this review, we provide an overview of different types of allergic adverse reactions that can potentially occur after vaccination and individual vaccine components capable of causing the allergic adverse reactions. We present the incidence of allergic adverse reactions during clinical studies and through post-authorization and post-marketing surveillance and provide plausible causes of these reactions based on potential allergenic components present in several common vaccines. Additionally, we review implications for individual diagnosis and management and vaccine manufacturing overall. Finally, we suggest areas for future research.

KEYWORDS
allergy, anaphylaxis, COVID-19, SARS-CoV-2, vaccine

1 | INTRODUCTION

The rapid development and the launch of several novel COVID-19 vaccines for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is an extraordinary and remarkable accomplishment of modern science. The Pfizer-BioNTech BNT162b2 was the first vaccine to be granted temporary authorization for emergency use by the Medicines and Healthcare Products Regulatory Agency (MHRA) in the U.K for the treatment of COVID-19 on 2 December 2020.[1] Soon after, on 11 December 2020, it also received emergency use authorization (EUA) by the U.S. Food and Drug Administration (FDA).[2] EUA is a mechanism to facilitate the availability of vaccines during public health emergencies, such as the current COVID-19 pandemic. Under an EUA, the FDA may allow the use of unapproved medical products (including vaccines) to prevent serious or life-threatening disease when certain statutory criteria have been met and no adequate and/or approved alternatives are available. The authorization of BNT162b2 was followed by an EUA for a second COVID-19 vaccine, the Moderna mRNA-1273 on 18 December 2020.[3] This was followed by the authorization of mRNA-1273 for use by other regulatory agencies such as the European Commission, UK MHRA, Israel Ministry of

TABLE 1 Key Characteristics of COVID-19 vaccines (approved and in phase 3 clinical trials)[7,162]

Developers	Vaccine Name (s)	Vaccine Platform	Authorization Status	Phase 3 Efficacy	Prod. capacity	% of doses to HICs for 2021	Agreement with COVAX?	No of Dose	Temp. for storage (Celsius)
Approved vaccines									
AstraZeneca with Oxford University	AZD1222	Viral vector (non-replicating)	Emergency use in U.K., E.U., other countries.	62%	3bn	27%	Yes	2	2–8
Bharat Biotech International Limited	BBV152, Covaxin	Inactivated virus	Emergency use in India.	–	700 m	0%	No	2	2–8
BioNTech with Pfizer+Fosum Pharma	BNT162b2, tozinameran, Comirnaty	RNA-based	Approved in several countries. Emergency use in United States, E.U., other countries.	95%	2bn	77%	Yes	2	−70
CanSino Biological Inc./ Beijing Institute of Biotechnology	Ad5-nCoV, Convidecia	Viral vector (non-replicating)	Limited use in China. Emergency use in Mexico, Pakistan.	65.7%	320 m	0%	No	1	2–8
Gamaleya Research Institute and Health Ministry of the Russian Federation	Sputnik V	Viral vector (non-replicating)	Early use in Russia. Emergency use in other countries.	92%	1bn	0%	No	2	−18
Moderna +National Institute of Allergy and Infectious Diseases (NIAID)	mRNA−1273	RNA-based	Approved in Switzerland. Emergency use in U.S., U.K., E.U., others	94%	1bn	97%	No	2	−20
Sinopharm with China National Biotec Group Co and Beijing Institute of Biological Products	BBIBP-CorV	Inactivated virus	Approved in China, U.A.E., Bahrain. Emergency use in Egypt, other countries.	79%	1bn	8%	No	2	2–8
Sinopharm and China National Biotec Group Co and Wuhan Institute of Biological Products	N/A	Inactivated virus	Limited use in China, U.A.E.	–	600 m	8%	No	2	2–8
Sinovac and Development Co., Ltd	CoronaVac	Inactivated virus	Approved in China. Emergency use in Brazil, other countries	50–91%	1bn	18%	No	2	Room temp.
Vector Institute	EpiVacCorona	Protein subunit	Early use in Russia	N/A	11 m	–	No	2	2–8
Vaccines in phase 3 trials (unapproved)									
AnGesand Takara Bio and Osaka University	AG0302-COVID19	DNA-based	Not approved	–	–	–	No	2	−70

(Continues)

TABLE 1 (Continued)

Developers	Vaccine Name (s)	Vaccine Platform	Authorization Status	Phase 3 Efficacy	Prod. capacity	% of doses to HICs for 2021	Agreement with COVAX?	No of Dose	Temp. for storage (Celsius)
Anhui ZhifeiLongcom Biopharmaceutical and Institute of Microbiology, Chinese Academy of Sciences	ZF2001	Protein subunit	Not approved	–	300 m	–	No	2 or 3	2–8
Biological E Limited and Dynavax and Baylor College of Medicine	N/A		Not approved	–	–	–	No	2	2–8
Chinese Academy of Medical Sciences and Institute of Medical Biology	N/A	Inactivated virus	Not approved						
Clover Pharmaceuticals Inc.,/ GSK/Dynavax	COVID-19 S-Trimer	Protein subunit	Not approved	–	1bn	–	No	2	2–8
Covaxx with Nebraska University and United Biomedical Inc	UB-612	Protein subunit	Not approved	–	1bn	0%	No	2	2–8
CureVacand Bayer	CVnCoV	RNA-based	Not approved	–	300 m	100%	No	2	5
Inovio Pharmaceuticals and International Vaccine Institute and Advaccine (Suzhou) Biopharmaceutical Co., Ltd	INO-4800	DNA-based	Not approved	–	100 m	–	No	2	2–8
Johnson & Johnson	Ad26		Not approved	66%	1bn	38%	Yes	1	2–8
Medicago, GSK	CoVLP	Virus-like particle	Not approved	–	80 m	100%	No	2	2–8
Novavax	NVX CoV2373	Protein subunit	Not approved	89%	2bn	31%	Yes	2	2–8
RIBSP	QazCovid-in®		Not approved	–	60 m	–	No	2	2–8
Research Institute for Biological Safety Problems	QazCovid	Inactivated virus	Not approved						
Sanofi with GlaxoSmithKline	N/A		Not approved	–	–	73%	Yes	2	2–8
SII with Max Planck Institute	VPM1002		Not approved	–	–	–	No	–	−50 to −15
SK Biosciences	GBP510		Not approved	–	–	–	No	–	2–8
University of Hong Kong	N/A		Not approved	–	–	–	No	–	−50 to −15
Zydus Cadila	ZyCoV-D	DNA-based	Not approved					3 doses (by skin patch)	

Health, and others.[4] On December 30, a third COVID-19 vaccine, the Oxford/AstraZeneca recombinant adenoviral AZD1222 or ChAdOx1-S was authorized for use by the UK MHRA.[5] In addition to the above authorized COVID-19 vaccines, a number of other novel COVID-19 vaccines are currently in different phases of clinicals development. Currently, used platforms in COVID-19 vaccines include classical and novel platforms, such as RNA- and DNA-based, viral vectors (non-replicating), protein subunits, virus-like particles, and inactivated viral platform (Table 1).[6–8]

With the authorization of COVID-19 vaccines, vaccination campaigns have been initiated in many areas throughout the world. Within the first few days of public vaccinations, however, BNT162b2 was associated with a few severe cases of anaphylaxis.[9] While severe allergic reactions may pose a potential risk with any vaccine (or systemic medications), the benefits of vaccination outweigh the potential risks of receiving the vaccine for the vast majority of individuals. However, the fear of allergic reactions may lead to vaccine hesitancy, which could compromise herd immunity and limit efforts to contain the pandemic. It is therefore critical that we understand the immunopathological changes in COVID-19,[10] risk of severe allergic reactions, and their mechanisms in order to improve individual safety and issue proper guidance with the goal of vaccinating the maximum number of individuals. Here, we review adverse events associated with vaccine-related allergic and non-allergic reactions to COVID-19 and other vaccines, mechanisms associated with allergic adverse reactions, and rates of severe allergic reactions with existing vaccines and with the novel COVID-19 active vaccines currently being administered to large parts of the world.

2 | SECTION I: VACCINE-ASSOCIATED ALLERGIC VS NON-ALLERGIC REACTIONS

There are a number of vaccines currently used with proven safety and efficacy. Vaccines are potentially associated with adverse events. An adverse event is defined as any untoward medical occurrence associated with the use of a drug in humans, whether or not considered drug related.[11] Adverse events can present as local or systemic, immediate or non-immediate, and immune or non–immune-mediated reactions. While all allergic reactions are immune-mediated, not all immune-mediated reactions are allergic. Local non-immediate reactions that are not allergenic are common and may include swelling and erythema at the injection site. These reactions can occur hours or days after administration and are not always mediated through the immune system. Systemic non-allergic reactions including mild fever and vasovagal reactions such as hypotension, nausea, and syncope are also relatively common. Neither the local nor the vasovagal reactions pose any serious risk. Although some of these reactions are immune-mediated, they are not allergic reactions. Rather, soreness at the injection site or fatigue are consequences of activation of the innate immune system.[12–15]

As of 10 February 2021, 44.77 million people had taken one or two doses of a COVID vaccine in the United States. 653 deaths and 12,697 total adverse events had been reported to following COVID-19 vaccinations to the CDC and Prevention's VAERS.[16] Adverse events, including allergic reactions, are graded according to severity as mild, moderate, and for purposes of this review, serious. Typical signs of an allergic reaction include bronchoconstriction, conjunctivitis, rhinorrhea, gastrointestinal symptoms, and/or characteristic skin lesions such as generalized urticaria and/or angioedema. These can occur in combination or alone, and onset can be immediate, within minutes, or up to several hours post-vaccination. Examples of mild allergic reactions are swelling with itching at the injection site, conjunctivitis, or rhinorrhea. Examples of moderate allergic reactions are bronchoconstriction that can be adequately treated with nebulized beta-agonists or generalized urticaria that may be treated with an antihistamine. Serious adverse events (SAE) are those events that are life-threatening, require inpatient hospitalization or prolongation of existing hospitalization, cause a persistent or significant incapacity or substantial disruption in the ability to conduct normal life functions, a congenital anomaly/birth defect, or death. Two examples of SAE that are allergic reactions are bronchospasm that requires intensive treatment and life-threatening anaphylaxis.[11,17]

Anaphylaxis, an immediate systemic multi-organ reaction, is rare but can be life-threatening. Organs affected include the cutaneous, gastrointestinal, respiratory, and cardiovascular systems. Anaphylaxis can be either immunological, non-immunological, or idiopathic. Idiopathic anaphylaxis is diagnosed through exclusion of other known causes and may mask a clonal mast cell disorder.[18–23] Non-immunological anaphylaxis was previously termed anaphylactoid reactions, but the World Allergy Organization (WAO) in 2004 suggested replacing anaphylactoid reactions with non-immunological anaphylaxis.[24] The change in terminology is to reinforce the risk and potential fatality of all types of anaphylaxis, regardless of the underlying mechanism. All three mechanisms of anaphylaxis produce the same clinical picture (see Section IV on mechanisms below). Distinguishing between systemic vasovagal reactions and anaphylaxis during immunization is critical to ensure that appropriate and immediate treatment can be administered (Table 2). Vasovagal reactions usually occur immediately or up to 30 minutes of vaccine administration. Similar to anaphylaxis, organs affected include the cutaneous, gastrointestinal, respiratory, neurological, and cardiovascular systems.[25,26]

Anaphylactic reactions are considered adverse events of special interest (AESI),[27] that is, adverse events that are of significant medical and scientific concern for which immediate medical action with ongoing monitoring and rapid communication by the investigator or sponsor is required. AESI reporting is a critical aspect of pharmacovigilance for characterization of the safety profile of a drug or vaccine in context of previous reports of the vaccine or of other vaccines with similar manufacturing processes, formulation, immunogenicity, and novelty. AESIs alert regulators to potential risks. Particularly in mass vaccination programs where a large number of adverse reactions may be reported, identification and assessment of AESIs are a high priority because they highlight potential risks that may alter risk-benefit profile and

TABLE 2 Differentiating features of vasovagal episode vs. anaphylaxis[25,26]

Vasovagal Episode	Anaphylaxis
Onset	
Immediately or up to 30 minutes after vaccine administration	Can be immediate or within minutes or up to several hours after vaccine administration
Respiratory	
Normal respiration - may be shallow, but not labored	Cough, wheeze, stridor, hoarseness, rhinorrhea Signs of respiratory distress (tachypnea, cyanosis, and rib recession) Upper airway swelling (lip, throat, tongue, uvula, or larynx)
Cardiovascular	
Bradycardia—weak/absent peripheral pulse- but with strong central pulse (carotid) Hypotension—usually transient and corrects in supine position Loss of consciousness—improves once supine or head-down position	Tachycardia, weak/absent central pulse Hypotension—sustained and no improvement without specific treatment (in infants and young children, limpness and pallor are signs of hypotension) Loss of consciousness—no improvement once supine or in head-down position
Skin	
Generalized pallor, cool clammy skin	Urticaria, angioedema, pruritus, erythema
Gastrointestinal	
Nausea or vomiting	Abdominal cramps, diarrhea, nausea, or vomiting
Neurological	
Nausea or vomiting	Feels faint, light-headed, headache, dizziness blurred vision, restless, seldom: seizures

may require immediate investigation, regulatory action, and prompt communication with the public.[28,29] Table 3 lists Pharmacovigilance Practices that follow authorization of a vaccine.

