Polyethylene glycol (PEG): The nature, immunogenicity, and role in the hypersensitivity of **PEGylated products** Mohamed Ibrahim, a,b Eslam Ramadan, a,b Nehal E. Elsadek, a Sherif E. Emam, a,c Taro Shimizu, a, Hidenori Ando, a Yu Ishima, a Omar Helmy Elgarhy, h Hatem A. Sarhan, h Amal K Hussein, h and Tatsuhiro Ishida^{a,*} ^a Department of Pharmacokinetics and Biopharmaceutics, Institute of Biomedical Sciences, Tokushima University; 1–78–1 Sho-machi, Tokushima 770–8505, Japan. ^b Department of Pharmaceutics and Industrial pharmacy, Faculty of Pharmacy, Minia University, 61519 Minia, Egypt. ^c Department of Pharmaceutics and Industrial Pharmacy, Faculty of Pharmacy, Zagazig University; Zagazig 44519, Egypt. *Corresponding author. Tel: +81-88-633-7260, Fax: +81-88-633-7259 E-mail address: ishida@tokushima-u.ac.jp

Abstract

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16 17

Polyethylene glycol (PEG) is a versatile polymer that is widely used as an additive in foods and cosmetics, and as a carrier in PEGylated therapeutics. Even though PEG is thought to be less immunogenic, or perhaps even non-immunogenic, with a variety of physicochemical properties, there is mounting evidence that PEG causes immunogenic responses when conjugated with other materials such as proteins and nanocarriers. Under these conditions, PEG with other materials can result in the production of anti-PEG antibodies after administration. The antibodies that are induced seem to have a deleterious impact on the therapeutic efficacy of subsequently administered PEGylated formulations. In addition, hypersensitivity to PEGylated formulations could be a significant barrier to the utility of PEGylated products. Several reports have linked the presence of anti-PEG antibodies to incidences of complement activation-related pseudoallergy (CARPA) following the administration of PEGylated formulations. The use of COVID-19 mRNA vaccines, which are composed mainly of PEGylated lipid nanoparticles (LNPs), has recently gained wide acceptance, although many cases of post-vaccination hypersensitivity have been documented. Therefore, our review focuses not only on the importance of PEGs and its great role in improving the therapeutic efficacy of various medications, but also on the hypersensitivity reactions attributed to the use of PEGylated products that include PEG-based mRNA COVID-19 vaccines.

18 19

20

Key words:

- 21 Polyethylene glycol (PEG); anti-PEG antibodies; Hypersensitivity; COVID-19 mRNA vaccines;
- 22 complement activation-related pseudoallergy (CARPA)

24

23

1. Introduction

1

2

3

4

5

6

7

8 9

10

11 12

13

14

15

16 17

18

19

20

21 22

23

24

25

26

27 28

29

30

31

Polyethylene glycol (PEG) is a synthesized polymer that is widely used in many industries and pharmaceutical formulations due to its well established safety and versatile physicochemical properties [1]. It is noteworthy that PEGs and their derivatives are widely employed in pharmaceutical products as a component, in non-pharmaceutical products as additives [2], and in cosmetics as emulsifiers, lubricants and humectants [3, 4]. PEGs were first used as a lubricant for medical equipment in the 1950s, and they have since been used as anti-freeze agents, food additives, and as a vehicle to carry additives in tablets and in dermatological formulations [3, 5]. PEGs with a high molecular weight are usually used in cosmetics as skin conditioners, surfactants, and as a cleansing agent, in addition to use in other topical preparations such as haircare products, lotions, creams, lipsticks, and toothpaste [6]. PEGs are also used as a suppository base due to approved hydrophilicity, which allows rapid miscibility with rectal mucosal fluids and a rapid release of drug molecules [7]. PEG 400 is used as a co-solvent in the preparation of Nifedipine soft gelatine capsules to improve oral absorption [8]. In addition, Abuchowski and colleagues pioneered the use of PEGs in the delivery of proteins in a technique known as PEGylation [9]. PEGylation is considered a breakthrough in the field of drug delivery with dozens of applications. PEGylation enhances the biological half-lives of biopharmaceuticals while reducing toxicity and improving stability [10, 11]. Despite the great importance of PEGylation, several limitations have been connected to the use of PEGylation in pharmaceutical formulations; there is growing evidence of the immunogenicity of PEG when conjugated with large molecules such as proteins, and when used as nanocarriers for liposomes and other drug molecules [11-13].

Over the past few decades, PEG has been considered a non-immunogenic molecule that can be safely used as an excipient in medications, cosmetics, and as a food additive [12, 14]. Recently, however, a growing number of reports have suggested that PEG is an immunogenic molecule; therefore, care should be taken when these molecules are used in pharmaceutical formulations, particularly with proteins, lipid nanoparticles, and liposomes [15, 16]. Anti-PEG antibodies could compromise the therapeutic efficacy of subsequently administered PEGylated products both in patients who have received PEGylated products and in healthy individuals. These antibodies tend to affect the distribution and enhance the clearance of PEGylated products, which is the so-called accelerated blood clearance (ABC) phenomenon [17]. Our group and

others have demonstrated how the first dose of PEGylated products could enhance the clearance of a second dose injected between 5 and 7 days later, which in turn affects the therapeutic efficacy of administered PEGylated products [18-21]. Unfortunately, these antibodies not only affect the therapeutic efficacy of PEGylated products but also cause immediate hypersensitivity reactions upon administration of PEGylated products. The PEGylated liposomal formulation doxorubicin (Doxil®) is known to cause immediate hypersensitivity reactions that cannot be explained based on the conventional hypothesis of IgE-mediated type I hypersensitivity. Szebeni *et al.* suggested a rationale and supplied empirical evidence for the concept that these responses represent a novel type of drug-induced hypersensitivity, which is referred to as complement activated-related pseudoallergy (CARPA) [22]. Also, several cases of hypersensitivity have been reported following the use of Oncaspar® (Pegaspargase), which is PEGylated L-asparaginase that has been approved for the treatment of Acute Lymphoblastic Leukemia (ALL). Reports of hypersensitivity reactions to Oncaspar® have also been attributed to anti-PEG antibodies that develop CARPA [23-25].

Due to the rapid spread of the recent COVID-19 pandemic, public health officials now look to the development of treatments or vaccines to limit the spread of diseases. Pharmaceutical companies successfully developed mRNA vaccines such as Pfizer/BioNTech (BNT162b2) [26] and Moderna (mRNA-1273) [27] COVID-19, which can help in limiting the spread of COVID-19, decrease the symptoms, and decrease the mortality rate [28-30]. Unfortunately, despite the success and approved efficacy of the mRNA COVID-19 vaccines, cases of allergies to these vaccines have been reported [31-33]. Thus far, the main causes and exact mechanisms of hypersensitivity to mRNA COVID-19 vaccines have not been fully elucidated, but reports of hypersensitivity reactions have focused on the role of the PEG polymer that is used in the preparation of these vaccines [34-37]. These reports studied the previous history of these cases and pointed out that all these cases received PEG-containing medications such as Depo-Provera, esomeprazole, Naproxen, or osmotic laxative. The formulations resemble shaving foam with low and high molecular weights of PEG and PEG-containing cosmetics [31, 32, 35]. Accordingly, in this review we have attempted to provide insight into the importance of the use of PEG in various areas of life including therapeutic and non-therapeutic applications. In addition, we explain the potential role of PEG in the reports of the immunogenicity and hypersensitivity that has been encountered post-mRNA COVID-19 vaccination.

2. Polyethylene glycols (PEGs)

2.1. PEG properties

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16 17

18

19

20

21 22

23

24

25

26

27

28

29

30

31

PEG, also known as polyoxyethylene (POE) or polyethylene oxide (PEO), is a bio-inert, biocompatible polymer. It is a synthetic hydrophilic polymer composed of repeated units of ethylene oxide, as illustrated in Figure 1. Macrogol, Polikol, Polygol, Polyox, Polyoxirane, poly(oxy-1,2-ethanediyl), and CarbowaxTM are some of the PEG-based compounds on the market. PEGs are classified using two nomenclature systems: one is based on the Chemical Abstract Service (CAS), and the other is based on the Cosmetics, Toiletry and Fragrance Association (CTFA)/International Nomenclature Cosmetic Ingredients (INCI). In cosmetics, the suffix number to PEG denotes the number of repeating oxyethylene units; for example, PEG 50 denotes the presence of 50 oxyethylene subunits in this particular PEG polymer [6, 38]. PEG 50 is widely used in pharmaceutical applications due to its aqueous solubility, biocompatibility, and safety [39]. The amphiphilic nature of PEGs makes them soluble in a wide range of organic solvents, including chloroform, ethanol, acetonitrile, and acetone in addition to a high level of water solubility [10]. PEGs are thermally stable and electrically neutral at different levels of pH, and they have highly active multifunctional terminal groups. The terminal hydroxyl group (-OH) can bind with different molecules through covalent or hydrogen bonding interaction. For example, PEG-Intron®, a mono-PEGylated INF-α2b, is synthesized using a succinimidyl carbonate PEG reagent (12-kDa mPEG SC). The mPEG SC reagent forms a covalent carbamate and/or urethane linker with amine groups on the protein [40]. Also, every molecule of PEG 3,350 has the ability to attach to 100 molecules of water through hydrogen bonding [41]. Therefore, PEG polymer is widely used as a hydrophobic drug carrier to promote aqueous solubility and dissolution. In addition, the terminal end of PEGs has a great ability to attach to various bioactive functional groups for a variety of applications [42].

PEGs can be synthesized in a wide range of molecular weights with a variety of properties. Commercially available PEGs have molecular weights ranging from 200 to 35,000 Da and come in a variety of forms and degrees of branching [43]. Low molecular weight PEGs are usually synthesized through the addition of an ethylene oxide subunit together with hydroxyl group donors such as water or any diols in the presence of alkaline catalysts. High molecular weight PEGs, on the other hand, are usually prepared using suspension polymerization in order to achieve large-scale production. They also can be prepared through the anionic polymerization of

ethylene oxide in an inert solvent or through anionic ring-opening polymerization of epoxides [44, 45]. According to their degree of polymerization and molecular weights, PEGs have distinct states and melting temperatures. PEGs with low molecular weights (100-700 Da) are viscous, colorless liquids, while those with molecular weights that range from 1,000 to 2,000 Da are soft solids, and those with higher molecular weights (>2,000 Da) are solid, waxy, and white in color, with melting points proportional to their molecular weights [46, 47]. Penetration of PEG molecules depends mainly on their molecular weight, except in the case of compromised skin, which PEG molecules can penetrate irrespective of their molecular weight — as in the case of burns [48]. Poor penetration of PEGs could be attributed to their hydrophilicity [49]. On the other hand, PEG derivatives can be used as penetration enhancers. For example, PEG stearate has a low molecular weight that enhances the penetration of other drug molecules by decreasing the skin surface tension and conditioning the stratum corneum [4].

2.2. Pharmacokinetics and the fate of PEG in the body

The gastrointestinal absorption and skin penetration of PEG molecules depends mainly on their molecular weight. Chadwick *et al.* reported that PEG 400 is well absorbed from the gastrointestinal tract after oral administration and half of administered doses is excreted mainly through the kidney within 24 h in humans [50]. PEGs are believed to be barely absorbed via intact skin, however, and studies have shown that those with higher molecular weights (4,000 Da or more) may not be absorbed at all [51]. On the other hand, as observed by Herold *et al.* [48] and Tsai *et al.* [52], the presence of injury or damage in the epidermal layers may accelerate the penetration of PEG molecules regardless of their molecular weight. After reaching systemic circulation, PEG is metabolized via slow oxidation of their hydroxyl group to form carboxylic acid, diacids, and hydroxy acid metabolites, which are catalysed by the alcohol dehydrogenase enzyme [53] and some other oxidase enzymes such as cytochrome P-450 [54]. PEG molecules (20,000 Da) are excreted mainly via the renal route, whereas PEGs with molecular weights between 20,000-50,000 Da are primarily excreted via the biliary route rather than the renal route, while PEG with molecular weights larger than 50,000 Da are primarily engulfed by liver macrophages [55, 56].

2.3. Pharmaceutical applications of PEGs

2.3.1. Solubility enhancing agent

Drugs with poor aqueous solubility exhibit poor bioavailability, particularly drugs belonging to biopharmaceutical classification system (BCS) classes 2 and 4 [57]. PEGs are employed as a solubility enhancer because of their strong polarity and solubility in a variety of aqueous and organic solvents, which allows them to interact more effectively with hydrophobic drug molecules [58]. In parenteral and oral preparations, liquid PEGs (up to 1,000 Da) are commonly utilized as water-miscible solubilizing agents. In parenteral and oral preparations, liquid PEGs (up to 1,000 Da) are commonly utilized as water-miscible solubilizing agents. PEGs with high molecular weight (1,000-6,000 Da) are mostly employed to improve the aqueous solubility of microencapsulated hydrophobic medicines where solubilization occurs at higher concentrations than the critical micelle concentration (CMC) of PEG derivatives. Paclitaxel is a potent chemotherapeutic agent that has been approved by the Food and Drug Administration (FDA) for breast and ovarian cancer treatment. PEGylation of Paclitaxel improves its water solubility, and this reflects its liposomal encapsulation efficiency and physical stability. Also, the bioavailability of PEGylated paclitaxel was 3.9-fold higher than conventional non-PEGylated paclitaxel. The increased bioavailability of PEGylated paclitaxel might have resulted from the physicochemical properties of the PEGylated paclitaxel, which is a water-soluble compound and can easily permeate through the gastrointestinal mucosa than non-PEGylated paclitaxel which leads to increase the concentration of paclitaxel in the plasma and subsequently improve the bioavailability than the parent drug [59].