3 | SECTION II: ALLERGIC REACTIONS TO VACCINES

In the last 120 years, global vaccination programs have eradicated or vastly reduced the incidence of debilitating infectious diseases such as smallpox, polio, and measles.[30] According to the Institute of Medicine, epidemiologic and mechanistic evidence support a causal relationship between anaphylaxis and several vaccines, including those for measles, mumps, and rubella (MMR), varicella, influenza, hepatitis B, meningococcus, human papillomavirus, and the combined diphtheria, tetanus, pertussis (DTaP or TdaP) vaccine.[31] Although approved vaccines have been rigorously tested for safety, anaphylactic reactions, although rare, can occur in individuals.[32] An analysis of reported anaphylaxis to the Vaccine Adverse Reporting System (VAERS) in the United States over a 26-year period found that out of the almost 500,000 reports to VAERS, only 828 were classified as anaphylaxis based on either on physician's diagnosis or in according to the Brighton Collaboration case standards.[33] A 2003 study analyzing over eight million routine vaccinations in the Vaccine Safety Datalink (VSD) Project[34] found the risk of anaphylaxis ranged between 0.65 and 1.53 cases/million doses. They also noted that most anaphylactic episodes occurred when multiple vaccines were administered during the same visit.[35] Similarly, a 2016 study used health data from VSD and found 33 confirmed cases of anaphylaxis after 25,173,965 vaccine doses or an anaphylaxis rate of 1.31 per million vaccine doses.[36] The study also found that 85% of cases of anaphylaxis had pre-existing atopic disease, which was consistent with earlier reports emphasizing coexisting atopic disease, particularly asthma, as being clinical risk factors for anaphylaxis.[37] In Asian children, analysis of a large-linked database found risk of anaphylaxis to be 1.21 cases/million doses.[38] A study conducted in Australia found that estimated incidence rate of anaphylaxis for DTaP vaccines was 0.36 cases per 100,000 doses, and 1.25 per 100,000 doses for MMR vaccines.[39] Overall, rates of reported anaphylaxis occur at a rate of about 1 per 100,000 to one per 1,000,000 depending on the vaccine.[40] Figure 1 and Table 4 show the frequency of anaphylaxis for specific vaccines. Table 5 provides population-specific considerations for common vaccines.[41–44]

4 | SECTION III: ALLERGIC REACTIONS TO COVID-19 MRNA VACCINES

Development of the SARS-CoV-2 mRNA vaccine occurred in record time. So far, three candidate vaccines (mRNA BNT162b2, mRNA-1273, and adenovirus vectored ChAdOx1) have been authorized for COVID-19 in the European Union and the United Kingdom. Of these, mRNA BNT162b2 and mRNA-1273 are authorized for emergency use in the United States. Additional candidates have entered or are

TABLE 3 General pharmacovigilance practices for monitoring vaccine reactions[163]

Goal	Steps
Observed vs. expected (O/E) analysis of AESIs during a mass vaccination program	• Collect background incidence rates of AESIs • Create system to process and display real-time vaccination data
Routine pharmacovigilance	• Provide AESI standard case definitions • Present age-stratified data on AESI incidence rate in target population
Follow-up for an adverse reaction	Collect data on: • Patient • Adverse reaction • Vaccination history • Vaccination and diluent (including manufacturer, batch number, batch release specifications, expiry date, laboratory test results about the batch) • Route of administration • Storage and handling conditions

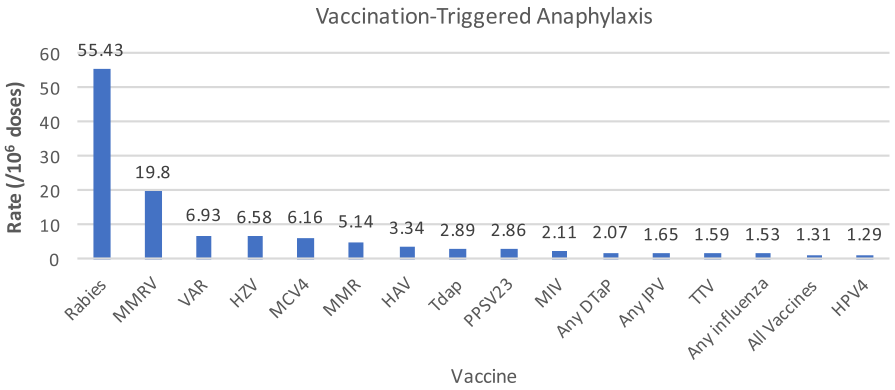

FIGURE 1 Vaccination-triggered anaphylaxis rates for major vaccines[19]

completing the pivotal stage of clinical development programs. At the time of this review, there are 64 vaccines in several stages of clinical development and 173 in pre-clinical development worldwide.[45] Details of their composition, mode of action, and developmental stage can be found in the review by Rodriguez-Coira et al.[46]

Even after formal authorization, vaccine rollout in the United Kingdom, United States, and European Union has been and continues to be challenging. Navigating complex distribution logistics, determining ethical allocation of a limited resource, and de-mystifying widespread news coverage of anaphylactic events that perpetuate vaccine hesitancy is among the most pressing. While anaphylaxis after routine vaccination is very rare, it is important for the scientific community to be informed and prepared to treat adverse vaccine reactions to increase safety and acceptability. During the COVID-19 pandemic, vast quantities of vaccine are expected to be administered over a very short period of time, with increased public awareness and surveillance. In this situation, the probability of reporting anaphylaxis likely increases without the added context of the denominator, which is the millions of individuals receiving the vaccine.

Of the vaccines authorized for use in Europe and the United States, the first vaccine to be administered and distributed was the mRNA BNT162b2 vaccine in the UK. Following two reports of allergic reactions in the UK on 8 December 2020, the MHRA updated its guidelines to state that individuals should not receive the vaccine if they have had previous anaphylaxis to a vaccine, medicine, or food. In addition, MHRA recommended that individuals who experience anaphylaxis after their first dose of the mRNA BNT162b2 vaccine should not receive the second dose and that everyone should be monitored for a minimum of 15 min after vaccination. However, on December 30, after a review of additional data, the guidelines were further updated to indicate that anyone with a previous history of allergic reactions to the ingredients of the vaccine should not receive it, but those with any other allergies such as a food allergy can receive the vaccine.[47]

A CDC report of the VAERS monitoring database indicates that between 14 December 2020 and 23 December 2020, 1,893,360 first doses of the mRNA BNT162b2 COVID-19 vaccine were administered, and 21 cases of anaphylaxis were reported (11.1 cases per million doses). Seventy-one percent of these reactions occurred within 15 minutes of vaccination.[48] Similarly, the CDC reported that between 21 December 2020 and 10 January 2021, 10 cases of anaphylaxis were reported after administration of 4,041,396 first doses of the mRNA-1273 vaccine (2.5 cases per million doses administered). In nine cases, onset occurred within 15 minutes of vaccination. No anaphylaxis-related deaths were reported.[49] This suggests that the incidence of anaphylaxis in the mRNA BNT162b2 (11.1 cases per million doses) and mRNA-1273 COVID-19 vaccines (2.5 cases per million doses) may be about 2 to 8.5 times as high as the incidence reported in the 2016 VSD study for all vaccines (1.31 per million doses). The US FDA authorized labeling currently lists past severe allergic reactions (eg, anaphylaxis) to any component of the vaccine as a contraindication to the mRNA BNT162b2 vaccine.

TABLE 4 Frequency of anaphylaxis after vaccines

Vaccine	Anaphylaxis rate per 10^6 doses	Comment	Reference
Rabies	55.43–86.1		McNeil et al JACI[36]
HPV	1.29–26	Different rates according to the type of HPV vaccine	Brotherton et al[164] Erlewyn-Lajeunesse et al[165] McNeil et al JACI[36]
TBE	20	Polygeline-free TBE vaccine with lower rate of anaphylaxis	Zent et al[166]
MMRV	19.8		McNeil et al JACI[36]
BNT162b COVID-19	11		CDC MMWR[48]
mRNA 1273	2.5		CDC MMWR[45]
Varicella	1.2–10.3	Gelatin-free varicella vaccine with lower rate of anaphylaxis	Sakaguchi et al[95] Ozaki et al[167] Su et al[33]
Herpes zoster	6.16–9.6		McNeil et al JACI[36]
Pandemic A/H1 N1 vaccine	6.8–8	Increased risk of anaphylaxis compared with seasonal influenza vaccines	Rouleau et al[168]
Yellow fever	7.6		Kelso et al[169]
MMR	0.6–5.14	Egg allergens no longer clinically relevant	D'Souza et al[170] Pool et al Pediatrics[93] McNeil et al JACI[36] Su et al[33]
MCV4	6.16		McNeil et al JACI[36]
HAV	3.34		McNeil et al JACI[36]
DTaP	0.51–3.6	During last two decades, the content of gelatin in this type of vaccine decreased followed by decreased frequency of anaphylaxis	Cheng et al[39]
PPSV23	0.2–2.48		McNeil et al JACI[36] Su et al[33]
Influenza	0.1–1.83	No significant differences by types of vaccine or manufacturer 2-fold higher rate for LAV compared with IIV regarding immediate hypersensitivity reactions	Kawai et al[171] Ropero-Alvarez et al[172] McNeil et al JACI[36] Halsey et al[173] Su et al[33]
HBV	0–1.67		McNeil et al JACI[36] Duclos[174]
Japanese encephalitis	0–0.26	Anaphylaxis reported in live attenuated vaccine	Li et al[175] McNeil et al JACI[36]
Hib	0	Very rare event	McNeil et al JACI[36]
PCV13	0	Very rare event	McNeil et al JACI[36]
Rotavirus vaccines	0	Very rare event	McNeil et al JACI[36]

Abbreviations: BNT162b2, mRNA vaccine against COVID-19; DTaP, diphtheria-tetanus-acellular pertussis vaccine; HAV, hepatitis A vaccine; Hib, *Haemophilus influenza* type b vaccine; IIV, inactivated influenza vaccine; LAIV, live attenuated influenza vaccine; MCV4, 4-valent meningococcal conjugated vaccine; MMR, measles-mumps-rubella vaccine; MMRV, measles-mumps-rubella-varicella vaccine; mRNA 1273, mRNA vaccine against COVID-19; PCV13, pneumococcal conjugated 13-valent vaccine; PPSV23, 23-valent pneumococcal polysaccharide vaccine; TBE, tick-borne encephalitis vaccine.

TABLE 5 Population-specific considerations for vaccination[41–44]

Population	Considerations
Healthy	(A) Follow routine guidelines/schedule for vaccination. (B) Monitor mRNA vaccine recipients for 15–30 minutes per local guidelines
History of food allergies	*Egg*: Proceed with vaccination under supervision. *Yeast*: Seek allergist evaluation prior to Hepatitis A, B, HPV, DTaP, Meningococcal, Pneumococcal vaccines. *Gelatin*: Seek allergist evaluation prior to MMR, Zoster, Influenza, Rabies, Yellow fever, Typhoid vaccines.
History of immunosuppression	(A) Defer live vaccination. (B) Administer vaccine prior to immunosuppression if possible.
History of vaccine, drug, or antibiotic allergy	(A) Refer to allergist. (B) Identify specific components that may be similar in other vaccines. (C) Graded administration of vaccine/drug after discussion of risks and benefits

Yet, the CDC also recommends that individuals with a history of immediate allergic reactions to other vaccines weigh their risk of exposure, risk of severe disease or death due to COVID-19, and consider whether they were previously infected with COVID-19 (because of lower rates of reinfection in the three-month period after infection), when deciding whether to delay or forego vaccination.[50,51]

After the mRNA BNT162b2 vaccine received central European marketing approval on 21 December 2020, the Paul-Ehrlich-Institut, Federal Institute for Vaccines and Biomedicines in Germany, and the European Medicines Agency (EMA) released its own guidance for vaccination of people with allergies. These guidelines recommended that only individuals in the European Union with an allergy to a specific vaccine component not receive the vaccine.[52] The European Academy of Allergy and Clinical Immunology (EAACI) position paper on diagnosis, management, and prevention of severe allergic reactions to the COVID-19 vaccines states that the vaccines are contraindicated only when there is an allergy to one of the vaccine components or if there was a severe allergic reaction to the first dose. The paper also provides a simplified algorithm of prevention, diagnosis and treatment of severe allergic reactions and a list of recommended medications and equipment for vaccine centers.[53]

5 | SECTION IV: GENERAL MECHANISMS IN PATHWAYS OF IMMUNE-MEDIATED AND NON-IMMUNE-MEDIATED REACTIONS

Immunological hypersensitivity is either IgE-mediated (hypersensitivity reaction type 1),[54] IgG-mediated (hypersensitivity reaction type 2), immune complex and/or complement-mediated (hypersensitivity reaction type 3), or cell-mediated (hypersensitivity reaction type 4) (Figure 2).[55] The mechanisms underlying IgE-mediated hypersensitivity are best understood. Individuals with IgE-mediated allergic reactions first undergo an initial sensitization phase during which allergen-specific IgE antibodies bind to high-affinity FcεRI receptors on mast cells and basophils. Subsequent allergen exposure can cross-link the cell-bound IgE antibodies and triggers degranulation of mast cells and/or basophils, with subsequent release of histamine and other inflammatory chemical mediators (cytokines, interleukins, leukotrienes, and prostaglandins) into the surrounding tissue causing several systemic effects, such as vasodilation, mucous secretion, tissue eosinophilic infiltration, and airway smooth muscle contraction.[56]

Type II IgG-mediated immune vaccine reactions are rare but have been observed with an MMR vaccine containing dextran before those preparations were taken off the market.[57] The occurrence of large, local injection-site reactions with TdaP vaccines has been reported to be due to a local immune complex type III hypersensitivity reaction.[58] Type IV hypersensitivity reactions occur when an individual's T cells provoke an inflammatory response against allergens, leading to T-cell activation and the release of cytokines and chemokines.[59] Type IV reactions to vaccines induce local eczema, which may start between 2 hours to up to 2 days after vaccinations. These reactions are typically observed following vaccines containing aluminum and antimicrobial agents. The occurrence of such an event is not a contraindication for further vaccinations.[60]

Non-immunologic anaphylaxis is caused by agents or events that induce sudden, massive mast cell and/or basophil degranulation in the absence of immunoglobulins. These reactions may be due to activation of complement by nanoparticles, colloidal solutions, or liposomes without immune complex formation, commonly termed complement activation-related pseudoallergy or CARPAs[61] (eg, medications containing Cremophor EL[62]), direct mast cell and basophil activation resulting in histamine release (vancomycin[63] and opiate medications[61]), or other mechanisms (activation of the kallikrein-kinin pathway[64]). Nonsteroidal anti-inflammatory drugs (NSAIDs),[65] local anesthetics,[66] monoclonal antibodies, and chemotherapeutic agents have also been reported to induce non-immunologic anaphylaxis. Recently, transient receptor potential cation receptor, subfamily V (TRPV4) channel has been implicated as a driver of IgE-independent mast cell-dependent bronchospasm via cysteinyl leukotrienes release.[67] In addition, MRGPRX2, a Mas-related G protein-coupled receptor, has been identified as a mechanism for mast cell degranulation. Since 2015, evidence has accumulated that

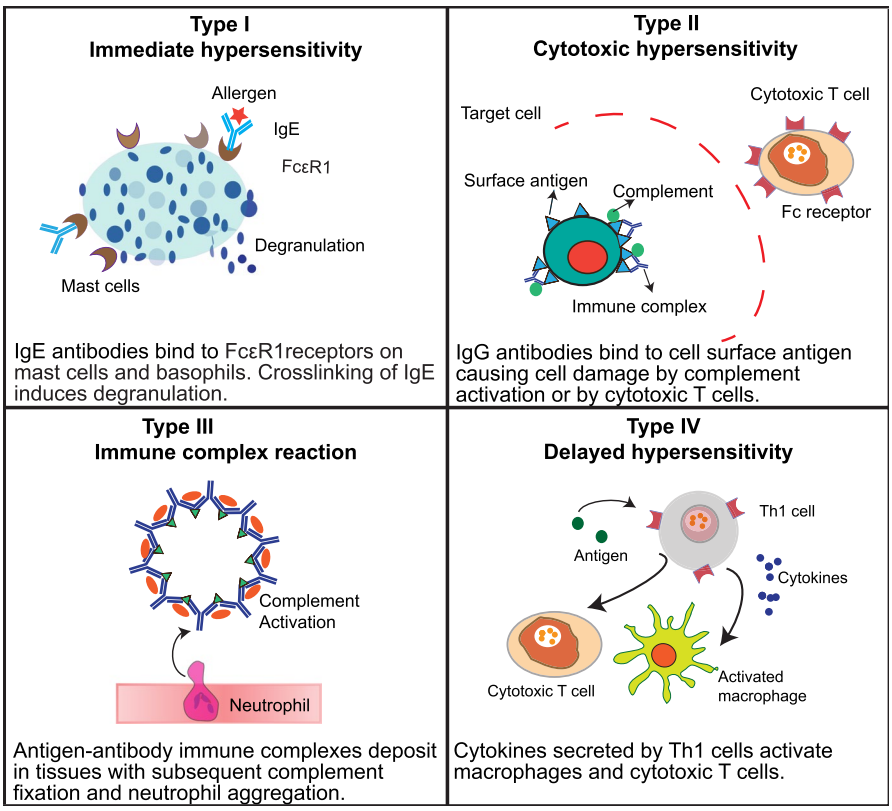

FIGURE 2 Types of hypersensitivity reactions

off-target occupation of this receptor by different therapy regimens, such as neuromuscular blocking agents (NMBA) and opioids could constitute an additional mechanism of non-immune immediate drug hypersensitivity. [68–70] While radio-contrast agents have traditionally been considered to be non-immunologic, some of the newer, low-osmolar agents may induce IgE-mediated reactions.[21]

Although the clinical presentations of both IgE-mediated anaphylaxis and non-immunologic anaphylaxis are similar, measurements of tryptase and SC5b-9may assist in differentiating the two types.[71,72] Tryptase is a marker of mast cell activation which is released following mast cell degranulation while SC5b-9 is a marker of complement activation and is a terminal complement complex.[71,73] As both tryptase and SC5b-9 are transiently elevated soon after an anaphylaxis episode, blood should ideally be collected between 30 and 90 min after the onset of reaction.[71] Acute serum total tryptase should be at least 20% plus 2 ng/mL over the baseline tryptase level.[73] Another novel emerging biomarker is hereditary α-tryptasemia which is present in mastocytosis and may be useful for determining the individual patient's risk of developing severe anaphylaxis.[74] Figure 3 depicts the mechanisms of IgE-mediated and non-immunological anaphylaxis.[75–81]

6 | SECTION V: PROVEN AND SUSPECTED ALLERGENIC COMPONENTS OF VACCINES

Anaphylaxis to vaccines is rare and occurs primarily among individuals who have histories of allergies to the components of the vaccines.[19] Allergic reactions after vaccination can be due to any of the vaccine components such as antigens, adjuvants, stabilizers, preservatives, emulsifiers, leached packaging components, residual antibiotics, cell culture materials, and inactivating ingredients (Box 1).[82] Table 6 lists components that have been implicated in allergenic reactions and related adverse events. Here, we discuss some of the most common allergenic or potentially allergenic components of vaccines.

Many vaccines contain small amounts of the egg protein ovalbumin. Influenza, yellow fever, and rabies vaccines tend to have higher concentrations of ovalbumin because they are cultured in embryonated chicken eggs.[85] Vaccines cultured in chicken embryo fibroblasts, such as the MMR vaccine, have lower concentrations of egg protein than those cultured in embryonic eggs.[86] While egg allergy is common in childhood, studies have shown that vaccinating egg-allergic children with MMR and influenza vaccines is well tolerated and risk of an allergic reaction is similar in the general population.[87,88] Specifically, egg-allergic children, including those who have had anaphylaxis, were successfully vaccinated with yellow fever[89] vaccines with no serious adverse events reported. Since severe allergic reactions to egg-based influenza vaccines are rare, the CDC and its Advisory Committee on Immunization Practices (ACIP) guidelines state that individuals with mild egg allergy can receive any licensed and recommended age-appropriate flu vaccine and no longer need to be observed for 30 minutes after receiving the vaccine. However, in those with severe egg allergy, the vaccines should only be given under the supervision of a health care provider who is capable of recognizing and managing serious allergic conditions.[90,91]

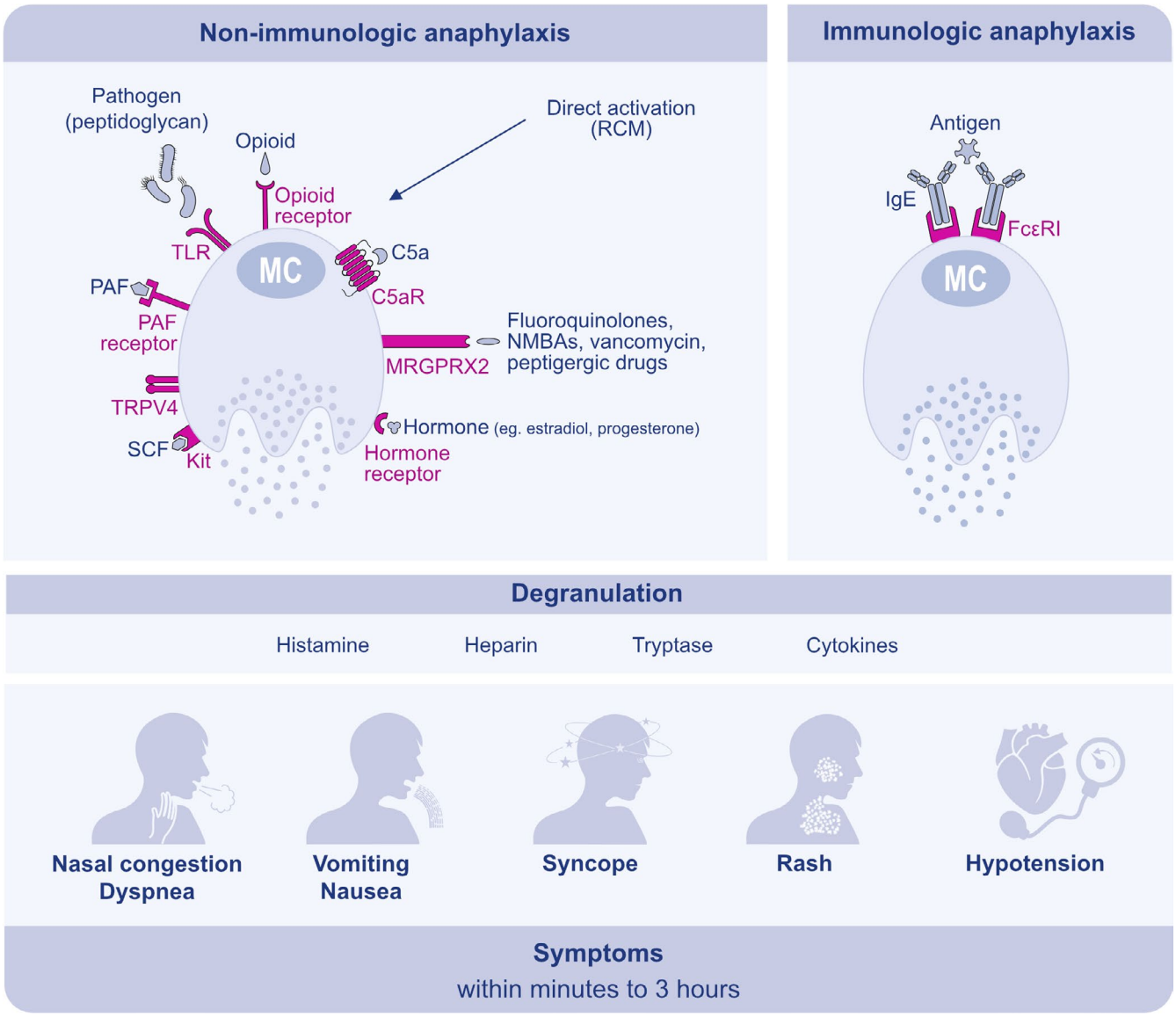

FIGURE 3 Immunological (IgE-mediated) and non-immunological anaphylaxis

Gelatin, a protein derived from bovine or porcine sources, is added to both live and inactivated vaccines as a stabilizing agent.[60] Sensitivity to gelatin was confirmed with both skin-prick tests and by immunoassay in a 17-year-old female who had an anaphylactic reaction to an MMR vaccine.[92] A retrospective case-control study which interviewed and collected sera from individuals who had suffered anaphylaxis after receiving MMR found that 27% of them had anti-gelatin IgE. In comparison, anti-gelatin IgE was not detected in any of the vaccinated subjects who did not present with adverse events.[93] It was subsequently shown that patients who have anaphylaxis to MMR were sensitized to gelatin present in the DTaP vaccine,[94,95] and that cellular immunity to gelatin from the DTaP vaccine can persist for more than three years.[96] However, sensitization may also persist due to exposure to gelatin in foods or through cross-reactivity to other allergens such as egg, chicken, and cow's milk.[97] Gelatin is also a source of alpha-gal, an carbohydrate allergen that causes meat allergy.[98] Anaphylaxis was observed after vaccination with MMR, Varicella, and DTaP/IPV in pediatric subjects with alpha-gal allergy.[99] Removal of gelatin from vaccines has dramatically reduced allergic reactions to these vaccines.[100]