2.3.2. Drug passive targeting

1

2

3

4

5

6

7

8

9

10

11 12

13

14 15

16

17

18

19

20

21

22

23

24

25

26

27 28

29

30

31

Uneven biodistribution of pharmaceuticals as well as their rapid clearance represent the main challenges in systemic drug administration, which can be alleviated by PEGylation. PEGylation is used to cover the drug surface with a protective hydrophilic coat, which results in an increase in drug particle size, reducing its glomerular filtration. Furthermore, the PEG coat protects drug molecules from enzymes and plasma protein adsorption, resulting in improved in vivo stability and extended circulation time, allowing for enhanced passive drug targeting [60]. The effect of PEGylation on the formulation's stability and passive drug targeting is summarized in **Figure 2**.

PEGylation is thought to be a useful approach for delivering anti-cancer drugs encapsulated in nanocarrier systems. PEGylation is a procedure that involves covalently grafting PEG chains onto the surfaces of other molecules to form nanocarrier systems. PEGylation is

known to improve the stability and plasma half-life of various medications [61, 62]. PEGylation is suggested to prolong the half-life of drug plasma by reducing the protein opsonin adsorption on the surface of nanocarrier systems, which prevents their uptake by the cells of the mononuclear phagocyte system (MPS) (the Stealth effect) [10, 63]. The potential of long-circulating PEGylated nanocarrier systems to pass through leaky blood vessels and accumulate within tumors via enhanced permeability and retention (EPR) [64, 65] is the proposed mechanism for passive drug targeting of tumors [66].

Fukuda et al. reported that PEGylated liposomes of doxorubicin (Doxil®) improved the pharmacokinetics and minimized the toxicity of doxorubicin by improving the biodistribution and enhancing the accumulation of doxorubicin in tumor tissues. They also reported, however, that although the use of non-PEGylated liposomes of doxorubicin, as Myocet®, reduced doxorubicin cardiotoxicity and gastrotoxicity; the drug was released more rapidly and had a short plasma circulation time [67]. Park et al. reported that encapsulation of doxorubicin into PEGylated nanoparticles maximized therapeutic efficacy while decreasing dose-related cardiotoxicity, and found that using PEG to make nanoparticles allowed for effective and safe doxorubicin administration [68]. Also, a report by O'Shaughnessy et al. associated a formulation of doxorubicin in the form of a PEGylated liposomal system with increasing the therapeutic index of conventional doxorubicin. They have also reported that PEGylated liposomal doxorubicin improves drug targeting efficacy without many of the side effects usually reported with the use of conventional doxorubicin therapy such as nausea, vomiting, alopecia, myelosuppression, and cardiac toxicity [69]. In the same manner, Safara et al. reported that Doxil® can reduce the risk of cardiomyopathy incidence in patients with solid tumors compared with those receiving free doxorubicin [70]

Some medications' short plasma half-lives may limit their therapeutic use. The cells of MPS are capable of engulfing hydrophobic materials, liposomes, peptides, and genes from systemic circulation [71]. The physicochemical features of drug molecules, such as particle size, hydrophilicity, and surface charge have a significant impact on the recognition of drug molecules by MPS and subsequently on the fate of drug molecules in the body [72, 73]. PEGylation could reduce the clearance of drug molecules by increasing the particle size and preventing the interaction with MPS cells.

The first approved application for the PEGylation method was ADAGEN® (pegademase bovine), which was first approved by the FDA in 1990 for treating severe combined immunodeficiency disease [74]. Doxil® also can ensure effective drug distribution with reduced toxicity [75, 76]. PEGylation of peptides, such as Pegasys® (peginterferon alfa-2a), a clinically approved PEGylated protein for the treatment of hepatitis B and C [40, 77], can also help protect peptides against enzyme hydrolysis, which improves the therapeutic results and stability of particular peptides. In the same manner, in 2004 Macugen® was the first approved PEGylated aptamer for the treatment of neovascular age-related macular degeneration [78]. Examples of clinically approved PEGylated products on the market with improved pharmacokinetics are summarized in **Table 1**.

Table 1. Examples of clinically approved PEGylated products

Marketed	PEGylated entity	Type of PEG	Half-life	Half-life	Therapeutic use	Year	Ref.
product			Before	After			
			PEGylation	PEGylation			
Adagen®	Adenosine	5 kDa PEG	11-22 h	72-144 h	Severe Combined	1990	[79]
	deaminase				Immunodeficiency		
					Disease (SCID)		
Doxil®	PEGylated	2 kDa PEG	17.3 h	69.3 h	Ovarian cancer,	1995	[80]
	liposomal				Breast cancer,		
	Doxorubicin				Kaposi's sarcoma		
PEG-	Interferon-alfa-2b	12 kDa PEG	12 h	48-72 h	hepatitis C	2001	[81]
Intron®							
Pegasys®	Interferon-alfa-2a	40 kDa bis-	3-8 h	65 h	Hepatitis C	2002	[82]
		monomethox					
		y PEG					
Somavert®	Human growth	5 kDa PEG	24-36 h	144 h	Acromegaly	2003	[83]
	hormone						
Macugen [®]	Anti-Vascular	40 kDa	9 h	240 h	Age-related	2004	[84]
	endothelial	mPEG			muscular		
	growth factor				degeneration		
	(anti-VEGF)						
Mircera®	Erythropoietin	30 kDa	7-20 h	134-139 h	Anaemia related to	2007	[85, 86]
		Methoxy			kidney disorders		
		polyethylene					
		glycol					
Cimzia®	Anti-tumor	20 kDa PEG	4.6 h	313 h	Rheumatoid	2008	[87]
	necrosis factor				arthritis		
	antibody						
Krystexxa®	Recombinant	10 kDa	4 h	154-331 h	Chronic gout	2010	[88]
	uricase	mPEG					
Rebinyn®	Recombinant	40 kDa PEG	19.34 h	92.76 h	Haemophilia B	2017	[89]
	coagulation factor						
	1X						

Jivi®	Recombinant	30 kDa PEG	13 h	17-21 h	Haemophilia A	2017	[90]
	antihemophilic						
	factor VIII						
Asparlas®	L-asparaginase	31-93* 5 kDa	31.2 h	384 h	Leukemia	2018	[91, 92]
Palynziq®	Recombinant	20 kDa PEG	21 h	60 h	Phenylketonuria	2018	[93]
	phenylalanine						
	ammonia lyase						
Esperoct®	Recombinant	40 kDa PEG	11.8 h	17-22 h	Haemophilia A	2019	[94]
	antihemophilic						
	factor VIII						
Ziextenzo®	Granulocyte	20 kDa PEG	3-4 h	15-80 h	Infection during	2019	[95]
	colony				chemotherapy		
	stimulating factor						
	(G-CSF)						
Udenyca®	G-CSF	20 kDa PEG	3-4 h	15-80 h	Neutropenia	2019	[96]
Nyvepria®	G-CSF	20 kDa PEG	3-4 h	15-80 h	Neutropenia	2020	[97]
					associated		
					chemotherapy		
Besremi®	Interferon	40 kDa PEG	2-3 h	60-70 h	Polycythaemia vera	2021	[98, 99]
Skytrofa [®]	Human growth	40 kDa PEG	2-4 h	25 h	Growth hormone	2021	[100]
	hormone				deficiency		
Empaveli®	Pentadecapeptide	40 kDa PEG	4 h	192 h	Paroxysmal	2021	[101]
					Nocturnal		
					Hemoglobinuria		
					(PNH)		

2.4. Immunogenicity of PEGs

2.4.1. Immunogenicity of free PEG

Many studies have been published concerning how the immunogenicity of PEGs could jeopardize the efficacy and safety of PEGylated products [102, 103]. Over the past few decades PEG has generally been recognized as safe and as such is commonly used as a food additive and in pharmaceutical formulations. However, much recent evidence has suggested the presence of pre-existing anti-PEG antibodies in healthy people [104-106]. Anti-PEG antibodies have been found in the blood of about 25% of healthy blood donors, as reported by Garay *et al.* [14]. Similarly, Yang *et al.* reported that substantial levels of anti-PEG antibodies were found in a large percentage of people (> 42%) who had never received PEGylated pharmaceuticals [107].

The presence of anti-PEG antibodies in individuals who never received PEGylated pharmaceuticals may be attributed to their frequent usage of PEG-containing products such as in cosmetics where PEG is a commonly used ingredient. Yang and Lai have also provided a possible explanation for the occurrence of anti-PEG antibodies in healthy people, claiming that any irritation, injury, or abrasion in the skin triggers local inflammatory reactions. Upon frequent use of commonly used cleaning or cosmetic products containing PEG molecules, these molecules can penetrate a site of inflammation and come into contact with inflammatory cells, which would trigger the formation of anti-PEG antibodies [107]. In the same manner, Jakasa *et al.* reported that depending on their molecular weight, PEGs could penetrate the stratum corneum and reach systemic circulation. They also reported that the condition of the skin may represent a critical factor in determining PEG skin permeability. The presence of any defect in the skin barrier, as in the case of atopic dermatitis (AD), could enhance the penetration of PEG molecules of different molecular weights. They also reported that the permeation coefficient of PEG molecules was doubled in cases of compromised skin compared with the rate for normally intact skin [108].

2.4.2. Immunogenicity of PEGylated products

Immunogenicity can be induced not only by free PEG molecules but also by PEGylated products. Numerous researchers have observed rapid clearances of a second dose of PEGylated products compared with the first doses when those products were repeatedly injected within one week [17, 19]. This effect is attributed to the formation of anti-PEG antibodies upon administration of a first dose of PEGylated products [109]. Immunogenicity of PEGylated

products was first described by Richter and Akerblom in 1983, when they found that anti-PEG antibodies could develop in rabbits following the intramuscular or subcutaneous administration of various PEG-modified proteins in Complete Freund's Adjuvant [110]. Our research group previously reported that intravenous administration of PEGylated liposome induces anti-PEG IgM production in both rats and mice [111]. We found that PEGylated liposomes act as T-cell independent antigens during anti-PEG antibody production [103]. We also observed that intravenous administration of PEGylated ovalbumin (OVA) or PEGylated bovine serum albumin (BSA) elicited anti-PEG antibodies similar to PEGylated liposomes [112]. Recently, several reports have described the effect of pre-existing or induced anti-PEG antibodies in humans and the effect this exerts on the therapeutic efficacy of PEGylated products in patients [113-116].

2.5. Accelerated blood clearance (ABC) of PEGylated products

The ABC phenomenon was introduced by Dams *et al.* in 2000. With ABC the first dose of PEGylated liposomes injected into rhesus monkeys or rats led to the enhanced clearance of a second dose of PEGylated liposomes injected within one week [19]. This phenomenon may limit the use of different types of PEGylated products in the future because the treatment with PEGylated products, which would induce anti-PEG antibodies, might affect the clearance as well as the therapeutic efficacy of subsequently administered PEGylated products.

Our research group has reported that an intravenous (i.v.) injection of PEGylated liposomes enhances the clearance of a second dose injected a few days later [20]. We gave a tentative explanation for this ABC phenomenon (**Figure 3**), where the initial dosage of PEGylated liposomes primes the immune system to produce anti-PEG IgM, which selectively interacts with PEG molecules in the second dose of PEGylated liposomes, and results in a complement activation and increased engulfment of the second dose of PEGylated liposomes by Kupffer cells in the liver [117, 118]. We recently reported that i.v. injection of PEGylated lipid nanoparticles (LNP) induces the production of anti-PEG IgM, which then triggers the ABC phenomenon [119]. We also reported that the anti-PEG IgM induced either by i.v. injection of PEGylated OVA or by the subcutaneous (s.c.) administration of Pegasys® effectively enhances the rapid clearance of subsequent Pegasys® doses [21]. We also found a similar phenomenon with repeated doses of pegfilgrastim (PEG-G-CSF), a clinically approved treatment for neutropenia [120]. Similarly, we found that i.v. injection of PEGylated exosomes induces the

production of anti-PEG IgM, which enhances the clearance of a second dosage of PEGylated exosomes or PEGylated liposomes administered via i.v. a few days after the initial dose [121].

Furthermore, anti-PEG antibodies may compromise the therapeutic efficacy of PEGylated therapeutics and/or develop undesirable adverse drug reactions. Pre-existing anti-PEG antibodies are known to enhance the clearance of an initial dose of PEGylated products, which may negatively affect the therapeutic efficacy of these drugs. We previously reported that pre-existing anti-PEG antibodies affect the *in vivo* fate of PEGylated liposomes. We found that pre-existing anti-PEG IgM induced via the intraperitoneal inoculation of anti-PEG IgM-producing hybridoma cells (HIK-M09 and HIK-M11) decreases the tumor accumulation level of subsequently administered PEGylated liposomes and accelerates liposome clearance by enhancing its accumulation in the liver and spleen [122]. Similarly, Hsieh *et al.* reported that pre-existing anti-PEG antibodies alter the pharmacokinetics and decrease the tumor accumulation and therapeutic efficacy of LipoDox. They reported that the therapeutic efficacy of LipoDox was significantly diminished in a mouse model bearing anti-PEG antibodies compared with a naïve model [104]. Other research groups have reported that these antibodies may be a potential source of hypersensitive reactions following the administration of PEGylated therapeutics [13, 17, 123].