Milk proteins are used as growth media in DTaP and Tdap vaccines. Although bovine casein is present in nanogram quantities in these vaccines, they rarely cause anaphylaxis. Kattan et al reported eight children with severe cow's milk allergy who reacted with anaphylaxis to the booster dose of DTaP or Tdap vaccine and suggested casein present in the vaccines may play role in the induction of anaphylaxis in atopic children.[101,102] However, the methods used in this report were questioned[102] and to our knowledge, there have been no subsequent data that support a causative role for DTaP or Tdap vaccines in the induction of allergic disease. It is the position of EAACI (or subcommittee) that these vaccines do not contribute to the pathogenesis of allergic disease and that atopy is not a contraindication to these vaccines.[32]

TABLE 6 Major components, function, and allergic reactions[a]

Component	Function	Vaccine	Allergic reactions
Gelatin	Stabilizer	MMR, Zoster, Influenza Rabies, Yellow fever, Typoid	Anaphylaxis, Urticaria, Local
Albumin (eggprotein, bovine/ calf/fetal serum)	Residual médium, Stabilizer	Yellow fever, MMR, Influenza, Rabies, Influenza	Anaphylaxis
Casein	Medium nutrient	DTaP, Meningococcal, Hib, Tdap, Influenza	Anaphylaxis
Aluminum	Adjuvant	Adenovirus Antrax, DTaP, Hib, Hepatitis A/B, HPV, Japanese EncephalitisMeningococcalPheumococcal, Tdap	Local
2-Phenoxyethanol	Stabilizer, Preservative	DTaP, Influenza, Polio, Tdap	Local
Thimerosal	Preservative	Influenza, Td	Local
Yeast	Medium nutrient	Hepatitis A, B, HPV, DTaP, Meningococcal, Pneumococcal, Typhoid	Anaphylaxis
Natural latex	Pharmaceuticalclosure	Tdap, Meningococcal, Anthrax, Hepatitis A, B, Influenza, DTaP, Rotavirus, Td, Yellowfever	Anaphylaxis, Urticaria
Neomycin	Antimicrobial	Polio, DTaP, Hepatitis A, Influenza, MMR, Rabies, Polio, Smallpox, Varicella, Zoster	Anaphylaxis
Polymyxin B	Antimicrobial	DTaP, Influenza, Polio, Smallpox	NA
Streptomycin	Antimicrobial	DTaP, Polio	NA
Kanamycin	Antimicrobial	Meningococcal, Influenza	Anaphylaxis
Gentamicin	Antimicrobial	Influenza	NA
Amphotericin B	Antimicrobial	Rabies	NA
Dextran	Stabilizer Medium nutrient	MMR[b], Rotavirus	Anaphylaxis
Formaldehyde	Inactivation of virus, Detoxification of bacterial toxin (Inactivating agent)	Polio, DTaP, Hib, Hepatitis B, Influenza, Japanese Encephalitis, Meningococcal, Tdap, Thypoid	Local
Peptone (soy)	Medium nutrient	Pneumococcal Hepatitis B	
Polysorbate 80	Surfactant	DTaP, Hepatitis A, B, HPV, Influenza, Meningococcal, Pneumococcal, Rotavirus, Tdap, Zoster	Non-immunological anaphylaxis, Local
Polyethylene glycol	Surfactant of mRNA	COVID-19	Anaphylaxis

[a]All reasonable efforts have been made to ensure the accuracy of this information, but manufacturers can change product contents before that information is reflected here. Some major components of vaccines have higher levels in food.

[b]Currently, MMR vaccines containing Dextran not on the market.

BOX 1 Description of common components and contaminants present in vaccine formulations[43,82-84]

Vaccine Component Category	Function
Antigen or its genetic code (DNA, RNA)	Molecules of the pathogen that cause the formation of antibodies and development of specific immune protection (humoral, cellular)
Adjuvants	To stimulate, broaden and optimize immune response
Stabilizers	To keep the vaccine potent and safe during storage and transportation
Preservatives	To prevent contamination
Residual antibiotics	To prevent contamination by bacteria during the vaccine manufacturing process
Residual cell culture materials	To grow enough of the virus or bacteria to make the vaccine
Residual inactivating ingredients	To kill viruses or inactivate toxins during the manufacturing process
Latex	Found in the vial and syringes used to contain and administer the vaccine. It is a potential contaminant.

As stated above, dextran present in one preparation of MMR vaccine was responsible for numerous cases of anaphylaxis, but this brand of MMR vaccine has since been withdrawn from the market.[103] It was used as a medium nutrient or as a stabilizer. Similarly, during Brazil's national MMR vaccination campaign in 2004, the rate of hypersensitivity following MMR vaccination was unexpectedly high while its case-control study showed no association with a history of allergy. However, subsequent studies implicated dextran as the likely cause of these hypersensitivity events.[104]

Many vaccines contain antigens that are created in cell lines. For example, hepatitis B vaccines and the human papillomavirus (HPV) vaccines contain antigens that are recombinant proteins expressed in Baker's yeast.[105] Purification removes most of the cellular material, but it is impossible to remove all trace components. Between 1990 and 2004, only 15 reports were identified of probable or possible anaphylaxis following vaccination of individuals with a reported history of yeast allergy. Eleven of these occurred after administration of the hepatitis B vaccine, which contains trace amounts of yeast proteins. Because these subjects were not tested for yeast allergy, it cannot be confirmed that sensitivity to yeast caused these adverse reactions. These data indicate that recombinant yeast-derived hepatitis B vaccine poses minimal risk of allergic reactions in yeast-sensitive individuals. Therefore, evaluation by an allergist is recommended for people who have a history of severe yeast allergy before administration of hepatitis B and HPV vaccines.[105] According to VAERS, there were 107 reports of adverse events in those with a history of yeast allergy present prior to vaccination; of these 11 recipients of hepatitis B vaccine had probable or possible anaphylaxis events.[105] By contrast, another study found no episodes of anaphylaxis in a large cohort of women who had positive skin tests to yeast extract after the HPV vaccine.[106]

Antibiotics such as neomycin, streptomycin, polymyxin B, kanamycin, and gentamicin are well known to cause mild to life-threatening allergic reactions. An individual receiving an MMR vaccine containing neomycin was reported to have experienced anaphylaxis shortly after vaccination.[107] Although a skin test to neomycin alone could not be performed in this individual due to a lack of a commercial preparation suitable for skin testing, patient history indicated systemic sensitivity on topical application of neomycin during infancy to disrupted skin. In another case, a history of previous reaction and positive skin test to neomycin was not associated with immediate or delayed hypersensitivity reactions following MMR vaccination.[108] In a report of anaphylaxis after rabies vaccination, the presence of residual kanamycin in the vaccine and a positive kanamycin result to an antibiotic skin sensitivity test suggested that kanamycin was the likely cause of the adverse event.[109] Finally, there is one report of anaphylaxis after applying eye drops containing polymyxin B, an excipient used in DTaP and other vaccines.[43,110] To our knowledge, no other antibiotics have been associated with vaccine-associated anaphylaxis. 2-Phenoxyethanol is widely used as preservative in cosmetics and vaccines due to its large spectrum of antimicrobial activity and is considered as one of the most well-tolerated preservatives.[111]

Natural latex allergy is well characterized among healthcare personnel and latex content in vaccines as vial stopper or syringe plunger may pose safety concerns in this population. However, Smith et al could not detect latex allergens in adult vaccines.[112] In 2004, an analysis of VAERS revealed only 28 cases of immediate hypersensitivity with latex allergy in vaccine recipients among 160,000 reports of vaccine-associated adverse events.[113]

Thimerosal, which is approximately 50% mercury by weight, has been one of the most widely used preservatives in vaccines to prevent growth of harmful microbes. All vaccines routinely recommended for children 6 years of age and younger in the United States are available in formulations that do not contain thimerosal.[114] A risk assessment study revealed that except for local hypersensitivity reactions, there is no evidence of harm caused by thimerosal in vaccines.[115] Thus, while thimerosal is the most prevalent preservative inducing contact dermatitis, it is considered irrelevant to vaccine-induced anaphylaxis.[116]

Formaldehyde and beta-propiolactone (BPL) have been used to inactivate viruses during the production of vaccines. However, approximately 6% of individuals who receive a booster dose of the rabies human diploid cell vaccine (HDCV) develop an immune complex-like reaction in the 2–21 days that follow.[117,118] These reactions have been associated with the presence of BPL altered human albumin contained in the HDCV.[119,120] Currently, SinoPharm' BBIBP-CorV[121] and Sinovac Life Sciences's CoronaVac's[122] COVID-19 vaccine are using BPL to inactivate SARS-CoV-2. Both vaccines are approved for use in China. Although formaldehyde is only found in residual quantities in vaccine preparation, it has been reported to aggravate eczematous dermatitis following hepatitis B vaccination.[123] It is used to inactivate virus or for the detoxification of bacterial toxin. Formaldehyde-specific contact dermatitis had also been reported following formaldehyde-containing influenza vaccine.[124] It is hypothesized that the introduction of carbonyl groups on antigens by formaldehyde in vaccines profoundly affects its immunogenicity, thus explaining adverse effects due to formaldehyde-containing vaccines.[125,126]

Adjuvants are incorporated into some vaccines to boost T-cell immunity and increase helper T-cell function. The most commonly used adjuvants in vaccines are aluminum hydroxide and aluminum phosphate. Despite its long-standing use as an adjuvant in vaccines, aluminum has always been the target of controversy. Although no association between direct toxicity of aluminum and vaccines has been established, several delayed-type hypersensitivity reactions have been reported.[127–129] In Denmark, 39 out of 42 children with persistent skin reactions following vaccination had positive patch tests for aluminum.[130] In another study, vaccination-induced granulomas and contact allergy to aluminum was reported in 60 out of 63 Swedish children receiving DTP vaccines.[131] In contrast, an in vivo study in a mouse model of peanut allergy found that the severity of peanut-hypersensitivity was reduced by an alum/CpG-adjuvanted vaccine while exposure to endotoxin and alum did not influence allergic symptoms.[132] Other novel adjuvants such as MF59, AS03, and AF03, which are squalene-based are also used in vaccines.[133]

TABLE 7 Excipients in the COVID-19 vaccines mRNA BNT162b2, mRNA-1273, and ChAdOx1-Sand their functions

Type of Excipient	Excipient	Vaccine
Lipid nanoparticles: stability and transport of mRNA		
PEGylated lipids	((4-hydroxybutyl)azanediyl)bis(hexane-6,1-diyl)bis(2-hexyldecanoate) (ALC-0159)	mRNA BNT162b2
	1,2-dimyristoyl-rac-glycero-3-methoxypolyethylene glycol-2000 (PEG2000 DMG)	mRNA-1273
Phospholipids	1,2-distearoyl-sn-glycero-3-phosphocholine (DSPC)	mRNA BNT162b2, mRNA-1273
Lipids	((4-hydroxybutyl)azanediyl)bis(hexane-6,1-diyl)bis(2-hexyldecanoate) (ALC-0315)	mRNA BNT162b2
	Lipid SM-102 (patented ionizable lipid)	mRNA-1273
	Cholesterol	mRNA BNT162b2, mRNA-1273
Buffer: stability of lipid nanoparticles/radical oxidation inhibition		
Phosphate buffer	Potassium dihydrogen phosphate/disodium phosphate dihydrate)	mRNA BNT162b2
Tromethamol	Tromethamol/tromethamol hydrochloride	mRNA-1273
Acetic acid/acetate	Acetic acid/sodium acetate trihydrate	mRNA-1273
Histidine	L-histidine/L-histidine hydrochloride monohydrate	ChAdOx1-S
Other stabilizers: Ionic strength, surfactant, metal-ion chelates		
	Potassium chloride	mRNA BNT162b2
	Sodium chloride	mRNA BNT162b2, ChAdOx1-S
	Magnesium chloride hexahydrate	ChAdOx1-S
	Ethanol	ChAdOx1-S
	Disodium edetate dihydrate (EDTA)	ChAdOx1-S
	Polysorbate 802	ChAdOx1-S
Thermostabilization		
	Sucrose	mRNA BNT162b2, mRNA-1273, ChAdOx1-S

Note: Adapted from: Borgsteede S, Tjerk G, Tempels-Pavlica Z, Other excipients than PEG might cause serious hypersensitivity reactions in COVID-19 vaccines. Allergy. Accepted January 2021. In Press (2021).

Although safety concerns were raised due to presence of antibodies to squalene, clinical evidence clearly suggested that squalene is poorly immunogenic and that low titers of antibodies to squalene are found in healthy individuals. Further, neither the presence of anti-squalene antibodies nor their titer is significantly increased by immunization with vaccines containing squalene.[134] Details on the mechanism of these and other vaccine adjuvants under clinical investigation are detailed in the review by Shi et al.[135]

Polysorbate 80 is an emulsifier which has been widely used to solubilize agents in foods and medicines, including vaccines. This nonionic detergent induces local and systemic allergic reactions, including IgE-mediated and non-immune anaphylaxis. The hydrophilic polymer polyethylene glycol (PEG) is structurally similar to polysorbate 80 and PEG, and its derivatives are frequently found in household products including toothpaste, cosmetics, pharmaceuticals, and foods. In addition, PEG is often conjugated to biological therapeutics to form a depot agent. It is now well understood that sensitivity to PEG can cause IgE-mediated anaphylaxis after administration of PEG-conjugated biological therapeutics,[136–141] and that severe allergic reactions to PEG have been associated with pre-existing anti-PEG antibodies induced by PEG-containing household products.[142] Polysorbate 80 is present in many vaccines and some foods as well. Polysorbates are obtained from PEG moieties but have lower molecular weights and thus may be much less likely to trigger an allergic reaction. PEG may also be cross-reactive with polysorbates, which are contained in some COVID-19 vaccines.[141,143–145] However, measures of pre-existing anti-PEG antibodies vary widely, range from 0.2% to 72% of healthy individuals.[146] This has become immediately relevant because PEG 2000, a high-molecular weight version of PEG, is a component in two of the three authorized COVID-19 vaccines. The mRNA BNT162b2 and mRNA-1273 COVID-19 vaccines are lipid nanoparticles containing mRNA that codes for the spike protein in the coronavirus.[147] The lipid nanoparticle delivery system prevents premature degradation of the genetic instructions necessary for individuals to eventually become protected against SARS-CoV-2.[148] PEG 2000-lipid is a component of the mRNA-1273 vaccine. The lipid

BOX 2 Future safety research objectives

General	COVID–19
Develop individualized prediction algorithms to determine risk of a vaccine reaction. Refine reaction reporting guidelines for continued accurate vaccine pharmacovigilance. Identify vaccine components that mediate allergic reactions. Understand mechanisms for reactions to various vaccine components. Develop better adjuvants	Compare various COVID–19 vaccines to each other to understand key differences in safety and efficacy, stratified by patient age and health status. Compare intradermal PEG tests to oral challenges in evaluating PEG sensitization.

nanoparticles stabilize and improve the aqueous solubility of the two mRNA vaccines, and also act as an adjuvant. While more research is needed to determine the cause of the potentially increased rate of anaphylaxis to COVID-19 compared to other vaccines, based on the experience with PEG-conjugated biologics, PEG 2000 in the vaccines is considered the most likely culprit.[138,139,149–153] Therefore, individuals with a known allergy to PEG should be excluded from vaccination with these vaccines for the time being.[154] In addition to PEG, other excipients present in COVID-19 vaccines, such as distearoyl phosphatidylcholine, tromethamol, polysorbate 80, and EDTA, should also be evaluated as potential allergenic components Table 7 lists the excipients present in the mRNA and ChAdOx1-S vaccines.[155]

7 | SECTION VI: MANAGEMENT OF VACCINE ALLERGY

Treatment of anaphylaxis in the setting of vaccine administration is reviewed in Sokolowska et al[156] and Castells et al.[137] Briefly, due to the possibility of an anaphylactic reaction to the vaccine, any professional administering the vaccine must be capable of managing an anaphylactic reaction and should have the necessary medications and tools on hand. There must be a mandatory observation period after vaccine administration of at least 15 minutes for all individuals, to allow for the administration of adrenaline in an adequate dose.[157] Individuals with a suspected allergic reaction to the first dose of the vaccine should be followed up by an allergist so that administration of the second dose can be performed in a specialized setting equipped to treat anaphylaxis. One approach used successfully for many vaccine-allergic individuals, but which has not been evaluated for the COVID-19 vaccines, is to administer the vaccine in incremental doses. Any adverse allergic reactions should be promptly reported including any additional information including individual characteristics.

8 | SECTION VII: CONCLUSIONS

Vaccinations benefit public health and help to reduce the risk of disease across the entire population. Despite early reports of waning of antibody responses over 20 days following SARS-CoV2 infection, evidence is now accumulating that similarly to other infections, recovered patients develop long-lasting immunity.[158] Recently, Hartley et al. reported that patients who have recovered from SARS-CoV-2 infection have stable virus-specific memory B cells that recognize the spike or nucleocapsid proteins of the SARS-CoV-2 virus for at least eight months post infection.[159] Another study found that neutralizing antibody titers against the SARS-CoV-2 spike protein persisted for at least 5 months after infection.[160] These findings support optimism that the currently licensed vaccines will be efficacious despite the emergence of highly infectious mutant viruses and are consistent with the limited reports of natural reinfection after confirmed illnesses. These mRNA vaccines are currently not available for children. Moreover, research by Pfizer and researchers from the University of Texas have determined that antibodies from 20 recipients of the mRNA BNT162b2vaccine can neutralize the mutant strains in vitro (as yet not peer reviewed).[161]

It is crucial that further research is conducted to better understand to which components of the currently available vaccines individuals are reacting and how to identify individuals who may be at risk of an adverse allergic reaction (Box 2). Individuals who may be at risk for an allergic response or who have a history of allergic responses to vaccinations (or their components) should be evaluated by an allergist.[60]

The best mechanism to further our understanding is to study individuals who have had reactions through a variety of in vitro experiments and clinical testing. In vitro experiments including analysis of plasma IgE markers as well as, basophil and mast cell line activation tests can help to better characterize potential allergens and identify individuals at risk of an anaphylaxis. Clinical testing, including skin-prick and intradermal tests, can also help identify allergens and at-risk individuals. All of these tests can involve testing the individual components of the vaccine to determine the reaction-inducing antigen. However, the critical and pragmatic evaluation of these tests' results in relationship to particular patient's clinical problems remains crucial.

Increased understanding of vaccine-related allergies will help to further improve the manufacturing processes and safety of vaccines. Specifically, through identifying specific vaccine components that cause allergic reactions, vaccine manufacturers can either try to remove or create replacements for those components. This also has an impact on the management of vaccine distribution, specifically

with regards to ensuring that those who need vaccination will not have an allergic reaction.

ACKNOWLEDGMENTS

We would like to thank the Parker foundation, the Sunshine foundation, the Crown Foundation, and the NIAID SARS Vaccine study for funding the study.

CONFLICT OF INTEREST

Dr. Diamant reports personal fees from ALK, AstraZeneca, Boehringer Ingelheim, GlaxoSmithKline, HAL Allergy, Merck Sharp & Dohme, and Sanofi-Genzyme-Regeneron and acted as Research Directorat QPS-NL, an institution which received research support from several bio-pharmaceutical companies, esp within respiratory: HAL Allergy, Foresee Pharmaceuticals, Patara Pharma (now Respivant), Novartis; Dr. Jesenak received honoraria, consultancy and speaker fees from Sanofi-Pasteur, Pfizer, GlaxoSmithKline, Merck Sharp&Dohme, Takeda, ALK, Stallergenes-Greer, Angelini, Novartis, Mundipharma and Berlin-Chemie and acted as principal investigator in trials sponsored by BioCryst, Pharming, Octapharma, and Baxalta; Dr. Vieths reports personal fees from Swiss Society for Allergy and Immunology, SchattauerAllergologieHandbuch, Elsevier Nahrungsmittelallergien und Intoleranzen, and Karger Food Allergy: Molecular Basis and Clinical Practice and also reports non-financial support from German Research Foundation, European Directorate for the Quality of Medicines and Health Care, European Academy of Allergy and Clinical Immunology, German Chemical Society (GDCh), AKM Allergiekongress, International Union of Immunological Societies, and from Spanish Society for Allergy and Clinical Immunology (SEAIC); Dr. Agache is an Associate Editor at Allergy and PAI; Dr. Chinthrajah reports grants from NIAID, CoFAR, Aimmune, DBV Technologies, Astellas, Regeneron, and FARE and is an advisory board member of Alladapt Therapeutics, Genentech, Novartis, and Sanofi; Dr. Eiwegger acts as local PI for company-sponsored trials by DBV and sub-investigator for Regeneron, holds grants from Innovation fund Denmark, CIHR. He is Co-Investigator or scientific lead in three investigator-initiated oral immunotherapy trials supported by the Food Allergy and Anaphylaxis Program SickKids and serves as associate editor for Allergy. He/his laboratory received unconditional/kind contributions from Macro Array Diagnostics and ALK. He holds advisory board roles for ALK; Dr. Barber reports grants from ALK, ALLERO therapeutics, personal fees from ALK and Aimmune; Dr. Ollert reports personal fees from Hycor Biomedical and is Scientific co-founder of TolerogenicsSarL; Dr. Palomares received research grants from Inmunotek S.L., Novartis and MINECO, received fees for giving scientific lectures or participation in Advisory Boards from Allergy Therapeutics, Amgen, AstraZeneca, Diater, GlaxoSmithKline, S.A, Inmunotek S.L, Novartis, Sanofi-Genzyme and Stallergenes; Dr. Pfaar reports grants and personal fees from ALK-Abelló, Allergopharma, Stallergenes Greer, HAL Allergy Holding B.V./HAL Allergie GmbH, BencardAllergie GmbH/Allergy Therapeutics, Lofarma, ASIT Biotech Tools S.A, Laboratorios LETI/LETI Pharma, Anergis S.A, and Glaxo Smith Kline, and grants from Biomay, Circassia, Pohl-Boskamp, and Inmunotek S.L, and personal fees from MEDA Pharma/MYLAN, Mobile Chamber Experts (a GA2LEN Partner), Indoor Biotechnologies, Astellas Pharma Global, EUFOREA, ROXALL Medizin, Novartis, Sanofi-Aventis and Sanofi-Genzyme, Med Update Europe GmbH, streamedup! GmbH, John Wiley and Sons, AS, and Paul-Martini-Stiftung (PMS); Dr. Sokolowska reports grants from Swiss National Science Foundation (SNF) and from GSK; Dr. Torres reports grants from European Comission, SEAIC, and ISCIII and personal fees from Diater laboratory, Leti laboratory, and consultancy from Aimmune Therapeutics; Dr. Traidl-Hoffmann reports personal fees from Sanofi Genzyme, Lilly Pharma, Novartis, Töpfer, and grants and personal fees from Sebapharma; Dr. van Zelm reports grants from NHMRC and has a patent pending, "Methods for detecting immune response"; Dr. Akdis reports grants from Allergopharma, Idorsia, Swiss National Science Foundation, Christine Kühne-Center for Allergy Research and Education, European Commission's Horizon's 2020 Framework Programme, Cure, Novartis Research Institutes, Astra Zeneca, Scibase, advisory board from Sanofi/Regeneron, Glaxo Smith-Kline, Scibase, and Novartis; Dr. Nadeau reports grants from National Institute of Allergy and Infectious Diseases (NIAID), National Heart, Lung, and Blood Institute (NHLBI), and National Institute of Environmental Health Sciences (NIEHS); Food Allergy Research & Education (FARE), Director of World Allergy Organization (WAO) Center of Excellence at Stanford; Advisor at Cour Pharma; Co-founder of Before Brands, Alladapt, Latitude, and IgGenix; National Scientific Committee member at Immune Tolerance Network (ITN) and National Institutes of Health (NIH) clinical research centers; DSMB member for NHLBI, US patents for basophil testing, multifood immunotherapy and prevention, monoclonal antibody from plasmablasts, and device for diagnostics. All other authors report no COI.