3. Toxicity of PEG in PEGylated products

Although PEG and its derivatives are considered inert and almost non-toxic molecules, some safety-related problems of free PEG or PEG conjugated with nanoparticles and proteins with different molecular weights have been noticed by some researchers. Smyth *et al.* reported a case of chronic oral toxicity in rats upon oral administration of PEG oligomer with a low molecular weight [124]. In the same manner, undesirable toxicity has been reported in monkeys [125]. Thiele *et al.* have been reported that mice intraperitoneally injected with PEG 200 at a dose of 8 mL/kg did not tolerate PEG 200 well, and half of the animals had to be euthanized. The results demonstrate that although PEG 200 is generally considered to be harmless, it can be toxic when it is intraperitonially injected and is painful for the recipient mice [126]. In the same manner, Liu *et al.* have reported that PEG-based monomers including poly (ethylene glycol) methyl ether acrylate (mPEGA) and poly (ethylene glycol) methyl ether methacrylate (mPEGMA) showed obvious cytotoxicity. They reported that PEG-400 and PEG-2000 seem to be non-cytotoxic in their research. PEG-1000, PEG-4000, and mPEGMA-950 showed moderate cytotoxicity, especially at high concentrations. Triethylene glycol (TEG) and mPEGMA-500

showed significant cytotoxicity, and mPEGA-480 showed acute cytotoxicity [127]. Shiraishi *et al.* have been studied the toxicity associated with the use of polymeric micelles composed of poly (ethylene glycol)-*b*-poly(aspartate) block copolymers using Donryu strain rats. They reported that intravenous injection five times with either a low dose (20 mg/kg) or a high dose (200 mg/kg) leads to an increase in the number of foamy cells in the lungs and lymph nodes in micelle-injected rats at the low dose. At the high dose, they observed a significant increase in the number of foamy cells in the spleen. Also, they observed a marked increase in the CD68-positive macrophages in the spleen, liver, and lungs of treated rats, which may confirm the toxicity of PEGylated polymeric micelles [128]. On the other hand, Turecek *et al.* studied the toxicological effects of PEGylated proteins and reported that there was a significant cellular vacuolation was observed in 5 of the 11 approved PEG-protein conjugates and 10 of the 17 PEG-protein conjugates, which may also represent another sign of toxicity of PEGylated products [40].

4. Hypersensitivity reaction (HSR) to PEG or PEGylated products

4.1. Background

Hypersensitivity is a group of undesirable reactions caused by the immune system. The severity of these reactions' ranges from mild to life-threatening. Hypersensitivity is classified into four types (I-IV) according to the onset and the immunological mechanism involved in these reactions [129]. Type I hypersensitivity is also referred to as an immediate type and is mediated by IgE specific for allergens. This is a mast cell-mediated hypersensitivity with examples that include asthma, urticaria, allergic rhinitis, and angioedema. It is noteworthy that some disorders occur via IgE-independent and non-specific activation of mast cells, which are considered subtypes of type I hypersensitivity such as systemic reactions to iodinated contrast reagents, some biological drugs, and opiates [130]. Type II hypersensitivity refers to an antibody-mediated cytotoxic reaction where IgG and IgM antibodies bind to allergens and help eliminate them via different mechanisms. Type II hypersensitivity is further classified into type IIa, which are antibody-mediated cytotoxic reactions characterized by the cytolytic destruction of targeted cells. Type IIb refers to antibody-mediated cell-stimulating reactions such as Graves' disease and chronic idiopathic urticaria [131]. Type III hypersensitivity is an immune complex-mediated reaction where IgG and IgM antibodies bind to antigens and form immune complexes. These complexes activate a complement system, which after several cascades ends with engulfment

and damage of antigens. Type IV hypersensitivity is characterized by delayed reactions, and T cells are the main effector cells in this type. Type IV hypersensitivity is further classified into 4 types as follows. Type IVa is characterized by Th1 cell-mediated reactions and macrophage activation as seen in type 1 diabetes and contact dermatitis. Type IVb refers to Th2 cell-mediated reactions with eosinophilic inflammation such as persistent asthma and allergic rhinitis. Type IVc is made up of cytotoxic T cell-mediated diseases such as Stevens-Johnson syndrome. Type IVd is T-cell-mediated neutrophilic inflammation such as that seen in acute generalized exanthemata's pustulosis and Behcet disease [132].

The administration of PEGylated nanocarriers can interact with the immune system and result in undesirable HSRs. These reactions are also known as CARPA or infusion reactions [133-135]. CARPA is classified as a non-IgE-mediated allergy because it occurs after a single exposure to PEGylated nanocarriers with no past history of exposure to PEGylated nanostructures. This is opposed to type I hypersensitivity, which requires prior allergen sensitization [136]. Symptoms associated with PEG hypersensitivity are characterized by a rapid onset with different degrees of severity. Common manifestations are pruritus, flushing, angioedema, hypotension, and even bronchospasm, which may lead to respiratory failure and death [38]. The role of complement activation in HSRs was first reported in the 1970s by Lang *et al.*, where complement activation was considered to be a marker of allergies caused by radiocontrast media [137]. This role of the complement system in HSRs was verified by Szebeni *et al.* in 2004 [138]. The factors that affect the incidence and the severity of HSRs will be discussed later.

4.2. HSRs to free PEG

Despite the notion that PEG is a biologically inert polymer, there have been reports of a relationship between PEG and the occurrence of HSRs. Bordere *et al.* reported a case of anaphylaxis, which is a life-threatening allergic reaction, caused by PEG 3,350, with symptoms of itching, erythema, and hypotension following the intra-articular administration of Depo-Medrol Lidocaine[®], which contains methylprednisolone acetate as an active constituent in addition to PEG 3,350 and other excipients [139]. In addition, Gachoka *et al.* reported symptoms of allergic reactions such as urticaria, angioedema, and anaphylaxis after the administration of a barium enema containing PEG to empty the bowels before an X-ray examination of the colon [140].

Hyry *et al.* reported two cases of anaphylaxis after administration of medications containing macrogol (PEG 6,000), namely V-Pen MEGA® tablets (for tonsillitis treatment) and Fludent® lozenges (for caries prevention). The reported cases had shown short episodes of urticaria, dizziness, and tachycardia within minutes after receiving the medications. Positive skin prick tests to PEG 6,000 in the reported cases confirmed the potential role of PEG 6,000 in HSRs [141]. Reported cases of free PEG-associated hypersensetivity are summarized in **Table 3.**

4.3. HSRs to PEGylated products

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

Several reports have elucidated the role of the interaction between PEGylated pharmaceuticals (PEGylated proteins, PEGylated liposomes, and PEGylated lipid nanoparticles) and the immune system and the development of hypersensitivity reactions, which is the so-called CARPA, or infusion reaction. This type of hypersensitivity is classified as a non-IgE-mediated pseudoallergy and is initiated by activating the complement system [136, 142-144]. CARPA mainly affects the cardiopulmonary system with various symptoms such as arrythmia, angioedema, bronchospasm, hyperventilation, cardiogenic shock, and myocardial infarction [133]. PEGylation is a commonly used approach to enhance the stability of therapeutic proteins, enzymes and aptamers. A typical example of CARPA is JIVI® (Factor VIII PEGylated protein used in the treatment of Haemophilia A patients) where reports have stated that patients treated with JIVI® for severe haemophilia A developed anti-PEG antibodies and experienced hypersensitivity reactions [145, 146]. Also, anti-PEG antibody-mediated infusion reactions have been reported in the treatment of gout using Krystexxa® (Pegloticase) [147]. In addition, 8.7-23.5% of patients treated with Oncaspar® (pegaspargase) have developed HSRs due to the induction of anti-PEG antibodies [148, 149]. A previous study reported that about 96% of 261 patients with phenylketonuria developed anti-PEG antibodies and experienced hypersensitivity reactions after receiving subcutaneous injections of pegvaliase [150]. In the same manner, severe immediate allergic reactions were reported following treatment with Pegnivacogin (Modified 31nucleotide RNA aptamer). Pegnivacogin is a PEGylated aptamer prepared by conjugation with 40-kD branched PEG polymer and is used to inhibit factor IXa in coronary artery disease patients. Povsic et al. reported that acute allergic reactions to Pegnivacogin occurred in patients with pre-existing anti-PEG antibodies, which might be associated with complement activation due to the interaction between anti-PEG antibodies and PEG in PEGylated aptamer [151].

It is noteworthy that not only PEGylated proteins, but also PEGylated nanoparticles are known to cause hypersensitivity reactions. As many as 45% of cancer patients are known to have developed HSRs upon receiving Doxil® without premedication with antihistaminic and steroids, but this percentage was decreased to between 4.0 and 7.1% in patients premedicated with antihistaminic [152-154]. Likewise, PEGylated liposomes encapsulating oligonucleotides or plasmid DNA generate anti-PEG antibodies in mice, which has led to severe hypersensitivity reactions including facial puffing, vasodilatation, and anaphylactic shock following a second dose of liposomes [155, 156]. Moreover, infusion-related reactions (IRRs) have also been reported following treatment with Onpattro®, which is an siRNA drug (Patisiran) encapsulated within PEGylated LNP. Onpattro® received approval in 2018 for the treatment of hereditary transthyretin-mediated (hATTR) amyloidosis. Its long-term safety studies demonstrated a high incidence of flushing and IRRs (22% each) [157]. Reported cases of PEGylated products-associated hypersensetivity are summarized in **Table 4**.

Recently, Pfizer/BioNTech BNT162b2 and Moderna mRNA-1273 vaccines were approved for the prevention of SARS-CoV-2 infection. Both are mRNA-based vaccines encoding SARS-CoV-2 spike protein delivered by PEGylated LNP (a full list of the components of both vaccines appears in Table 2) [158-160]. On December 8th 2020, the National Health System (NHS) in UK started a vaccination campaign for high-risk people. One day later, the healthcare workers reported two cases of allergic reactions after vaccine administration. As a result, the UK's Medicines and Healthcare Products Regulatory Agency (MHRA) advised healthcare workers not to provide BNT162b2 vaccines to anyone who had a history of allergic reactions [161]. In North America, the Vaccine Adverse Event Reporting System (VAERS) created by the Centers for Disease Control (CDC) reported severe allergic reactions in 6 cases out of 272,001 vaccinations up to December 19th 2020 [162], which increased to 3,942 cases of allergic reactions among 483,847,837 doses (of both BNT162b2 and mRNA-1273) in the US by December 24th 2021 [163]. This represents one case of allergic reactions in every 122,742 doses, which is about 8.15 times higher than the expected rate of one case per million. Interestingly, there are numerous reports regarding allergic reactions to COVID-19 mRNA vaccines, and the majority of them link allergic reactions to the presence of PEG in vaccine components. Wolfson et al. reported that among 65 patients with immediate allergic reactions to a first dose of mRNA vaccines, a total of 14 patients had a positive skin test for PEG (5 cases) and/or polysorbate 80 (12 cases) [164]. Another case reported by Sellaturay *et al.* showed a 52-year-old woman who suffered from an immediate severe allergic reaction to Pfizer/BioNTech COVID-19 vaccine. The woman had a history of allergic reactions to a PEG-containing medication in addition to some cosmetic products. Surprisingly, she developed systemic anaphylaxis after only a skin prick test with 1% PEG 4,000. PEG allergy was confirmed as the reason for the woman's allergic reaction to the Pfizer/BioNTech COVID-19 vaccine [165].

Furthermore, a search of the VAERS database for incidences of allergic reactions to COVID-19 mRNA vaccines BNT162b2 and mRNA-1273 revealed 3,942 cases where symptoms appeared within one day following injection. After further refinement of the results to identify the cases related to PEG allergy, 25 cases with either a positive PEG/polysorbate allergy test or a previous history of PEG/polysorbate allergy were identified (cases are summarized in **Table 5**). The reported symptoms varied from hives to severe life-threatening anaphylactic reactions.

4.4. Mechanism of PEG-associated hypersensitivity

Although the exact mechanism of PEG-induced hypersensitivity has not been fully elucidated, a growing body of evidence suggests that complement activation plays a role in the development of HSRs. Complement activation plays a vital role in the innate immune defense mechanism against foreign antigens [166]. Complement activation is normally controlled by a collection of cell-surface proteins to prevent auto harm to normal tissues. If, however, the complement is hyperactive, as it is in autoimmune diseases, it can cause serious damage to a variety of organs [167].

In this model, the complement system contributes to the occurrence of HSRs by releasing anaphylatoxins C3a and C5a in response to complement activation via the three known activation pathways: classical, alternative and lectin. The released anaphylatoxins bind to their receptors C3aR and C5aR, respectively, causing inflammatory cells such as macrophages, basophiles, and mast cells to become activated. Activated inflammatory cells secrete a group of inflammatory mediators as histamine, leukotrienes, platelet activating factor (PAF), and tryptase. The cardiopulmonary symptoms associated with HSRs generated by PEGylated products are caused by the action of these mediators on their specific receptors [133, 168]. Recent results have highlighted the role of anti-PEG antibodies in PEG-induced CARPA via the classical pathway in the case of PEGylated liposomes and PEG-G-CSF [120, 134]. **Figure 4** represents a simple demonstration for the mechanism of PEG-induced anaphylactic reactions.

Hugli *et al.* investigated the function and structure of anaphylatoxins, concluding that C3a,

2 C4a, and C5a are genetically related and are the key regulators of cardiopulmonary function.

They also found that complement activation and the resultant HSRs are linked to the

4 overexpression of anaphylatoxins, particularly C3a and C5a [169].

The double-hit theory is another hypothesis that explains the mechanism of PEG-induced HSRs mainly in the case of pre-existing anti-PEG IgMs in circulation. Anaphylactic reactions, according to this theory, are caused by two hits on immune modulatory cells such as mast cells, basophils, and macrophages; the first is an anaphylatoxin signal, and the second is a direct engagement of drugs or particles with these cells via surface receptors. Interaction with these receptors stimulates a signal transduction network that mediates the secretory response [133]. Binding of the secreted anaphylatoxins to their specific receptors on mast cells or basophiles causes a release of vasoactive inflammatory mediators, which are responsible for HSR symptoms. PEG is believed to act on the same hypothesis, in which PEG on the surface of PEGylated nanocarriers binds to macrophages and mast cells via specific surface receptors to stimulate a secretory response [13, 170].

4.5. Factors affecting PEG-associated hypersensitivity

Several factors could affect the incidence and the severity of PEG hypersensitivity; these are summarized in the following section.

4.5.1. Factors affecting free PEG-induced hypersensitivity

4.5.1.1. PEG molecular weight

Because of the diversity in polymer properties that occurs when the molecular weight of the polymer changes, a wide range of PEG molecular weights are commercially available for use in pharmaceutical formulations, cosmetics, or as food additives. PEG molecular weight is known to have a significant effect on the onset of the severity of the HSRs. Shah *et al.* reported that PEGs with a lower molecular weight can permeate the skin and mucosa more effectively than those of a larger molecular weight, which increases the risk of sensitization. They also reported that PEGs with a high molecular weight can trigger HSRs at low concentrations upon sensitization by comparison with low molecular weight PEGs [171].

Among the various molecular weights (from 300 to 20,000 Da), PEGs with molecular weights of 3,350 and 4,000 Da make up the majority of reported cases of HSRs [38]. Stone *et al.* showed that the serum of patients who reacted clinically to PEG 3,350, was more reactive

towards PEG with a higher molecular weight, indicating that a high molecular weight of PEG is an important factor in PEG-related HSRs. Although PEGs with a high molecular weight are frequently associated with positive PEG skin prick tests (SPT) [172, 173], both low and high molecular weight PEGs can induce HSRs [171, 174].

4.5.1.2. Reaction threshold dose

Not only the molecular weight of PEG but also the dosage of PEG may play a crucial role in the development of PEG hypersensitivity. To investigate the effect of PEG dosage on the development of PEG allergies, Sohy *et al.* used different concentrations of oral PEG 4,000 (starting from 1 mg and increasing the dose every 30 minutes). The results showed that a positive allergic response was observed 30 minutes following the administration of 7.1 mg of PEG 4,000, while lower dosages showed no significant allergic responses [175]. According to Bommarito *et al.*, PEG 4,000 and PEG 6,000 at low concentrations (0.0001%) showed a positive response in the basophile activation test, whereas PEG 400 at various concentrations showed no response [172]. These findings point to the need for a particular test for various PEGs in order to investigate the possibility of PEG hypersensitivity. Furthermore, PEGs with different molecular weights, particularly the lower molecular weight versions, gave false SPT results since the measured dose was insufficient to surpass the patient's reactivity-threshold dose. As a result, each patient should be individually checked against the dose and molecular weight.

4.5.2. Factors affecting PEGylated product-induced hypersensitivity

4.5.2.1. Morphological properties of PEGylated nanocarriers

PEGs are commonly used for surface decoration of nanocarriers such as liposomes, nanoparticles, and exosomes to improve the circulation half-life and stability. The external shape of nanostructures is reported to have a significant impact on complement activation and on the development of PEG allergic reactions. Nanostructures with irregular, oval, or elongated external surfaces generally activate C5 convertases, which are responsible for complement activation [176].

Pedersen *et al.* have reported that the structure curvature has a significant impact on the binding of human IgM antibodies to antigen surfaces. They demonstrated that the presence of curvature in peptidoglycan (PGN) fragments allows for an efficient interaction with IgM, which results in activation of a strong classical pathway complement [177]. According to Szebeni *et al.*, the reactogenicity and increase in the SC5b-9 formation after Doxil® administration compared

with that of free doxorubicin may be attributed to the surface modification of PEGylated liposomes. This reactogenicity may be due to the presence of elongated crystals of doxorubicin and/or to the irregular liposome surface, which causes an ovaliform transition of spherical vesicles followed by an increase in the ratio of flat surfaces to curved areas, and this results in a build-up of multimolecular complexes and complement activation [176].

4.5.2.2. Surface charge and composition of PEGylated nanocarriers

The surface charge of PEGylated products is known to have a significant impact on the extent of complement activation and HSR development. Cationic nanocarriers increase complement activation *in vitro* in human serum analysis, which could be attributed to the efficient binding ability of positively charged molecules with serum and plasma proteins [178]. Gao *et al.* reported that negatively charged plasma proteins such as opsonin protein, which is responsible for exogenous molecule opsonization and then phagocytic engulfment, can efficiently bind positively charged nanoparticles [179]. Similarly, Yallapu *et al.* reported that the interaction of curcumin nanoformulations with plasma proteins is primarily influenced by the surface charge of the nanoformulations with cationic ones enhancing binding with negatively charged physiological membranes and plasma proteins [180]. In addition, large multilamellar vesicles prepared with negatively charged phospholipids are known to have a greater vasoactive effect than those prepared with neutral phospholipids, indicating that charged vesicles can stimulate the immune system via complement activation to a greater extent than uncharged vesicles [181].

Furthermore, the composition of PEGylated formulations has a significant impact on HSR induction. According to Baranyi *et al.*, i.v. injection of multilamellar vesicles with high cholesterol content (71%) can cause pulmonary and myocardial manifestations as a result of complement activation [182]. Szebeni *et al.* reported that the pulmonary hypertensive effect of liposomal administration is directly proportional to cholesterol content [183]. Excess cholesterol content can aggregate and accumulate on the surface of nanocarriers, which makes them available to interact with naturally existing anti-cholesterol antibodies in blood circulation [183, 184]. Caracciolo *et al.* used neutral lipids in the preparation of liposomes to investigate the impact on their interaction with plasma proteins. They prepared neutral dioleoyl phosphatidylethanolamine (DOPE)-based liposomes instead of cationic 1,2-dioleoyl-3-trimethylammonium-propane (DOTAP)-based liposomes. They reported that positively charged

- 1 cationic liposomes interact substantially with negatively charged plasma proteins, boosting
- 2 complement activation, whereas neutral lipids have a poor affinity for plasma protein interaction.
- 3 They also discovered that employing high cholesterol levels in liposome synthesis can induce
- 4 interactions of liposomes with complement proteins and immunoglobulins [185].

4.5.2.3. Size and homogeneity of PEGylated formulations

The degree and intensity of complement activation is largely determined by the affinity of antibodies for foreign substances. The binding of anti-PEG, anti-cholesterol, or anti-phospholipid antibodies to PEGylated-lipidic nanocarriers is dependent on their size. This impact can be explained by the fact that increasing the size of an antigen increases the surface area that is accessible for antibody-specific antigen interaction [186]. Highly homogenous unilamellar vesicles are considered highly safe with no vasoactive properties. Increasing the diameters of PEGylated nanovesicles, as in large multilamellar vesicles (LMV), could assure efficient antibody binding, which would trigger higher levels of complement activation and HSRs [187, 188].

Szebeni *et al.* proposed another explanation for the effect of surface area on complement activation, based on the fact that complement activation requires a specific threshold dose of antibodies to be initiated, and that increasing the surface area allows for an optimal arrangement of antibodies on the molecule surface, allowing for a large amount of antibody to bind to the molecule surface, which could be sufficient to initiate complement activation [183]. The same assumption can be extended to PEGylated proteins, since Zang *et al.* proved the importance of particle size in determining the efficacy of anti-PEG antibodies in accelerating the clearance of PEGylated particles [189]. They reported that a particle size greater than 40 nm is required for anti-PEG antibody binding and complement activation, and that the presence of aggregated PEG-uricase increased the particle size of PEG-uricase (around 38 nm) to greater than 60 nm, facilitating antibody binding and the ABC phenomenon. This could also explain why second-dose PEG-OVA and PEG-G-CSF (10 nm) failed to induce the ABC phenomenon in the presence of anti-PEG IgM but not PEGylated liposomes (100 nm) [120].

4.5.2.4. Route and rate of administration

Oral, intravenous (i.v.), intramuscular, intravaginal, and intraarticular administration of PEG-containing products, as well as topical application of PEG-containing products, have all been linked to PEG-induced hypersensitivity [171-173, 190-192]. In terms of the i.v. route, slow

i.v. infusion of PEGylated nanocarriers was found to have a pulmonary hypertensive effect that was lower than that of bolus i.v. injection. The pulmonary hypertensive impact is attributed to an increase in the generation of anaphylatoxins after complement activation [193]. The level of anaphylatoxins in blood is controlled by two main rates: the first is the rate of production (complement activation), and the second is the rate of clearance. Because the rate of clearance is relatively consistent and unaffected by the route of administration, the degree of complement activation is the main contributor to the level of anaphylatoxins. As a result, the route of administration has a significant impact on the level of anaphylatoxins, which is higher in the case of bolus i.v. injection compared with slow i.v. infusion, which allows for substantial complement activation and the generation of anaphylatoxins [13, 194].

The amount of PEG available for absorption affects the development and severity of HSRs when PEG-containing products are applied topically. Low molecular weight PEGs have limited absorption through healthy skin. High molecular weight PEGs (more than 4,000 Da) have difficulty being absorbed through intact skin, which is why PEGs with high molecular weights are favored in cosmetic preparations [4]. On the other hand, the presence of injury or damage in the skin or in the gastrointestinal mucosa allows enough PEGs to be absorbed and subsequently stimulate complement activation. Symptoms of urticaria, pruritus, and oedema have been reported after using oral and topical PEG-containing products in cases with compromised skin or intestinal mucosa [192, 195, 196].

Conclusions

PEGs are widely used synthetic polymers with different molecular weights and different properties. PEGs have promising characteristics such as low toxicity, biocompatibility, and inert nature. Therefore, PEGs have been frequently used in foods, cosmetics and pharmaceutical products as solubility enhancing agents and stabilizing agents. PEGylation can stabilize nanoparticles and protein drugs in vials during storage by preventing their aggregation. PEGylation also improves circulation properties of nanoparticles and protein drugs by preventing adsorption of plasma proteins (opsonisation) and recognition by the cells of MPS. The long circulating effect provided by PEGylation is the main mediator for drug passive targeting in highly perfused tissues such as solid tumors. Despite the widespread usage of PEGs, immunological reactions to PEGs themselves, PEG-containing products and PEGylated products are recognized. The administration of PEGylated products induces production of anti-PEG

antibodies. The presence of pre-existing anti-PEG antibodies, presumably due to extensive use of PEGs in foods and cosmetics, increases the risk of the accelerated blood clearance (ABC) phenomenon, which could lessen the therapeutic efficacy of PEGylated products in clinical settings, as well as, increases the risk of Hypersensitivity reaction (HSR). HSR is currently being reported in many cases following mRNA-based COVID-19 vaccination. Although the mechanism behind HSR induced by mRNA-based COVID-19 vaccines is still uncertain, understanding the mechanism and exchanging the knowledge between the nanomedicine and vaccine field are important since PEGylated lipid nanoparticles are used for all mRNA-based COVID-19 vaccines as delivery vehicles. With widespread use of PEG household items and PEGylated therapeutics, an increase in the incidence of HSR is predictable. The role of PEGs in induction of anti-PEG antibodies and PEG-induced HSRs and the mechanism behind these immunological reactions should be further elucidated to unearth more facts, attaining more effective preventive measures.

Conflicts of interest

There is no conflict of interest for any of the authors.

Acknowledgement

M.I. and E.R. are funded by scholarships from the Ministry of Higher Education of the Arab Republic of Egypt. S.E. is funded by JSPS International Fellowships for Research in Japan. This study was in part supported by a Grant-in-Aid for JSPS Research Fellow (20F20411), a Grant-in-Aid for Fostering Joint International Research (B) (19KK0279) and a Grant-in-Aid for Transformative Research Areas (A) (Publicly Offered Research) (21H05526) from the Japan Society for the Promotion of Science, by the Egyptian Government represented in the Cultural Affairs and Missions Sectors (Ministry of High Education), by the Nagai Foundation Tokyo, by the KOSÉ Cosmetology Research Foundation, and by a research program for the development of an intelligent Tokushima artificial exosome (iTEX) from Tokushima University.