ORCID

Zuzana Diamant https://orcid.org/0000-0003-0133-0100
Milos Jesenak https://orcid.org/0000-0001-7976-2523
Ioana Agache https://orcid.org/0000-0001-7994-364X
Mübeccel Akdis https://orcid.org/0000-0003-0554-9943
Domingo Barber https://orcid.org/0000-0002-5488-5700
Heimo Breiteneder https://orcid.org/0000-0003-2022-8689
Sharon Chinthrajah https://orcid.org/0000-0003-2467-4256
Thomas Eiwegger https://orcid.org/0000-0002-2914-7829
Wytske Fokkens https://orcid.org/0000-0003-4852-229X
Robyn E. O'Hehir https://orcid.org/0000-0002-3489-7595
Liam O'Mahony https://orcid.org/0000-0003-4705-3583
Oscar Palomares https://orcid.org/0000-0003-4516-0369
Oliver Pfaar https://orcid.org/0000-0003-4374-9639
Carmen Riggioni https://orcid.org/0000-0002-8745-0228
Mohamed H Shamji https://orcid.org/0000-0003-3425-3463
Maria Jose Torres https://orcid.org/0000-0001-5228-471X
Claudia Traidl-Hoffmann https://orcid.org/0000-0001-5085-5179
Menno van Zelm https://orcid.org/0000-0003-4161-1919
De Yun Wang https://orcid.org/0000-0002-0909-2963
Luo Zhang https://orcid.org/0000-0002-0910-9884
Cezmi A. Akdis https://orcid.org/0000-0001-8020-019X
Kari C. Nadeau https://orcid.org/0000-0002-2146-2955

REFERENCES

1. Pfizer and BioNTech achieve first authorization in the world for a vaccine to combat COVID-19; 2020. https://www.pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-achieve-first-authorization-world. Accessed January 14, 2021.
2. Pfizer. Pfizer and BioNTech celebrate historic first authorization in the U.S. of vaccine to prevent COVID-19; 2020. https://www.pfizer.com/news/press-release/press-release-detail/pfizer-and-biontech-celebrate-historic-first-authorization. Accessed January 13, 2021.
3. U.S. FDA Takes Additional Action in Fight Against COVID-19 By Issuing Emergency Use Authorization for Second COVID-19 Vaccine; 2020. https://www.fda.gov/news-events/press-announcements/fda-takes-additional-action-fight-against-covid-19-issuing-emergency-use-authorization-second-covid. Accessed January 14, 2021.
4. Moderna. Press Release; 2021. https://investors.modernatx.com/news-releases. Accessed January 22, 2021.
5. AstraZeneca's COVID-19 vaccine authorised for emergency supply in the UK; 2020. https://www.astrazeneca.com/media-centre/press-releases/2020/astrazenecas-covid-19-vaccine-authorised-in-uk.html. Accessed January 14, 2021.
6. van Riel D, de Wit E. Next-generation vaccine platforms for COVID-19. *Nat Mater*. 2020;19(8):810-812.
7. World Health Organization. Draft landscape of COVID-19 candidate vaccines; 2021. https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines. Accessed January 14, 2021.
8. Karamloo F, Konig R. SARS-CoV-2 immunogenicity at the crossroads. *Allergy*. 2020;75(7):1822-1824.
9. Klimek L, Jutel M, Akdis CA, et al. ARIA-EAACI statement on severe allergic reactions to COVID-19 vaccines - an EAACI-ARIA position paper. *Allergy*. 2020;76(6):1624-1628.
10. Azkur AK, Akdis M, Azkur D, et al. Immune response to SARS-CoV-2 and mechanisms of immunopathological changes in COVID-19. *Allergy*. 2020;75(7):1564-1581.
11. FDA US. CFR - Code of Federal Regulations Title 21; 2010. https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/cfrsearch.cfm?fr=312.32. Accessed January 22, 2021.
12. Morris L, Swofford S. Vaccine Safety. *Prim Care*. 2020;47(3):431-441.
13. Siegrist CA. Mechanisms underlying adverse reactions to vaccines. *J Comp Pathol*. 2007;137(Suppl 1):S46-50.
14. Spencer JP, Trondsen Pawlowski RH, Thomas S. Vaccine adverse events: separating myth from reality. *Am Fam Physician*. 2017;95(12):786-794.
15. Kelso JM, Greenhawt MJ, Li JT, et al. Adverse reactions to vaccines practice parameter 2012 update. *J Allergy Clin Immunol*. 2012;130(1):25-43.
16. Views TD-CsHDNa. 653 Deaths +12,044 Other Injuries Reported Following COVID Vaccine, Latest CDC Data Show; 2021. https://childrenshealthdefense.org/defender/vaers-injuries-covid-vaccine-cdc-data/. Accessed March 8, 2021.
17. U.S. Department for Health and Human Services. Common Terminology Criteria for Adverse Events (CTCAE). Version 5.0; 2017. https://ctep.cancer.gov/protocoldevelopment/electronic_applications/docs/CTCAE_v5_Quick_Reference_5x7. pdf. Accessed May 28, 2021.
18. Sampson HA, Munoz-Furlong A, Campbell RL, et al. Second symposium on the definition and management of anaphylaxis: summary report–Second National Institute of Allergy and Infectious Disease/Food Allergy and Anaphylaxis Network symposium. *J Allergy Clin Immunol*. 2006;117(2):391-397.
19. McNeil MM, DeStefano F. Vaccine-associated hypersensitivity. *J Allergy Clin Immunol*. 2018;141(2):463-472.
20. Radice A, Carli G, Macchia D, Farsi A. Allergic reactions after vaccination: translating guidelines into clinical practice. *Eur Ann Allergy Clin Immunol*. 2019;51(2):51-61.
21. Watts MM, Marie DA. Anaphylaxis. *Allergy Asthma Proc*. 2019;40(6):453-456.
22. Bilo MB, Martini M, Tontini C, Corsi A, Antonicelli L. Anaphylaxis. *Eur Ann Allergy Clin Immunol*. 2021;53(1):4-17.
23. Jensen F, Woudwyk M, Teles A, et al. Estradiol and progesterone regulate the migration of mast cells from the periphery to the uterus and induce their maturation and degranulation. *PLoS One* 2010;5(12):e14409.
24. Johansson SG, Bieber T, Dahl R, et al. Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol*. 2004;113(5):832-836.
25. National Advisory Committee on Immunization. Canadian Immunization Guide. Anaphylaxis and other acute reactions following vaccination 2020. https://www.canada.ca/en/public-health/services/publications/healthy-living/canadian-immunization-guide-part-2-vaccine-safety/page-4-early-vaccine-reactions-including-anaphylaxis.html. Accessed January 23, 2021.
26. Department for Health and Wellbeing, Government of South Australia. Vasovagal Episode or Anaphylaxis; 2019. https://www.sahealth.sa.gov.au/wps/wcm/connect/7bf436004f0c7c268dc7af791a12b24c/Vasovagal+episode+or+anaphylaxis+Chart+-+201907+%28v1.0%29.pdf?MOD=AJPERES&CACHEID=ROOTWORKSPACE-7bf436004f0c7c268dc7af791a12b24c-niQpYNW. Accessed January 14, 2021.
27. Kochhar S, Salmon DA. Planning for COVID-19 vaccines safety surveillance. *Vaccine*. 2020;38(40):6194-6198.
28. Ruggeberg JU, Gold MS, Bayas JM, et al. Anaphylaxis: case definition and guidelines for data collection, analysis, and presentation of immunization safety data. *Vaccine*. 2007;25(31):5675-5684.
29. Wittes J, Crowe B, Chuang-Stein C, et al. The FDA's Final Rule on Expedited Safety Reporting: Statistical Considerations. *Stat Biopharm Res*. 2015;7(3):174-190.
30. Minor PD. Live attenuated vaccines: Historical successes and current challenges. *Virology*. 2015;480:379-392.
31. Stratton K, Ford A, Rusch E, Clayton EW, editors. *Adverse Effects of Vaccines: Evidence and Causality*. Washington, DC: National Academies Press; 2011.
32. Nilsson L, Brockow K, Alm J, et al. Vaccination and allergy: EAACI position paper, practical aspects. *Pediatr Allergy Immunol*. 2017;28(7):628-640.
33. Su JR, Moro PL, Ng CS, Lewis PW, Said MA, Cano MV. Anaphylaxis after vaccination reported to the Vaccine Adverse Event Reporting System, 1990–2016. *J Allergy Clin Immunol*. 2019;143(4):1465-1473.
34. CDC. Vaccine Safety Datalink (VSD); 2020. https://www.CDC.gov/vaccinesafety/ensuringsafety/monitoring/vsd/index.html. Accessed January 23, 2021.
35. Bohlke K, Davis RL, Marcy SM, et al. Risk of anaphylaxis after vaccination of children and adolescents. *Pediatrics* 2003;112(4):815-820.
36. McNeil MM, Weintraub ES, Duffy J, et al. Risk of anaphylaxis after vaccination in children and adults. *J Allergy Clin Immunol*. 2016;137(3):868-878.
37. Clark S, Wei W, Rudders SA, Camargo CA Jr. Risk factors for severe anaphylaxis in patients receiving anaphylaxis treatment in US emergency departments and hospitals. *J Allergy Clin Immunol*. 2014;134(5):1125-1130.
38. Choe YJ, Lee H, Kim JH, Choi WS, Shin JY. Anaphylaxis following vaccination among children in Asia: A large-linked database study. *Allergy*. 2021;76(4):1246-1249.
39. Cheng DR, Perrett KP, Choo S, Danchin M, Buttery JP, Crawford NW. Pediatric anaphylactic adverse events following

immunization in Victoria, Australia from 2007 to 2013. *Vaccine*. 2015;33(13):1602-1607.
40. Dreskin SC, Halsey NA, Kelso JM, et al. International Consensus (ICON): allergic reactions to vaccines. *World Allergy Organ J*. 2016;9(1):32.
41. Sarti L, Lezmi G, Mori F, Giovannini M, Caubet JC. Diagnosis and management of hypersensitivity reactions to vaccines. *Expert Rev Clin Immunol*. 2020;16(9):883-896.
42. CDC. Recommendations of the Advisory Committee on Immunization Practices (ACIP): Use of Vaccines and Immune Globulins in Persons with Altered Immunocompetence; 1993. https://www.cdc.gov/mmwr/preview/mmwrhtml/00023141.htm. Accessed January 21, 2021.
43. CDC. Vaccine Excipient Summary: Excipients Included in U.S. Vaccines, by Vaccine; 2020. https://www.cdc.gov/vaccines/pubs/pinkbook/downloads/appendices/b/excipient-table-2.pdf. Accessed January 14, 2021.
44. Echeverria-Zudaire LA, Ortigosa-del Castillo L, Alonso-Lebrero E, et al. Consensus document on the approach to children with allergic reactions after vaccination or allergy to vaccine components. *Allergol Immunopathol (Madr)*. 2015;43(3):304-325.
45. World Health Organization. Draft landscape of COVID-19 candidate vaccines; 2021. https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines. Accessed January 10, 2021.
46. Rodriguez-Coira J, Sokolowska M. SARS-CoV-2 candidate vaccines - composition, mechanisms of action and stages of clinical development. *Allergy*. 2020;76(6):1922-1924.
47. Medicines and Healthcare products Regulatory Agency. Confirmation of guidance to vaccination centres on managing allergic reactions following COVID-19 vaccination with the Pfizer/BioNTech vaccine; 2020. https://www.gov.uk/government/news/confirmation-of-guidance-to-vaccination-centres-on-managing-allergic-reactions-following-covid-19-vaccination-with-the-pfizer-biontech-vaccine. Accessed January 14, 2021.
48. CDC. Allergic Reactions Including Anaphylaxis After Receipt of the First Dose of Pfizer-BioNTech COVID-19 Vaccine — United States. December 14–23, 2020; 2021. https://www.cdc.gov/mmwr/volumes/70/wr/mm7002e1.htm. Accessed January 14, 2021.
49. CDC. Allergic Reactions Including Anaphylaxis After Receipt of the First Dose of Moderna COVID-19 Vaccine — United States. December 21, 2020–January 10, 2021; 2021. https://www.cdc.gov/mmwr/volumes/70/wr/mm7004e1.htm. Accessed January 24, 2021.
50. CDC. Interim Clinical Considerations for Use of mRNA COVID-19 Vaccines Currently Authorized in the United States; 2021. https://www.cdc.gov/vaccines/covid-19/info-by-product/clinical-considerations.html#SARS-CoV-2-infection. Accessed January 24, 2021.
51. CDC. Moderna COVID-19 Vaccine Questions; 2021. https://www.cdc.gov/vaccines/covid-19/info-by-product/moderna/moderna-faqs.html. Accessed January 24, 2021.
52. Paul-Ehrlich-Institut. Corona vaccination in individuals suffereing from allergies; 2020. https://www.pei.de/EN/newsroom/positions/covid-19-vaccines/positions-corona-vaccination-allergies.html. Accessed January 14, 2021.
53. Sokolowska M, Eiwegger T, Ollert M, et al. EAACI statement on the diagnosis, management and prevention of severe allergic reactions to COVID-19 vaccines. *Allergy*. 2021. http://dx.doi.org/10.1111/all.14739. Epub ahead of print.
54. Janeway CA, Travers P, Walport M, Shlomchik MJ. *Immunobiology: The immune system in health and disease*, 5th edn. New York, NY: Garland Publishing; 2001. Part V, The Immune System in Health and Disease. https://www.ncbi.nlm.nih.gov/books/NBK10775/
55. Dispenza MC. Classification of hypersensitivity reactions. *Allergy Asthma Proc*. 2019;40(6):470-473.
56. Sampath V, Sindher SB, Alvarez Pinzon AM, Nadeau KC. Can food allergy be cured? What are the future prospects? *Allergy*. 2020;75(6):1316-1326.
57. Kelos J. Allergic reactions to vaccines; 2021. https://www.uptodate.com/contents/allergic-reactions-to-vaccines. Accessed January 14, 2021.
58. Pool V, Mege L, Abou-Ali A. Arthus Reaction as an Adverse Event Following Tdap Vaccination. *Vaccines (Basel)* 2020;8(3):385.
59. Marwa K. Type IV Hypersensitivity Reaction. In: Bayot ML, Abdelgawad I, editors. *StatPearls [Internet]*. StatPearls Publishing; 2021. https://www.ncbi.nlm.nih.gov/books/NBK562228/
60. Chung EH. Vaccine allergies. *Clin Exp Vaccine Res*. 2014;3(1):50-57.
61. Zhang B, Li Q, Shi C, Zhang X. Drug-Induced Pseudoallergy: A Review of the Causes and Mechanisms. *Pharmacology*. 2018;101(1–2):104-110.
62. Kim YN, Kim JY, Kim JW, et al. The Hidden Culprit: A Case of Repeated Anaphylaxis to Cremophor. *Allergy Asthma Immunol Res*. 2016;8(2):174-177.
63. De Luca JF, Holmes NE, Trubiano JA. Adverse reactions to vancomycin and cross-reactivity with other antibiotics. *Curr Opin Allergy Clin Immunol*. 2020;20(4):352-361.
64. Oza NB, Ryan JW, Ryan US, Berryer P, Pena G. Pulmonary anaphylaxis and the kallikrein-kinin system. *Adv Exp Med Biol*. 1979;120A:473-486.
65. Thong BY. Nonsteroidal anti-inflammatory drug hypersensitivity in the Asia-Pacific. *Asia Pac Allergy*. 2018;8(4):e38.
66. Navines-Ferrer A, Serrano-Candelas E, Lafuente A, Munoz-Cano R, Martin M, Gastaminza G. MRGPRX2-mediated mast cell response to drugs used in perioperative procedures and anaesthesia. *Sci Rep*. 2018;8(1):11628.
67. Bonvini SJ, Birrell MA, Dubuis E, et al. Novel airway smooth muscle-mast cell interactions and a role for the TRPV4-ATP axis in non-atopic asthma. *Eur Respir J*. 2020;56(1):1901458.
68. Ali H. Emerging Roles for MAS-Related G Protein-Coupled Receptor-X2 in Host Defense Peptide, Opioid, and Neuropeptide-Mediated Inflammatory Reactions. *Adv Immunol*. 2017;136:123-162.
69. McNeil BD, Pundir P, Meeker S, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature*. 2015;519(7542):237-241.
70. Tatemoto K, Nozaki Y, Tsuda R, et al. Immunoglobulin E-independent activation of mast cell is mediated by Mrg receptors. *Biochem Biophys Res Commun*. 2006;349(4):1322-1328.
71. CDC. Lab Tests to Collect Shortly After Severe Allergic Reaction/Anaphylaxis Following COVID-19 Vaccination; 2020. https://www.cdc.gov/vaccines/covid-19/clinical-considerations/testing-after-allergic-reaction.html#:~:text=Tryptase%20is%20released%20from%20mast,characterize%20the%20severe%20allergic%20reaction. Accessed January 24, 2021.
72. Sahiner UM, Yavuz ST, Buyuktiryaki B, et al. Serum basal tryptase may be a good marker for predicting the risk of anaphylaxis in children with food allergy. *Allergy*. 2014;69(2):265-268.
73. Valent P, Akin C, Arock M, et al. Definitions, criteria and global classification of mast cell disorders with special reference to mast cell activation syndromes: a consensus proposal. *Int Arch Allergy Immunol*. 2012;157(3):215-225.
74. Greiner G, Sprinzl B, Gorska A, et al. Hereditary alpha tryptasemia is a valid genetic biomarker for severe mediator-related symptoms in mastocytosis. *Blood*. 2020;37:238-247.
75. Porebski G, Kwiecien K, Pawica M, Kwitniewski M. Mas-Related G Protein-Coupled Receptor-X2 (MRGPRX2) in Drug Hypersensitivity Reactions. *Front Immunol*. 2018;9:3027.
76. Supajatura V, Ushio H, Nakao A, et al. Differential responses of mast cell Toll-like receptors 2 and 4 in allergy and innate immunity. *J Clin Invest*. 2002;109(10):1351-1359.