Table 2. Components of BNT162b2 and mRNA-1273 vaccines

Vaccine	mRNA	Lipids				Other additives	Ref.
	encoded	Ionizable lipid	Helper lipid	Cholesterol	PEG-lipid		
Pfizer/Bio	Nucleoside-	ALC-0315 =	DSPC = 1,2-	Cholesterol	ALC-0159 =	Dibasic sodium	[197]
NTech	modified	((4-	Distearoyl-	(plant	2-	phosphate	
BNT162b2	mRNA	hydroxybutyl)	sn-glycero-3-	derived)	[(polyethylen	dihydrate	
	encoding	azanediyl)bis(phosphocholi		e glycol)-	Monobasic	
	SARS-CoV-	hexane-6,1-	ne		2000]-N,N-	potassium	
	2 spike (S)	diyl)bis(2-			ditetradecyla	phosphate	
	glycoprotein	hexyldecanoat			cetamide	Potassium	
		e)				chloride	
						Sodium chloride	
						Sucrose	
Moderna	Nucleoside-	SM-102 =	DSPC = 1,2-	BotaniChol	PEG2000-	Sodium acetate	[198]
mRNA-	modified	heptadecane-	Distearoyl-	® (non-	DMG = 1,2-	Sucrose	
1273	mRNA	9-yl 8-((2-	sn-glycero-3-	animal	dimyristoyl-	Tromethamine	
	encoding	hydroxyethyl)	phosphocholi	origin	rac-glycerol,	Tromethamine	
	SARS-CoV-	(6-oxo-6-	ne	cholesterol)	methoxypoly	hydrochloride	
	2 spike (S)	(undecyloxy)			ethylene	Acetic acid	
	glycoprotein	hexyl) amino)			glycol		
		octanoate					

 Table 3. Cases with PEG-related allergic reactions to free PEG [31]

PEG-containing product	Active constituent	Type of PEG	Allergic symptoms	Previous history of allergy
- Depo-Provera®	- Medroxyprogesterone	- PEG-3350	Sneezing, rhinorrhoea,	None
	PEG-3350		urticaria, ocular	
			irritation, hypotension,	
			chest tightness,	
			Biphasic urticaria.	
- MoviPrep®	- Osmotic laxative PEG-	- PEG-3350	Urticaria, Pruritus,	None
	3350		urticaria, angioedema,	
			swelling	
			of hands and feet,	
			hypotension, Contact	
			urticaria.	
- Vimovo®	- Esomeprazole,	- PEG-8000	Urticaria, angioedema,	None
	naproxen.		syncope, presyncope.	
			Generalized pruritus.	
- Effervescent	- Effervescent vitamin C:			
vitamin C®	HMW-PEG			
- Klean Prep®	- Osmotic laxative, PEG-	- PEG-3350	Dyspnea, angioedema,	None
	3350		visual	
- Phosphate Sando®	- Effervescent phosphate,	- PEG-4000.	disturbance, syncope,	
	PEG-4000.		presyncope.	
- Motilium	- Domperidone, PEG-400	- PEG-400 and	Angioedema, throat	Chronic
Suppository®	and 1000.	1000.	tightness, paraesthesia,	Spontaneous
- Nurofen®	-Ibuprofen, PEG-6000.	- PEG-6000.	throat tightness.	Urticaria (CSU)
- Betadine®	- Povidone-iodine, PEG-	- PEG-400,	Urticaria, angioedema,	CSU
	400, 6000	6000	presyncope, hypotension,	
- Voltorol Oral®	- Diclofenac PEG-8000	- PEG-8000	respiratory distress,	
			Contact urticaria.	

Table 4. Cases with PEG-related allergic reactions to PEGylated products

Commercial product	PEGylated entity	Type of PEG	Allergic symptoms	Previous history of allergy	Ref.
$JIVI^{ ext{ iny R}}$	Recombinant	30 kDa PEG	Urticaria, angioedema,	None	[145]
	antihemophilic		dyspnea.		
	factor VIII				
Neulasta [®]	Pegfilgrastim	10 kDa PEG	Minimal rash on her arms and	None	[199]
			abdomen as well as a sore		
			throat, pruritis, erythematous,		
			lip swelling		
Cimzia®	Certolizumab pegol	40 kDa mPEG	Erythema, urticarial rash,	Movicol® (PEG	[200]
			dyspnoea, wheeze, and a	3350) allergy	
			sensation of presyncope.		
Oncaspar [®]	Pegaspargase	5 kDa PEG	Transient flushing or rash,	None	[201]
			urticaria, dyspnea,		
			symptomatic bronchospasm,		
			angioedema, hypotension,		
			anaphylaxis.		
Palynziq [®]	Pegvaliase	40 kDa PEG	lip swelling, flushing,	allergic rhinitis	[202]
			dyspnoea.		
Doxil®	PEGylated	2 kDa PEG	Shortness of breath, flushing,	None	[203]
	liposomal		feeling warm		
	Doxorubicin		and dizziness		
Macugen [®]	Pegaptanib	40 kDa mPEG	Tongue oedema, lip swelling,	None	[204]
			prolonged urticarial rash.		
Pegnivacogin [®]	RNA aptamer	40 kDa mPEG	Angioedema, flushing,	None	[205]
			difficulty of breathing.		
Onpattro [®]	Patisiran	PEG ₂₀₀₀ -DMG	Flushing, peripheral oedema,	None	[157]
			muscle spasm, dyspnoea.		

1 Table 5. Cases with PEG-related allergic reactions to mRNA COVID-19 vaccines according to the2 CDC [163]

Vaccine	Anaphylaxis	Symptoms	Onset	Current	Adverse	Current	Allergy test	History of	VAERS
				illness	events prior	medications		allergies	ID
					vaccinations				
Moderna	Yes	Massive bloody	Within 9 h	None.	None.	None.	Polysorbate	Seasonal Trees	1020162-1
		diarrhea, one large					allergy:	avocado	
		vomitus, hives, tongue					Negative		
		swelling, difficult to					PEG allergy:		
		speak and swallow.					positive		
Moderna	No	Tachycardia, tingling,	Within	None.	None.	None.	Polysorbate 80	Acyclovir, tree	1285640-1
		dizziness, hives	minutes				allergy: positive	pollens, grass	
								pollen, and dogs.	
Pfizer/BioN	No	Hives; face flushing	Within 5	None.	None.	None.	BNT162B2	Meat allergy	1405639-1
Tech			minutes				allergy: positive		
							PEG allergy:		
							positive		
							Polysorbate 80		
							allergy: positive		

Moderna	No	Cognitive issues;	Within the	None	None.	None.	Polysorbate 80	Allergic to dogs;	1614213-1
		headaches; sleep	first day				allergy: positive	Drug allergy	
		disturbance; vertigo;						(allergic to	
		fatigue; Red						acyclovir); Food	
		inflammation in vein of						allergy (poly	
		right foot; swollen						sorbate 80);	
		lymph nodes; skin felt						Pollen allergy	
		like crawling; Hands						(allergic to grass	
		tremoring; legs and						pollen.)	
		hands weakness;							
		freezing; Arm was very							
		sore and little achy;							
		rapid heartbeat; Feet							
		tingling; blood pressure							
		elevated; dizziness;							
		strange taste in mouth;							
		foggy							

Pfizer/BioN	Yes	Felt 'impending doom',	Within 15	Reflux	None.	Prevacid	PEG 3350	Allergic reaction	1853854-1
Tech		heart racing, tunnel	minutes				allergy: positive	to sushi	
		vision, chest pressure,							
		throat constriction							
		fatigue, SOB, and							
		nausea, chills, fever.							
Moderna	No	Pressure on the front of	Within 1.5	Fibromyal	Reaction to	Vitamin D	PEG allergy:	Amoxicillin,	1396793-1
		throat, high heart rate,	h	gia and	flu vaccine in	and	positive	penicillin, I'm	
		weakness, insomnia,		pre-	2012.	multivitami		sensitive to	
		stomach pain, arm		diabetic		n.		azithromycin,	
		swelling, headache,						allergic to	
		difficult breathing,						melons	
		numbness and pinching							
		on left side.							
Moderna	No	Itchy arms and legs,	Within	None	None.	Claritin	N/A	PEG, colophony,	0924196-1
		allergic dermatitis	minutes					thimerosal, gold	

Moderna	Yes	Cough, difficult	Within 15	None.	None.	Centrum	PEG allergy:	Penicillin,	1036813-1
		breathing.	minutes			Women's,	positive	Sugars (sucrose),	
						Zyrtec,	Polysorbate	Nickel,	
						Climentadin	allergy: positive	Shellfish, MSG,	
						e, vitamin		Perfumes,	
						B12, and D,		Makeups,	
						Levothyroxi		Cleaning, Tree	
						ne, Cinopril,		nuts	
						Lexapro			
Pfizer/BioN	Yes	Gasping for air; Tongue	Within	Hashimoto	None.	None.	N/A	Amlodipine	1150913-1
Tech		swelling; Swollen lips;	seconds	's disease;				which has PEG	
		Throat closing; tickle in		Undifferen				and Lisinopril	
		the throat; Headache;		tiated					
		Dizzy; Confusion;		connective					
		weak; fatigued; full		tissue					
		tongue affecting		disease					
		talking; tingling in lips							

Moderna	No	Arm swelling, Sense of	Within 2 h	None.	None.	N/A	N/A	Aspirin, codeine,	1709669-1
		taste lost, metallic taste,						penicillins, PEG,	
		Fingertips and throat						and possibly	
		swollen, itching all						more per patient.	
		over.						Patient reported	
								allergy to	
								Benadryl	
								(contains PEG)	
Pfizer/BioN	No	Sharp chest pain,	Within 8 h	None.	None.	None.	N/A	PEG, PCN,	1865357-1
Tech		shortness of breath,						ibuprofen	
		headache, and nausea.							
Pfizer/BioN	Yes	Tingling, tightness in	Within 3 h	None.	None.	5mg	N/A	Hashimoto's,	1139132-1
Tech		throat, tongue swelling,				loratadine		Asthma, Gluten	
		difficulty swallowing,				Ritual		Intolerance,	
		Hyperventilating,				Prenatal		Chronic Hives,	
		difficult breathing.				Vitamin		patient reported	
						1000mg		severe eczema	
						turmeric		after application	
						Probiotic		of eye cream	
								containing PEG	

Pfizer/BioN	No	Throat closing;	Within the	None.	None.	Metoprolol	N/A	Patient	1332760-1
Tech		Shortness of breath;	first day			tartrate;		previously had a	
		Swollen throat;				ativan;		reaction to	
		Dizziness/ Lightheaded;				lexapro		Miralax	
		Wheezing.						(contains PEG)	
Moderna	No	Blurred vision, high	Within the	None.	None.	None.	N/A	Propofol which	1581247-1
		blood pressure,	first day					contains	
		flushing, dizziness,						polysorbate,	
		back of head/neck						Demerol	
		soreness, arthralgia, and							
		fatigue.							

Pfizer/BioN	Yes	Mid-back pain;	Within 5-	None.	None.	Zyrtec;	N/A	Cephalasporins,	1645678-1
Tech		coughing; wheezing;	10 minutes			topamax;		oral Diflucan,	
		SOB; severe pain in				spironolacto		clindamycin,	
		chest; flushing and				ne; vitamin		PEG,	
		itching.				D; benadryl		Polysorbate,	
								Propylene	
								glycol, sulfa,	
								Levaquin The	
								patient had	
								known allergies	
								to polysorbate,	
								polyethylene	
								glycol, and	
								propylene glycol	

Pfizer/BioN	No	Itching; Allergic	Within 20	None.	None.	Benadryl;	N/A	Patient reported	1648915-1
Tech		reactions; Tachycardia;	minutes			claritin		PEG sensitivity	
		Numbness and tingling				[clarithromy			
		in her mouth; Feeling of				cin]; zyrtec			
		congestion in the back				[cetirizine			
		of her throat; was not				hydrochlori			
		able to swallow; Patient				de];			
		cannot talk in complete				prednisone			
		sentences.							
Moderna	Yes	Lip swelling; Tongue	Within 15	None.	None.	Fish oil;	N/A	Atenolol,	1691526-1
		Swelling; Chest	minutes			vitamin D;		Sensitivity to	
		tightness; dizziness;				probiotics		petroleum type	
		irregular heart rate.						products, Egg	
								allergy.	
								Patient reported	
								PEG allergy	

Pfizer/BioN	No	Swelling up the	Within the	None.	None.	None.	PEG allergy:	Patient reported	1823560-1
Tech		injection site; redness;	first day				Negative.	polysorbate	
		lots of soreness; 4x4						allergy	
		discoloration area; The							
		lymph nodes under arm							
		became rock hard and							
		was the size of palm.							
Pfizer/BioN	No	Allergic reaction to the	Within 15	None.	None.	None.	N/A	Sulfonamide	1903830-1
Tech		shot; Rash; Itching	minutes					allergy,	
								influenza	
								vaccine and	
								polysorbate 80	
Moderna	No	Nausea, tachycardia,	Within 15	None.	None.	Zyrtec	Polysorbate	Seasonal Trees	1030771-1
		rash on face and neck,	minutes			Vyvanse	20/80 allergy:	avocado	
		tongue swelling				birth control	positive		
							PEG 3350		
							allergy: positive.		
Pfizer/BioN	No	Hives	Within the	Fever	None.	Multivitami	Polysorbate 80	None	1959413-1
Tech			first day	3days		ns	allergy: positive		
				before					
				vaccinatio					
				n, asthma					

Pfizer/BioN	Yes	Rash and itching	Within 10	None.	Anaphylaxis;	Adderall,	N/A	Latex,	1205076-1
Tech			minutes		age 12; MMR	Lopressor,		pineapple.	
					vaccine.	Seasonique,		Sensitivities to	
					Anaphylaxis;	Ventolin,		PEG and	
					age 29; flu	Flovent,		polysorbate.	
					vaccine.	Seravent,			
						Fioricet			
Pfizer/BioN	No	Abnormal breathing	Within the	Chronic	None.	N/A	N/A	PEG/PG, eggs,	1675204-1
Tech		patterns, lightheaded,	first day	daily				wheat, dairy,	
		cold, clammy,		migraine,				peanut	
		respiratory/acute		hypothyroi					
		distress/breathing		d,					
		difficulty, swelling of		hypertensi					
		entire face and sinuses.		on					
Pfizer/BioN	Yes	Dizziness, tongue	Within the	None.	None.	None.	N/A	Depo-provera,	1749850-1
Tech		swelling and sensation	first day					PEG	
		of throat closing.							

References

- [1] A.A. D'souza, R. Shegokar, Polyethylene glycol (PEG): a versatile polymer for pharmaceutical applications, Expert Opin. Drug Deliv., 13 (2016) 1257-1275. https://doi.org/10.1080/17425247.2016.1182485.
- [2] M. Younes, P. Aggett, F. Aguilar, R. Crebelli, B. Dusemund, M. Filipič, M.J. Frutos, P. Galtier, D. Gott, Refined exposure assessment of polyethylene glycol (E 1521) from its use as a food additive, EFSA Journal, 16 (2018) e05293. https://doi.org/10.2903/j.efsa.2018.5293.
- [3] H.-J. Jang, C.Y. Shin, K.-B. Kim, Safety evaluation of polyethylene glycol (PEG) compounds for cosmetic use, Toxicol. Res., 31 (2015) 105-136. https://doi.org/10.5487/TR.2015.31.2.105.
- [4] C. Fruijtier-Pölloth, Safety assessment on polyethylene glycols (PEGs) and their derivatives as used in cosmetic products, Toxicology, 214 (2005) 1-38. https://doi.org/10.1016/j.tox.2005.06.001.
- [5] D. Hutanu, M.D. Frishberg, L. Guo, C.C. Darie, Recent applications of polyethylene glycols (PEGs) and PEG derivatives, Mod. Chem. Appl, 2 (2014) 1-6. http://dx.doi.org/10.4172/2329-6798.1000132.
- [6] M. Zacchigna, F. Cateni, S. Drioli, G.M. Bonora, Multimeric, multifunctional derivatives of poly (ethylene glycol), Polymers, 3 (2011) 1076-1090. https://doi.org/10.3390/polym3031076.
- [7] S. Ranjita, S. Kamalinder, In-vitro release of paracetamol from suppocire suppositories: role of additives, Malays. J. Pharm. Sci., 8 (2010) 57-71.
- [8] R.G. Strickley, Solubilizing excipients in oral and injectable formulations, Pharm. Res., 21 (2004) 201-230. https://doi.org/10.1023/B:PHAM.0000016235.32639.23.
- [9] A. Abuchowski, J.R. McCoy, N.C. Palczuk, T. van Es, F.F. Davis, Effect of covalent attachment of polyethylene glycol on immunogenicity and circulating life of bovine liver catalase, J. Biol. Chem., 252 (1977) 3582-3586. https://doi.org/10.1016/S0021-9258(17)40292-4.
- [10] J.M. Harris, R.B. Chess, Effect of pegylation on pharmaceuticals, Nat. Rev. Drug Discov, 2 (2003) 214-221. https://doi.org/10.1038/nrd1033.
- [11] N.E. Elsadek, A.S.A. Lila, T. Ishida, Immunological responses to PEGylated proteins: anti-PEG antibodies, in: Polymer-Protein Conjugates, Elsevier, 2020, pp. 103-123.https://doi.org/10.1016/B978-0-444-64081-9.00005-X.
- [12] R. P Garay, J. P Labaune, Immunogenicity of polyethylene glycol (PEG), Open Conf. Proc. J., 2 (2011) 104-107. https://doi.org/10.2174/2210289201102010104.
- [13] M. Mohamed, A.S. Abu Lila, T. Shimizu, E. Alaaeldin, A. Hussein, H.A. Sarhan, J. Szebeni, T. Ishida, PEGylated liposomes: immunological responses, Sci. Technol. Adv. Mater., 20 (2019) 710-724. https://doi.org/10.1080/14686996.2019.1627174.
- [14] R.P. Garay, R. El-Gewely, J.K. Armstrong, G. Garratty, P. Richette, Antibodies against polyethylene glycol in healthy subjects and in patients treated with PEG-conjugated agents, Expert Opin. Drug Deliv., 9 (2012) 1319-1323. https://doi.org/10.1517/17425247.2012.720969.
- [15] H. Schellekens, W.E. Hennink, V. Brinks, The immunogenicity of polyethylene glycol: facts and fiction, Pharm. Res., 30 (2013) 1729-1734. https://doi.org/10.1007/s11095-013-1067-7.
- [16] N. d'Avanzo, C. Celia, A. Barone, M. Carafa, L. Di Marzio, H.A. Santos, M. Fresta, Immunogenicity of polyethylene glycol based nanomedicines: mechanisms, clinical implications and systematic approach, Adv. Ther., 3 (2020) 1900170. https://doi.org/10.1002/adtp.201900170.
- [17] A.S.A. Lila, H. Kiwada, T. Ishida, The accelerated blood clearance (ABC) phenomenon: clinical challenge and approaches to manage, J. Control. Release, 172 (2013) 38-47. https://doi.org/10.1016/j.jconrel.2013.07.026.
- [18] T. Ishida, H. Kiwada, Accelerated blood clearance (ABC) phenomenon upon repeated injection of PEGylated liposomes, Int. J. Pharm., 354 (2008) 56-62. https://doi.org/10.1016/j.ijpharm.2007.11.005.
- [19] E.T. Dams, P. Laverman, W.J. Oyen, G. Storm, G.L. Scherphof, J.W. Van der Meer, F.H. Corstens, O.C. Boerman, Accelerated blood clearance and altered biodistribution of repeated injections of sterically stabilized liposomes, J. Pharmacol. Exp. Ther., 292 (2000) 1071-1079.

- [20] T. Ishida, K. Atobe, X. Wang, H. Kiwada, Accelerated blood clearance of PEGylated liposomes upon repeated injections: effect of doxorubicin-encapsulation and high-dose first injection, J. Control. Release, 115 (2006) 251-258. https://doi.org/10.1016/j.jconrel.2006.08.017.
- [21] N.E. Elsadek, E. Hondo, T. Shimizu, H. Takata, A.S. Abu Lila, S.E. Emam, H. Ando, Y. Ishima, T. Ishida, Impact of Pre-Existing or Induced Anti-PEG IgM on the Pharmacokinetics of Peginterferon Alfa-2a (Pegasys) in Mice, Mol. Pharm., 17 (2020) 2964-2970. https://doi.org/10.1021/acs.molpharmaceut.0c00366.
- [22] J. Szebeni, L. Baranyi, S. Savay, H.U. Lutz, E. Jelezarova, R. Bunger, C.R. Alving, The role of complement activation in hypersensitivity to pegylated liposomal doxorubicin (Doxil®), J. Liposome Res., 10 (2000) 467-481. https://doi.org/10.3109/08982100009031112.
- [23] A. Verma, K. Chen, C. Bender, N. Gorney, W. Leonard, P. Barnette, PEGylated E. coli asparaginase desensitization: an effective and feasible option for pediatric patients with acute lymphoblastic leukemia who have developed hypersensitivity to pegaspargase in the absence of asparaginase Erwinia chrysanthemi availability, Pediatr. Hematol. Oncol., 36 (2019) 277-286. https://doi.org/10.1080/08880018.2019.1634778.
- [24] L.K. Beaupin, B. Bostrom, M.J. Barth, I. Franklin, R. Jaeger, P. Kamath, B. Schreiber, A. Bleyer, Pegaspargase hypersensitivity reactions: intravenous infusion versus intramuscular injection—a review, J. Leuk. Lymphoma, 58 (2017) 766-772. https://doi.org/10.1080/10428194.2016.1218004.
- [25] R.E. Rau, Z. Dreyer, M.R. Choi, W. Liang, R. Skowronski, K.P. Allamneni, M. Devidas, E.A. Raetz, P.C. Adamson, S.M. Blaney, Outcome of pediatric patients with acute lymphoblastic leukemia/lymphoblastic lymphoma with hypersensitivity to pegaspargase treated with PEGylated Erwinia asparaginase, pegcrisantaspase: a report from the Children's Oncology Group, Pediatr. Blood Cancer, 65 (2018) e26873. https://doi.org/10.1002/pbc.26873.
- [26] S.J. Thomas, E.D. Moreira Jr, N. Kitchin, J. Absalon, A. Gurtman, S. Lockhart, J.L. Perez, G. Pérez Marc, F.P. Polack, C. Zerbini, Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine through 6 months, N. Engl. J. Med., 385 (2021) 1761-1773. https://doi.org/10.1056/NEJMoa2110345.
- [27] S. Meo, I. Bukhari, J. Akram, A. Meo, D.C. Klonoff, COVID-19 vaccines: comparison of biological, pharmacological characteristics and adverse effects of Pfizer/BioNTech and Moderna Vaccines, Eur. Rev. Med. Pharmacol. Sci., 25 (2021) 1663-1669. https://doi.org/10.26355/eurrev 202102 24877
- [28] Z. Chagla, The BNT162b2 (BioNTech/Pfizer) vaccine had 95% efficacy against COVID-19≥ 7 days after the 2nd dose, 174 (2021) JC15.
- [29] E. Mahase, Covid-19: Pfizer vaccine efficacy was 52% after first dose and 95% after second dose, paper shows, in, British Medical Journal Publishing Group, 2020,
- [30] Y. Ling, J. Zhong, J. Luo, Safety and effectiveness of SARS CoV 2 vaccines: A systematic review and meta analysis, J. Med. Virol., 93 (2021) 6486-6495. https://doi.org/10.1002/jmv.27203.
- [31] F. Cox, K. Khalib, N. Conlon, PEG that reaction: A case series of allergy to polyethylene glycol, J. Clin. Pharmacol., 61 (2021) 832-835. https://doi.org/10.1002/jcph.1824.
- [32] T.T. Shimabukuro, M. Cole, J.R. Su, Reports of anaphylaxis after receipt of mRNA COVID-19 vaccines in the US—December 14, 2020-January 18, 2021, JAMA, 325 (2021) 1101-1102. https://doi.org/10.1001/jama.2021.1967.
- [33] C. COVID, R. Team, Allergic reactions including anaphylaxis after receipt of the first dose of Moderna COVID-19 Vaccine—United States, December 21, 2020—January 10, 2021, Morb. Mortal. Wkly. Rep., 70 (2021) 125-129. https://doi.org/10.15585/mmwr.mm7004e1.
- [34] P. Sellaturay, S. Nasser, S. Islam, P. Gurugama, P.W. Ewan, Polyethylene glycol (PEG) is a cause of anaphylaxis to the Pfizer/BioNTech mRNA COVID 19 vaccine, Clin. Exp. Allergy, 51 (2021) 861-863. https://doi.org/10.1111/cea.13874
- [35] P. Sellaturay, S. Nasser, S. Islam, P. Gurugama, P.W. Ewan, Polyethylene glycol (PEG) is a cause of anaphylaxis to the Pfizer/BioNTech mRNA COVID-19 vaccine, Clin. Exp. Allergy, 51 (2021) 861-863. https://doi.org/10.1111/cea.13874