77. Urb M, Sheppard DC. The role of mast cells in the defence against pathogens. *PLoS Pathog*. 2012;8(4):e1002619.
78. McCurdy JD, Lin TJ, Marshall JS. Toll-like receptor 4-mediated activation of murine mast cells. *J Leukoc Biol*. 2001;70(6):977-984.
79. Kajiwara N, Sasaki T, Bradding P, et al. Activation of human mast cells through the platelet-activating factor receptor. *J Allergy Clin Immunol*. 2010;125(5):1137-1145.
80. Ferguson GT, Cole J, Aurivillius M, et al. Single-use autoinjector functionality and reliability for at-home administration of benralizumab for patients with severe asthma: GRECO trial results. *J Asthma Allergy*. 2019;12:363-373.
81. Ito T, Smrz D, Jung MY, et al. Stem cell factor programs the mast cell activation phenotype. *J Immunol*. 2012;188(11):5428-5437.
82. CDC. What's in Vaccines?; 2019. cdc.gov/vaccines/vac-gen/additives.htm. Accessed January 14, 2021.
83. CDC. Vaccine Recommendations and Guidelines of the ACIP; 2020. https://www.cdc.gov/vaccines/hcp/acip-recs/general-recs/adverse-reactions. html. Accessed May 28, 2021.
84. CDC. Latex in Vaccine Packaging; 2020. https://www.cdc.gov/vaccines/pubs/pinkbook/downloads/appendices/b/latex-table.pdf. Accessed January 14, 2021.
85. Magista S, Albanesi M, Chaoul N, et al. Safety of measles, mumps, and rubella vaccine in egg allergy: in vivo and in vitro management. *Clin Mol Allergy*. 2020;18(1):21.
86. Cerecedo Carballo I, Dieguez Pastor MC, Bartolome Zavala B, Sanchez Cano M, de la Hoz CB. Safety of measles-mumps-rubella vaccine (MMR) in patients allergic to eggs. *Allergol Immunopathol (Madr)*. 2007;35(3):105-109.
87. Upton JEM, Hummel DB, Kasprzak A, Atkinson AR. No systemic reactions to influenza vaccination in egg-sensitized tertiary-care pediatric patients. *Allergy, Asthma & Clinical Immunology*. 2012;8(1):2.
88. Kang J-H. Effectiveness and safety of seasonal influenza vaccination in children with underlying respiratory diseases and allergy. *Korean J Pediatr*. 2014;57(4):164.
89. Sharma K, Perrett KP, Wood N. Yellow Fever Vaccination In EGG-Allergic Children. *Pediatr Infect Dis J*. 2020;39(6):e76-e78.
90. CDC. Flu Vaccine and People with Egg. *Allergies*. 2020; https://www.cdc.gov/flu/prevent/egg-allergies.htm. Accessed January 14, 2021.
91. Grohskopf LA, Sokolow LZ, Broder KR, Walter EB, Bresee JS, Fry AM. Prevention and Control of Seasonal Influenza with Vaccines: Recommendations of the Advisory Committee on Immunization Practices — United States, 2017–18 Influenza Season. *MMWR. Recommend Rep*. 2017;66(2):1-20.
92. Kelso JM, Jones RT, Yunginger JW. Anaphylaxis to measles, mumps, and rubella vaccine mediated by IgE to gelatin. *J Allergy Clin Immunol*. 1993;91(4):867-872.
93. Pool V, Braun MM, Kelso JM, et al. Prevalence of anti-gelatin IgE antibodies in people with anaphylaxis after measles-mumps rubella vaccine in the United States. *Pediatrics* 2002;110(6):e71.
94. Nakayama T, Aizawa C, Kuno-Sakai H. A clinical analysis of gelatin allergy and determination of its causal relationship to the previous administration of gelatin-containing acellular pertussis vaccine combined with diphtheria and tetanus toxoids. *J Allergy Clin Immunol*. 1999;103(2):321-325.
95. Sakaguchi M, Inouye S. Systemic allergic reactions to gelatin included in vaccines as a stabilizer. *Jpn J Infect Dis*. 2000;53(5):189-195.
96. Kumagai T, Kamada M, Igarashi C, et al. Gelatin-specific cellular immune responses persist for more than 3 years after priming with gelatin containing DTaP vaccine. *Clin Exp Allergy*. 2002;32(10):1510-1514.
97. Patja A, Makinen-Kiljunen S, Davidkin I, Paunio M, Peltola H. Allergic Reactions to Measles-Mumps-Rubella Vaccination. *Pediatrics*. 2001;107(2):e27.
98. Swiontek K, Morisset M, Codreanu-Morel F, et al. Drugs of porcine origin-A risk for patients with alpha-gal syndrome? *J Allergy Clin Immunol Pract*. 2019;7(5):1687-1690.
99. Stone CA Jr, Commins SP, Choudhary S, et al. Anaphylaxis after vaccination in a pediatric patient: further implicating alpha-gal allergy. *J Allergy Clin Immunol Pract*. 2019;7(1):322-324.
100. Kuno-Sakai H, Kimura M. Removal of gelatin from live vaccines and DTaP-an ultimate solution for vaccine-related gelatin allergy. *Biologicals*. 2003;31(4):245-249.
101. Kattan JD, Konstantinou GN, Cox AL, et al. Anaphylaxis to diphtheria, tetanus, and pertussis vaccines among children with cow's milk allergy. *J Allergy Clin Immunol*. 2011;128(1):215-218.
102. Slater JE, Rabin RL, Martin D. Comments on cow's milk allergy and diphtheria, tetanus, and pertussis vaccines. *J Allergy Clin Immunol*. 2011;128(2):434; author reply 435.
103. Zanoni G, Puccetti A, Dolcino M, et al. Dextran-specific IgG response in hypersensitivity reactions to measles-mumps-rubella vaccine. *J Allergy Clin Immunol*. 2008;122(6):1233-1235.
104. Freitas DRC, Moura E, Araújo G, et al. Investigation of an outbreak of hypersensitivity-type reactions during the 2004 national measles-mumps-rubella vaccination campaign in Brazil. *Vaccine*. 2013;31(6):950-954.
105. DiMiceli L, Pool V, Kelso JM, Shadomy SV, Iskander J. Team VAERS. Vaccination of yeast sensitive individuals: review of safety data in the US vaccine adverse event reporting system (VAERS). *Vaccine*. 2006;24(6):703-707.
106. Brotherton JML, Gold MS, Kemp AS, McIntyre PB, Burgess MA, Campbell-Lloyd S. Anaphylaxis following quadrivalent human papillomavirus vaccination. *Can Med Assoc J*. 2008;179(6):525-533.
107. Kwittken PL, Rosen S, Sweinberg SK. MMR Vaccine and Neomycin Allergy. *Arch Pediatr Adolesc Med*. 1993;147(2):128.
108. Elliman D, Dhanraj B. Safe MMR vaccination despite neomycin allergy. *Lancet*. 1991;337(8737):365.
109. Huang S, Zhu Z, Cai L, et al. Analysis on the risks of severe adverse events in rabies post-exposure prophylaxis and appropriate decision-making procedure. *Human Vacc Immunotherap*. 2018;15(9):2121-2125.
110. Henao MP, Ghaffari G. Anaphylaxis to polymyxin B–trimethoprim eye drops. *Ann Allergy Asthma Immunol*. 2016;116(4):372.
111. Dréno B, Zuberbier T, Gelmetti C, Gontijo G, Marinovich M. Safety review of phenoxyethanol when used as a preservative in cosmetics. *J Eur Acad Dermatol Venereol*. 2019;33(S7):15-24.
112. Smith D, Gomez R, Davenport J. Latex content in adult vaccines. *Mil Med*. 2020;185(3–4):354-355.
113. Russell M, Pool V, Kelso JM, Tomazic-Jezic VJ. Vaccination of persons allergic to latex: a review of safety data in the Vaccine Adverse Event Reporting System (VAERS). *Vaccine*. 2004;23(5):664-667.
114. U.S. FDA. Thimerosal and Vaccines; 2018. https://www.fda.gov/vaccines-blood-biologics/safety-availability-biologics/thimerosal-and-vaccines. Accessed January 24, 2021.
115. Ball LK, Ball R, Pratt RD. An assessment of thimerosal use in childhood vaccines. *Pediatrics*. 2001;107(5):1147-1154.
116. Jacob SE, Brod B, Crawford GH. Clinically relevant patch test reactions in children-a United States based study. *Pediatr Dermatol*. 2008;25(5):520-527.
117. Centers for Disease C. Systemic allergic reactions following immunization with human diploid cell rabies vaccine. *MMWR Morb Mortal Wkly Rep*. 1984;33(14):185-187.
118. Huang DB, Wu JJ, Tyring SK. A review of licensed viral vaccines, some of their safety concerns, and the advances in the development of investigational viral vaccines. *J Infect*. 2004;49(3):179-209.
119. Swanson MC, Rosanoff E, Gurwith M, Deitch M, Schnurrenberger P, Reed CE. IgE and IgG antibodies to beta-propiolactone and human serum albumin associated with urticarial reactions to rabies vaccine. *J Infect Dis*. 1987;155(5):909-913.