- [36] L.H. Garvey, S. Nasser, Anaphylaxis to the first COVID-19 vaccine: is polyethylene glycol (PEG) the culprit?, Br. J. Anaesth, 126 (2021) e106-e108. https://doi.org/10.1016/j.bja.2020.12.020.
- [37] B. Cabanillas, C. Akdis, N. Novak, Allergic reactions to the first COVID 19 vaccine: a potential role of polyethylene glycol?, Allergy
- 76 (2020) 1617-1618. https://doi.org/10.1111/all.14711.
- [38] E. Wenande, L. Garvey, Immediate type hypersensitivity to polyethylene glycols: a review, Clin. Exp. Allergy, 46 (2016) 907-922. https://doi.org/10.1111/cea.12760.
- [39] T.T. Hoang Thi, E.H. Pilkington, D.H. Nguyen, J.S. Lee, K.D. Park, N.P. Truong, The importance of poly (ethylene glycol) alternatives for overcoming PEG immunogenicity in drug delivery and bioconjugation, Polym. J., 12 (2020) 298. https://doi.org/10.3390/polym12020298.
- [40] P.L. Turecek, M.J. Bossard, F. Schoetens, I.A. Ivens, PEGylation of biopharmaceuticals: a review of chemistry and nonclinical safety information of approved drugs, J. Pharm. Sci., 105 (2016) 460-475. https://doi.org/10.1016/j.xphs.2015.11.015.
- [41] X.-F. XIAO, X.-Q. JIANG, L.-J. ZHOU, Surface modification of poly ethylene glycol to resist nonspecific adsorption of proteins, Chinese J. Anal. Chem., 3 (2013) 445-453. https://doi.org/10.1016/S1872-2040(13)60638-6.
- [42] M.-Y. Bai, S.-Z. Liu, A simple and general method for preparing antibody-PEG-PLGA sub-micron particles using electrospray technique: An in vitro study of targeted delivery of cisplatin to ovarian cancer cells, Colloids Surf. B: Biointerfaces, 117 (2014) 346-353. https://doi.org/10.1016/j.colsurfb.2014.02.051.
- [43] L. Hong, Z. Wang, X. Wei, J. Shi, C. Li, Antibodies against polyethylene glycol in human blood: A literature review, J Pharmacol. Toxicol. Methods, 102 (2020) 106678. https://doi.org/10.1016/j.vascn.2020.106678.
- [44] J.M. Harris, Physics, Laboratory synthesis of polyethylene glycol derivatives, J. Macromol. Sci. Phys., 25 (1985) 325-373. https://doi.org/10.1080/07366578508081960.
- [45] Y. Hu, W.A. Daoud, K.K.L. Cheuk, C.S.K. Lin, Newly developed techniques on polycondensation, ring-opening polymerization and polymer modification: Focus on poly (lactic acid), J. Mater., 9 (2016) 133. https://doi.org/10.3390/ma9030133.
- [46] F. Zia, M.N. Anjum, M.J. Saif, T. Jamil, K. Malik, S. Anjum, I. BiBi, M.A. Zia, Alginate-poly (ethylene) glycol and poly (ethylene) oxide blend materials, in: Algae Based Polymers, Blends, and Composites, Elsevier, 2017, pp. 581-601.https://doi.org/10.1016/B978-0-12-812360-7.00016-1.
- [47] A. Thomas, S.S. Müller, H. Frey, Beyond poly (ethylene glycol): linear polyglycerol as a multifunctional polyether for biomedical and pharmaceutical applications, Biomacromolecules, 15 (2014) 1935-1954. https://doi.org/10.1021/bm5002608.
- [48] D.A. Herold, G.T. Rodeheaver, W.T. Bellamy, L.A. Fitton, D.E. Bruns, R.F. Edlich, Toxicity of topical polyethylene glycol, Toxicol. Appl. Pharmacol., 65 (1982) 329-335. https://doi.org/10.1016/0041-008X(82)90016-3.
- [49] T. Stylianopoulos, M.-Z. Poh, N. Insin, M.G. Bawendi, D. Fukumura, L.L. Munn, R.K. Jain, Diffusion of particles in the extracellular matrix: the effect of repulsive electrostatic interactions, Biophys. J., 99 (2010) 1342-1349. https://doi.org/10.1016/j.bpj.2010.06.016.
- [50] V. Chadwick, S. Phillips, A. Hofmann, Measurements of intestinal permeability using low molecular weight polyethylene glycols (PEG 400): II. Application to normal and abnormal permeability states in man and animals, Gastroenterology, 73 (1977) 247-251. https://doi.org/10.1016/S0016-5085(19)32197-3.
- [51] H.-W. Leung, B. Ballantyne, S.J. Hermansky, S.W. Frantz, Peroral subchronic, chronic toxicity, and pharmacokinetic studies of a 100-Kilodalton polymer of ethylene oxide (Polyox N-10) in the Fischer 344 Rat, Int. J. Toxicol., 19 (2000) 305-312. https://doi.org/10.1080/10915810050178752.

- [52] J.-C. Tsai, L.-C. Shen, H.-M. Sheu, C.-C. Lu, Tape stripping and sodium dodecyl sulfate treatment increase the molecular weight cutoff of polyethylene glycol penetration across murine skin, Arch. Dermatol. Res., 295 (2003) 169-174. https://doi.org/10.1007/s00403-003-0414-7.
- [53] D.A. Herold, K. Keil, D.E. Bruns, Oxidation of polyethylene glycols by alcohol dehydrogenase, Biochem. Pharmacol., 38 (1989) 73-76. https://doi.org/10.1016/0006-2952(89)90151-2.
- [54] M. Beranova, R. Wasserbauer, D. Vanćurová, M. Štifter, J. Očenášková, M. Mara, Effect of cytochrome P-450 inhibition and stimulation on intensity of polyethylene degradation in microsomal fraction of mouse and rat livers, Biomaterials, 11 (1990) 521-524. https://doi.org/10.1016/0142-9612(90)90070-7.
- [55] T. Yamaoka, Y. Tabata, Y. Ikada, Fate of water soluble polymers administered via different routes, J. Pharm. Sci., 84 (1995) 349-354. https://doi.org/10.1002/jps.2600840316.
- [56] P. Caliceti, F.M. Veronese, Pharmacokinetic and biodistribution properties of poly (ethylene glycol)—protein conjugates, Adv. Drug Deliv. Rev., 55 (2003) 1261-1277. https://doi.org/10.1016/S0169-409X(03)00108-X.
- [57] C.E. Mueller, Prodrug approaches for enhancing the bioavailability of drugs with low solubility, Chem. Biodivers., 6 (2009) 2071-2083. https://doi.org/10.1002/cbdv.200900114.
- [58] N. Seedher, S. Bhatia, Solubility enhancement of Cox-2 inhibitors using various solvent systems, AAPS PharmSciTech, 4 (2003) 36-44. https://doi.org/10.1208/pt040333.
- [59] J.-S. Choi, B.-W. Jo, Enhanced paclitaxel bioavailability after oral administration of pegylated paclitaxel prodrug for oral delivery in rats, Int. J. Pharm., 280 (2004) 221-227. https://doi.org/10.1016/j.ijpharm.2004.05.014.
- [60] P. Milla, F. Dosio, L. Cattel, PEGylation of proteins and liposomes: a powerful and flexible strategy to improve the drug delivery, Curr. Drug Metab., 13 (2012) 105-119. https://doi.org/10.2174/138920012798356934.
- [61] M. Swierczewska, K.C. Lee, S. Lee, What is the future of PEGylated therapies?, Expert Opin. Emerg. Drugs, 20 (2015) 531-536. https://doi.org/10.1517/14728214.2015.1113254.
- [62] G. Molineux, Pegylation: engineering improved pharmaceuticals for enhanced therapy, Cancer Treat. Rev., 28 (2002) 13-16. https://doi.org/10.1016/S0305-7372(02)80004-4.
- [63] A. Grigoletto, A. Mero, I. Zanusso, O. Schiavon, G. Pasut, Chemical and Enzymatic Site Specific PEGylation of hGH: The Stability and in vivo Activity of PEG N Terminal hGH and PEG -
- Gln141 hGH Conjugates, Macromol. Biosci., 16 (2016) 50-56. https://doi.org/10.1002/mabi.201500282.
- [64] A. Kolate, D. Baradia, S. Patil, I. Vhora, G. Kore, A. Misra, PEG—a versatile conjugating ligand for drugs and drug delivery systems, J. Control. Release, 192 (2014) 67-81. https://doi.org/10.1016/j.jconrel.2014.06.046.
- [65] A. Gabizon, A.T. Horowitz, D. Goren, D. Tzemach, H. Shmeeda, S. Zalipsky, In vivo fate of folate-targeted polyethylene-glycol liposomes in tumor-bearing mice, Clin. Cancer Res., 9 (2003) 6551-6559.
- [66] E.A. Azzopardi, E.L. Ferguson, D.W. Thomas, The enhanced permeability retention effect: a new paradigm for drug targeting in infection, J. Antimicrob. Chemother., 68 (2013) 257-274. https://doi.org/10.1093/jac/dks379.
- [67] A. Fukuda, K. Tahara, Y. Hane, T. Matsui, S. Sasaoka, H. Hatahira, Y. Motooka, S. Hasegawa, M. Naganuma, J. Abe, S. Nakao, H. Takeuchi, M. Nakamura, Comparison of the adverse event profiles of conventional and liposomal formulations of doxorubicin using the FDA adverse event reporting system, 12 (2017) e0185654. https://doi.org/10.1371/journal.pone.0185654.
- [68] J. Park, P.M. Fong, J. Lu, K.S. Russell, C.J. Booth, W.M. Saltzman, T.M. Fahmy, PEGylated PLGA nanoparticles for the improved delivery of doxorubicin, Nanomed.: Nanotechnol. Biol. Med., 5 (2009) 410-418. https://doi.org/10.1016/j.nano.2009.02.002.
- [69] J.A. O'Shaughnessy, Pegylated liposomal doxorubicin in the treatment of breast cancer, Clin. Breast Cancer, 4 (2003) 318-328. https://doi.org/10.3816/CBC.2003.n.037.

- [70] T. Safra, F. Muggia, S. Jeffers, D. Tsao-Wei, S. Groshen, O. Lyass, R. Henderson, G. Berry, A. Gabizon, Pegylated liposomal doxorubicin (doxil): reduced clinical cardiotoxicity in patients reaching or exceeding cumulative doses of 500 mg/m2, Ann. Oncol., 11 (2000) 1029-1034. https://doi.org/10.1023/A:1008365716693.
- [71] A.E. Nel, L. Mädler, D. Velegol, T. Xia, E.M. Hoek, P. Somasundaran, F. Klaessig, V. Castranova, M. Thompson, Understanding biophysicochemical interactions at the nano-bio interface, Nat. Mater., 8 (2009) 543-557. https://doi.org/10.1038/nmat2442.
- [72] F. Alexis, E. Pridgen, L.K. Molnar, O.C. Farokhzad, Factors affecting the clearance and biodistribution of polymeric nanoparticles, Mol. Pharm., 5 (2008) 505-515. https://doi.org/10.1021/mp800051m.
- [73] R. Firdessa, T.A. Oelschlaeger, H. Moll, Identification of multiple cellular uptake pathways of polystyrene nanoparticles and factors affecting the uptake: relevance for drug delivery systems, Eur. J. Cell Biol., 93 (2014) 323-337. https://doi.org/10.1016/j.ejcb.2014.08.001.
- [74] J.R. Mora, J.T. White, S.L. DeWall, Immunogenicity risk assessment for PEGylated therapeutics, AAPS PharmSciTech, 22 (2020) 35. https://doi.org/10.1208/s12248-020-0420-0.
- [75] N. James, R. Coker, D. Tomlinson, J. Harris, M. Gompels, A. Pinching, J. Stewart, Liposomal doxorubicin (Doxil): an effective new treatment for Kaposi's sarcoma in AIDS, J. Clin. Oncol., 6 (1994) 294-296. https://doi.org/10.1016/S0936-6555(05)80269-9.
- [76] M. Markman, Pegylated liposomal doxorubicin in the treatment of cancers of the breast and ovary, Expert Opin. Pharmacother., 7 (2006) 1469-1474. https://doi.org/10.1517/14656566.7.11.1469.
- [77] S. Foser, A. Schacher, K.A. Weyer, D. Brugger, E. Dietel, S. Marti, T. Schreitmüller, Isolation, structural characterization, and antiviral activity of positional isomers of monopegylated interferon α -2a (PEGASYS), Protein Expr. Purif., 30 (2003) 78-87. https://doi.org/10.1016/S1046-5928(03)00055-X.
- [78] D.H. Bunka, O. Platonova, P.G. Stockley, Development of aptamer therapeutics, Curr. Opin. Pharmacol., 10 (2010) 557-562. https://doi.org/10.1016/j.coph.2010.06.009.
- [79] M. Vellard, The enzyme as drug: application of enzymes as pharmaceuticals, Curr. Opin. Biotechn., 14 (2003) 444-450. https://doi.org/10.1016/S0958-1669(03)00092-2.
- [80] Y.C. Barenholz, Doxil®—the first FDA-approved nano-drug: lessons learned, J. Control. Release, 160 (2012) 117-134. https://doi.org/10.1016/j.jconrel.2012.03.020.
- [81] R.M. Bukowski, C. Tendler, D. Cutler, E. Rose, M.M. Laughlin, P. Statkevich, Treating cancer with PEG Intron: pharmacokinetic profile and dosing guidelines for an improved interferon alpha 2b formulation, Cancer, 95 (2002) 389-396. https://doi.org/10.1002/cncr.10663.
- [82] K.R. Reddy, M.W. Modi, S. Pedder, Use of peginterferon alfa-2a (40 KD)(Pegasys®) for the treatment of hepatitis C, Adv. Drug Deliv. Rev., 54 (2002) 571-586. https://doi.org/10.1016/S0169-409X(02)00028-5.
- [83] I. Schreiber, M. Buchfelder, M.t. Droste, K. Forssmann, K. Mann, B. Saller, C.J. Strasburger, Treatment of acromegaly with the GH receptor antagonist pegvisomant in clinical practice: safety and efficacy evaluation from the German Pegvisomant Observational Study, Eur. J. Endocrinol., 156 (2007) 75-82. https://doi.org/10.1530/eje.1.02312.
- [84] D.J. D'Amico, Pegaptanib Sodium for Neovascular Age-Related Macular Degeneration: Two-Year Safety Results of the Two Prospective, Multicenter, Controlled Clinical Trials, Ophthalmology, 113 (2006) 992-1001.e1006. https://doi.org/10.1016/j.ophtha.2006.02.027.
- [85] A. Sanchez-Fructuoso, L. Guirado, J. Ruiz, V. Torregrosa, E. Gonzalez, M. Suarez, R. Gallego, Anemia control in kidney transplant patients treated with methoxy polyethylene glycol-epoetin beta (mircera): the Anemiatrans Group, Transplant. Proc., 42 (2010) 2931-2934. https://doi.org/10.1016/j.transproceed.2010.09.012.
- [86] M.P. Curran, P.L. McCormack, Methoxy polyethylene glycol-epoetin beta, ADIS Drug Evaluation, 68 (2008) 1139-1156. https://doi.org/10.2165/00003495-200868080-00009.