120. Fishbein DB, Yenne KM, Dreesen DW, Teplis CF, Mehta N, Briggs DJ. Risk factors for systemic hypersensitivity reactions after booster vaccinations with human diploid cell rabies vaccine: a nationwide prospective study. *Vaccine*. 1993;11(14):1390-1394.
121. Xia S, Zhang Y, Wang Y, et al. Safety and immunogenicity of an inactivated SARS-CoV-2 vaccine, BBIBP-CorV: a randomised, double-blind, placebo-controlled, phase 1/2 trial. *Lancet Infect Dis*. 2021;21(1):39-51.
122. Zhang Y, Zeng G, Pan H, et al. Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine in healthy adults aged 18–59 years: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial. *Lancet Infect Dis*. 2021;21(2):181-192.
123. Ring J. Exacerbation of eczema by formalin-containing hepatitis B vaccine in formaldehyde-allergic patient. *Lancet*. 1986;328(8505):522-523.
124. Kuritzky LA, Pratt M. Systemic allergic contact dermatitis after formaldehyde-containing influenza vaccination. *J Cutaneous Med Surg*. 2015;19(5):504-506.
125. Moghaddam A, Olszewska W, Wang B, et al. A potential molecular mechanism for hypersensitivity caused by formalin-inactivated vaccines. *Nat Med*. 2006;12(8):905-907.
126. Peebles JRS, Sheller James R, Collins Robert D, Jarzecka K, Mitchell Daphne B, Graham BS. Respiratory Syncytial Virus (RSV)–Induced Airway Hyperresponsiveness in Allergically Sensitized Mice Is Inhibited by Live RSV and Exacerbated by Formalin-Inactivated RSV. *J Infect Dis*. 2000;182(3):671-677.
127. Lauren CT, Belsito DV, Morel KD, LaRussa P. Case report of subcutaneous nodules and sterile abscesses due to delayed type hypersensitivity to aluminum-containing vaccines. *Pediatrics*. 2016;138(4):e20141690.
128. Kelly E, Leahy R, McDermott M, et al. Persistent pruritic subcutaneous nodules at injection sites and other delayed type hypersensitivity reactions to aluminium adsorbed vaccines in Irish children: A case series. *Acta Paediatr*. 2020;109(12):2692-2693.
129. Gordon SC, Bartenstein DW, Tajmir SH, Song JS, Hawryluk EB. Delayed-type hypersensitivity to vaccine aluminum adjuvant causing subcutaneous leg mass and urticaria in a child. *Pediatr Dermatol*. 2018;35(2):234-236.
130. Salik E, Løvik I, Andersen K, Bygum A. Persistent Skin Reactions and Aluminium Hypersensitivity Induced by Childhood Vaccines. *Acta Derm Venereol*. 2016;96(7):967-971.
131. Bergfors E, Trollfors B. Sixty-four children with persistent itching nodules and contact allergy to aluminium after vaccination with aluminium-adsorbed vaccines—prognosis and outcome after booster vaccination. *Eur J Pediatr*. 2012;172(2):171-177.
132. Johnson-Weaver BT, McRitchie S, Mercier KA, et al. Effect of endotoxin and alum adjuvant vaccine on peanut allergy. *J Allergy Clin Immuno*. 2018;141(2):791-794.
133. Montana M, Verhaeghe P, Ducros C, Terme T, Vanelle P, Rathelot P. Safety review: squalene and thimerosal in vaccines. *Therapie*. 2010;65(6):533-541.
134. Lippi G, Targher G, Franchini M. Vaccination, squalene and anti-squalene antibodies: facts or fiction? *Eur J Intern Med*. 2010;21(2):70-73.
135. Shi S, Zhu H, Xia X, Liang Z, Ma X, Sun B. Vaccine adjuvants: Understanding the structure and mechanism of adjuvanticity. *Vaccine*. 2019;37(24):3167-3178.
136. Meller S, Gerber PA, Kislat A, et al. Allergic sensitization to pegylated interferon-alpha results in drug eruptions. *Allergy*. 2015;70(7):775-783.
137. Castells MC, Phillips EJ. Maintaining Safety with SARS-CoV-2 Vaccines. *N Engl J Med*. 2021;384(7):643-649.
138. Cabanillas B, Akdis C, Novak N. Allergic reactions to the first COVID-19 vaccine: a potential role of polyethylene glycol? *Allergy*. 2020;76(6):1617-1618.
139. Pfaar O, Mahler V. Allergic reactions to COVID-19 -vaccination - unveiling the secret(s). *Allergy*. 2021;76(6):1617-1618.
140. Kozma GT, Shimizu T, Ishida T, Szebeni J. Anti-PEG antibodies: Properties, formation, testing and role in adverse immune reactions to PEGylated nano-biopharmaceuticals. *Adv Drug Deliv Rev*. 2020;155:163-175.
141. Stone CA, Liu Y, Relling MV, et al. Immediate hypersensitivity to polyethylene glycols and polysorbates: more common than we have recognized. *J Allergy Clinical Immunol Pract*. 2019;7(5):1533-1540.
142. Povsic TJ, Lawrence MG, Lincoff AM, et al. Pre-existing anti-PEG antibodies are associated with severe immediate allergic reactions to pegnivacogin, a PEGylated aptamer. *J Allergy Clin Immunol*. 2016;138(6):1712-1715.
143. Banerji A, Wickner PG, Saff R, et al. mRNA Vaccines to Prevent COVID-19 Disease and Reported Allergic Reactions: Current Evidence and Suggested Approach. *J Allergy Clin Immunol Pract*. 2020;9(4):1423-1437.
144. Maggio E. Polysorbates, biotherapeutics and anaphylaxis: a review. *Bioprocess Int*. 2017;15(8). https://bioprocessintl.com/manufacturing/formulation/polysorbates-biotherapeutics-and-anaphylaxis-a-review/
145. Badiu I, Geuna M, Heffler E, Rolla G. Hypersensitivity reaction to human papillomavirus vaccine due to polysorbate 80. *BMJ Case Rep*. 2012;2012:bcr0220125797.
146. Hong L, Wang Z, Wei X, Shi J, Li C. Antibodies against polyethylene glycol in human blood: A literature review. *J Pharmacol Toxicol Methods*. 2020;102:106678.
147. Nanomedicine and the COVID-19 vaccines. *Nat Nanotechnol*. 2020;15(12):963. http://dx.doi.org/10.1038/s41565-020-00820-0
148. Jackson LA, Anderson EJ, Rouphael NG, et al. An mRNA Vaccine against SARS-CoV-2 - Preliminary Report. *N Engl J Med*. 2020;383(20):1920-1931.
149. de Vrieze J. Pfizer's vaccine raises allergy concerns. *Science*. 2021;371(6524):10-11.
150. Cabanillas B, Akdis C, Novak N. Allergic reactions to the first COVID-19 vaccine: a potential role of polyethylene glycol? *Allergy*. 2020;76(6):1621-1623.
151. Garvey LH, Nasser S. Anaphylaxis to the first COVID-19 vaccine: is polyethylene glycol (PEG) the culprit? *Br J Anaesth*. 2021;126(3):e106-e108.
152. Bruusgaard-Mouritsen MA, Johansen JD, Garvey LH. Clinical manifestations and impact on daily life of allergy to polyethylene glycol (PEG) in ten patients. *Clin Exp Allergy*. 2021;51(3):463-470.
153. Zhou ZH, Stone CA Jr, Jakubovic B, et al. Anti-PEG IgE in anaphylaxis associated with polyethylene glycol. *J Allergy Clin Immunol Pract*. 2020;9(4):1731-1733.e3. http://dx.doi.org/10.1016/j.jaip.2020.11.011
154. Klimek L, Jutel M, Akdis CA, et al. ARIA-EAACI statement on severe allergic reactions to COVID-19 vaccines – an EAACI-ARIA position paper. *Allergy*. 2020;76(6):1624-1628.
155. Hatziantoniou S, Maltezou HC, Tsakris A, Poland GA, Anastassopoulou C. Anaphylactic reactions to mRNA COVID-19 vaccines: A call for further study. *Vaccine*. 2021;39(19):2605-2607.
156. Sokolowska M, Lukasik ZM, Agache I, et al. Immunology of COVID-19: Mechanisms, clinical outcome, diagnostics, and perspectives-A report of the European Academy of Allergy and Clinical Immunology (EAACI). *Allergy*. 2020;75(10):2445-2476.
157. European Medicines Agency. Annex I: Summary of product characteristics. https://www.ema.europa.eu/en/documents/product-information/comirnaty-epar-product-information_en.pdf. Accessed January 10, 2021.
158. Sasisekharan V, Pentakota N, Jayaraman A, Tharakaraman K, Wogan GN, Narayanasami U. Orthogonal immunoassays for IgG antibodies to SARS-CoV-2 antigens reveal that immune response lasts beyond 4 mo post illness onset. *Proc Natl Acad Sci U S A*. 2021;118(5):e2021615118.

159. Hartley GE, Edwards ESJ, Aui PM, et al. Rapid generation of durable B cell memory to SARS-CoV-2 spike and nucleocapsid proteins in COVID-19 and convalescence. *Sci Immunol*. 2020;5(54):eabf8891.
160. Wajnberg A, Amanat F, Firpo A, et al. Robust neutralizing antibodies to SARS-CoV-2 infection persist for months. *Science*. 2020;370(6521):1227-1230.
161. Xie X, Zou J, Fontes-Garfias CR, et al. Neutralization of N501Y mutant SARS-CoV-2 by BNT162b2 vaccine-elicited sera. *bioRxiv*. 2001;2021(2021):2007.
162. Ashraf MU, Kim Y, Kumar S, Seo D, Ashraf M, Bae YS. COVID-19 vaccines (revisited) and oral-mucosal vector system as a potential vaccine platform. *Vaccines (Basel)*. 2021;9(2):171.
163. European Medicines Agency. Guideline on good pharmacovigilance practices (GVP). Product- or Population-Specific Considerations I: Vaccines for prophylaxis against infectious diseases; 2012. https://www.ema.europa.eu/en/documents/scientific-guideline/draft-guideline-good-pharmacovigilance-practices-gvp-product-population-specific-considerations-i_en.pdf. Accessed January 14, 2021.
164. Brotherton JM, Gold MS, Kemp AS, et al. Anaphylaxis following quadrivalent human papillomavirus vaccination. *CMAJ*. 2008;179(6):525-533.
165. Erlewyn-Lajeunesse M, Hunt LP, Heath PT, Finn A. Anaphylaxis as an adverse event following immunisation in the UK and Ireland. *Arch Dis Child*. 2012;97(6):487-490.
166. Zent O, Beran J, Jilg W, Mach T, Banzhoff A. Clinical evaluation of a polygeline-free tick-borne encephalitis vaccine for adolescents and adults. *Vaccine*. 2003;21(7–8):738-741.
167. Ozaki T, Nishimura N, Muto T, et al. Safety and immunogenicity of gelatin-free varicella vaccine in epidemiological and serological studies in Japan. *Vaccine*. 2005;23(10):1205-1208.
168. Rouleau I, De Serres G, Drolet JP, et al. Increased risk of anaphylaxis following administration of 2009 AS03-adjuvanted monovalent pandemic A/H1N1 (H1N1pdm09) vaccine. *Vaccine*. 2013;31(50):5989-5996.
169. Kelso JM, Mootrey GT, Tsai TF. Anaphylaxis from yellow fever vaccine. *J Allergy Clin Immunol*. 1999;103(4):698-701.
170. D'Souza RM, Campbell-Lloyd S, Isaacs D, et al. Adverse events following immunisation associated with the 1998 Australian Measles Control Campaign. *Commun Dis Intell*. 2000;24(2):27-33.
171. Kawai AT, Li L, Kulldorff M, et al. Absence of associations between influenza vaccines and increased risks of seizures, Guillain-Barre syndrome, encephalitis, or anaphylaxis in the 2012–2013 season. *Pharmacoepidemiol Drug Saf*. 2014;23(5):548-553.
172. Ropero-Alvarez AM, Whittembury A, Bravo-Alcantara P, Kurtis HJ, Danovaro-Holliday MC, Velandia-Gonzalez M. Events supposedly attributable to vaccination or immunization during pandemic influenza A (H1N1) vaccination campaigns in Latin America and the Caribbean. *Vaccine*. 2015;33(1):187-192.
173. Halsey NA, Griffioen M, Dreskin SC, et al. Immediate hypersensitivity reactions following monovalent 2009 pandemic influenza A (H1N1) vaccines: reports to VAERS. *Vaccine*. 2013;31(51):6107-6112.
174. Duclos P. Safety of immunisation and adverse events following vaccination against hepatitis B. *Expert Opin Drug Saf*. 2003;2(3):225-231.
175. Li X, Ma SJ, Liu X, et al. Immunogenicity and safety of currently available Japanese encephalitis vaccines: a systematic review. *Hum Vaccin Immunother*. 2014;10(12):3579-3593.

How to cite this article: Sampath V, Rabinowitz G, Shah M, et al. Vaccines and allergic reactions: The past, the current COVID-19 pandemic, and future perspectives. *Allergy*. 2021;76:1640–1660. https://doi.org/10.1111/all.14840