- [87] M. Connock, S. Tubeuf, K. Malottki, A. Uthman, J. Round, S. Bayliss, C. Meads, D. Moore, Certolizumab pegol (CIMZIA®) for the treatment of rheumatoid arthritis, Health Technol. Assess., 14 (2010) 1-10. https://doi.org/10.3310/hta14suppl2/01.
- [88] S.N. Alconcel, A.S. Baas, H.D. Maynard, FDA-approved poly (ethylene glycol)—protein conjugate drugs, Polym. Chem., 2 (2011) 1442-1448. https://doi.org/10.1039/C1PY00034A.
- [89] M. Ezban, M.B. Hermit, E. Persson, FIXing postinfusion monitoring: Assay experiences with N9 -
- GP (nonacog beta pegol; Refixia®; Rebinyn®), Haemophilia, 25 (2019) 154-161. https://doi.org/10.1111/hae.13671.
- [90] D.P. Roggeri, E. Zanon, C. Biasoli, A. Roggeri, Extended Half-life rFVIII for the Treatment of Hemophilia A: Drugs Consumption and Patients' Perspective, Farmeconomia. Health Economics and Therapeutic Pathways, 21 (2020) 59-68. https://doi.org/10.7175/fe.v21i1.1472.
- [91] A. Radadiya, W. Zhu, A. Coricello, S. Alcaro, N.G. Richards, Improving the treatment of acute lymphoblastic leukemia, Biochemistry, 59 (2020) 3193-3200. https://doi.org/10.1021/acs.biochem.0c00354.
- [92] R.-J. Li, R. Jin, C. Liu, X. Cao, M.L. Manning, X.M. Di, D. Przepiorka, F. Namuswe, A. Deisseroth, K.B. Goldberg, FDA approval summary: calaspargase pegol-mknl for treatment of acute lymphoblastic leukemia in children and young adults, Clin. Cancer Res., 26 (2020) 328-331. https://doi.org/10.1158/1078-0432.CCR-19-1255.
- [93] A. Markham, Pegvaliase: first global approval, BioDrugs, 32 (2018) 391-395 https://doi.org/10.1007/s40259-018-0292-3.
- [94] M. Ezban, M. Hansen, M. Kjalke, An overview of turoctocog alfa pegol (N8 GP; ESPEROCT®) assay performance: implications for postadministration monitoring, Haemophilia
- 26 (2020) 156-163. https://doi.org/10.1111/hae.13897.
- [95] M. Velinova, A. Bellon, R. Nakov, S. Schussler, S. Schier-Mumzhiu, C. Schelcher, S.D. Koch, A. Skerjanec, J. Wang, A. Krendyukov, Randomized, double-blind, cross-over phase I study comparing pharmacokinetics, pharmacodynamics, safety and immunogenicity of a biosimilar pegfilgrastim with EU and US references, Ann. Oncol., 30 (2019) v738. https://doi.org/10.1093/annonc/mdz265.060.
- [96] B. Finck, H. Tang, F. Civoli, J. Hodge, H. O'Kelly, V. Vexler, Pharmacokinetic and pharmacodynamic equivalence of pegfilgrastim-cbqv and pegfilgrastim in healthy subjects, Adv. Ther., 37 (2020) 4291-4307. https://doi.org/10.1007/s12325-020-01459-y.
- [97] J. Yang, R. Liu, A. Granghaud, O. Zaidi, J. Stephens, Biosimilar pegfilgrastim may offer affordable treatment options for patients in France: a budget impact analysis on the basis of clinical trial and real-world data, J. Med. Econ., 24 (2021) 665-674. https://doi.org/10.1080/13696998.2021.1922252. [98] L. Červinek, Ropeginterferon alfa-2 b for the therapy of polycythemia vera, Vnitr. Lek., 66 (2020) 309-313.
- [99] H. Gisslinger, C. Klade, P. Georgiev, D. Krochmalczyk, L. Gercheva-Kyuchukova, M. Egyed, V. Rossiev, P. Dulicek, H. Pylypenko, L. Sivcheva, PS1457 MAINTENANCE OF RESPONSE IN LONG-TERM TREATMENT WITH ROPEGINTERFERON ALFA-2B (BESREMI®) VS. HYDROXYUREA IN POLYCYTHEMIA VERA PATIENTS (PROUD/CONTINUATION-PV PHASE III TRIALS), HemaSphere, 3 (2019) 670-671. https://doi.org/10.1097/01.HS9.0000564092.98639.85.
- [100] Y.N. Lamb, Lonapegsomatropin: Pediatric First Approval, Paediatr. Drugs, 24 (2022) 83-90. https://doi.org/10.1007/s40272-021-00478-8.
- [101] S.M. Hoy, Pegcetacoplan: First Approval, Drugs, 81 (2021) 1423-1430. https://doi.org/10.1007/s40265-021-01560-8.
- [102] X. Hu, L. Miller, S. Richman, S. Hitchman, G. Glick, S. Liu, Y. Zhu, M. Crossman, I. Nestorov, R.S. Gronke, A novel PEGylated interferon beta 1a for multiple sclerosis: safety, pharmacology, and biology, J. Clin. Pharmacol., 52 (2012) 798-808. https://doi.org/10.1177/0091270011407068.
- [103] T. Ishida, X. Wang, T. Shimizu, K. Nawata, H. Kiwada, PEGylated liposomes elicit an anti-PEG IgM response in a T cell-independent manner, J. Control. Release

- 122 (2007) 349-355. https://doi.org/10.1016/j.jconrel.2007.05.015.
- [104] Y.-C. Hsieh, H.-E. Wang, W.-W. Lin, S.R. Roffler, T.-C. Cheng, Y.-C. Su, J.-J. Li, C.-C. Chen, C.-H. Huang, B.-M. Chen, Pre-existing anti-polyethylene glycol antibody reduces the therapeutic efficacy and pharmacokinetics of PEGylated liposomes, Theranostics, 8 (2018) 3164-3175. https://doi.org/10.7150/thno.22164.
- [105] N.J. Ganson, T.J. Povsic, B.A. Sullenger, J.H. Alexander, S.L. Zelenkofske, J.M. Sailstad, C.P. Rusconi, M.S. Hershfield, Pre-existing anti–polyethylene glycol antibody linked to first-exposure allergic reactions to pegnivacogin, a PEGylated RNA aptamer, J. Allergy Clin. Immunol., 137 (2016) 1610-1613. e1617. https://doi.org/10.1016/j.jaci.2015.10.034.
- [106] Q. Yang, T.M. Jacobs, J.D. McCallen, D.T. Moore, J.T. Huckaby, J.N. Edelstein, S.K. Lai, Analysis of pre-existing IgG and IgM antibodies against polyethylene glycol (PEG) in the general population, Anal. Chem., 88 (2016) 11804-11812. https://doi.org/10.1021/acs.analchem.6b03437.
- [107] Q. Yang, S.K. Lai, Anti PEG immunity: emergence, characteristics, and unaddressed questions, Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol., 7 (2015) 655-677. https://doi.org/10.1002/wnan.1339.
- [108] I. Jakasa, M.M. Verberk, M. Esposito, J.D. Bos, S. Kezic, Altered penetration of polyethylene glycols into uninvolved skin of atopic dermatitis patients, J. Invest. Dermatol., 127 (2007) 129-134. https://doi.org/10.1038/sj.jid.5700582.
- [109] X. Wang, T. Ishida, H. Kiwada, Anti-PEG IgM elicited by injection of liposomes is involved in the enhanced blood clearance of a subsequent dose of PEGylated liposomes, J. Control. Release, 119 (2007) 236-244. https://doi.org/10.1016/j.jconrel.2007.02.010.
- [110] A.W. Richter, E. Åkerblom, Antibodies against polyethylene glycol produced in animals by immunization with monomethoxy polyethylene glycol modified proteins, Int. Arch. Allergy Immu., 70 (1983) 124-131. https://doi.org/10.1159/000233309.
- [111] M. Ichihara, T. Shimizu, A. Imoto, Y. Hashiguchi, Y. Uehara, T. Ishida, H. Kiwada, Anti-PEG IgM response against PEGylated liposomes in mice and rats, Pharmaceutics, 3 (2011) 1-11. https://doi.org/10.3390/pharmaceutics3010001.
- [112] T. Shimizu, M. Ichihara, Y. Yoshioka, T. Ishida, S. Nakagawa, H. Kiwada, Intravenous administration of polyethylene glycol-coated (PEGylated) proteins and PEGylated adenovirus elicits an anti-PEG immunoglobulin M response, Biol. Pharm. Bull., 35 (2012) 1336-1342. https://doi.org/10.1248/bpb.b12-00276.
- [113] J.K. Armstrong, G. Hempel, S. Koling, L.S. Chan, T. Fisher, H.J. Meiselman, G. Garratty, Antibody against poly (ethylene glycol) adversely affects PEG asparaginase therapy in acute lymphoblastic leukemia patients, 110 (2007) 103-111. https://doi.org/10.1517/17425247.2012.720969.
- [114] M.S. Hershfield, N.J. Ganson, S.J. Kelly, E.L. Scarlett, D.A. Jaggers, J.S. Sundy, therapy, Induced and pre-existing anti-polyethylene glycol antibody in a trial of every 3-week dosing of pegloticase for refractory gout, including in organ transplant recipients, Arthritis Res. Ther., 16 (2014) 1-11. https://doi.org/10.1186/ar4500.
- [115] P.E. Lipsky, L.H. Calabrese, A. Kavanaugh, J.S. Sundy, D. Wright, M. Wolfson, M.A. Becker, Pegloticase immunogenicity: the relationship between efficacy and antibody development in patients treated for refractory chronic gout, Arthritis Res. Ther., 16 (2014) 1-8. https://doi.org/10.1186/ar4497.
- [116] N. Longo, C.O. Harding, B.K. Burton, D.K. Grange, J. Vockley, M. Wasserstein, G.M. Rice, A. Dorenbaum, J.K. Neuenburg, D.G. Musson, Single-dose, subcutaneous recombinant phenylalanine ammonia lyase conjugated with polyethylene glycol in adult patients with phenylketonuria: an openlabel, multicentre, phase 1 dose-escalation trial, Lancet, 384 (2014) 37-44. https://doi.org/10.1016/S0140-6736(13)61841-3.
- [117] T. Ishida, K. Masuda, T. Ichikawa, M. Ichihara, K. Irimura, H. Kiwada, Accelerated clearance of a second injection of PEGylated liposomes in mice, Int. J. Pharm., 255 (2003) 167-174. https://doi.org/10.1016/S0378-5173(03)00085-1.

- [118] T. Ishida, R. Maeda, M. Ichihara, K. Irimura, H. Kiwada, Accelerated clearance of PEGylated liposomes in rats after repeated injections, J. Control. Release, 88 (2003) 35-42. https://doi.org/10.1016/S0168-3659(02)00462-5.
- [119] T. Suzuki, Y. Suzuki, T. Hihara, K. Kubara, K. Kondo, K. Hyodo, K. Yamazaki, T. Ishida, H. Ishihara, PEG shedding-rate-dependent blood clearance of PEGylated lipid nanoparticles in mice: Faster PEG shedding attenuates anti-PEG IgM production, Int. J. Pharm., 588 (2020) 119792. https://doi.org/10.1016/j.ijpharm.2020.119792.
- [120] N.E. Elsadek, A.S.A. Lila, S.E. Emam, T. Shimizu, H. Takata, H. Ando, Y. Ishima, T. Ishida, Pegfilgrastim (PEG-G-CSF) induces anti-PEG IgM in a dose dependent manner and causes the accelerated blood clearance (ABC) phenomenon upon repeated administration in mice, Eur. J Pharm. Biopharm., 152 (2020) 56-62. https://doi.org/10.1016/j.ejpb.2020.04.026.
- [121] S.E. Emam, N.E. Elsadek, A.S. Abu Lila, H. Takata, Y. Kawaguchi, T. Shimizu, H. Ando, Y. Ishima, T. Ishida, Anti-PEG IgM production and accelerated blood clearance phenomenon after the administration of PEGylated exosomes in mice, J. Control. Release, 334 (2021) 327-334. https://doi.org/10.1016/j.jconrel.2021.05.001.
- [122] M.M. El Sayed, T. Shimizu, A.S.A. Lila, N.E. Elsadek, S.E. Emam, E. Alaaeldin, A. Kamal, H.A. Sarhan, H. Ando, Y. Ishima, A mouse model for studying the effect of blood anti-PEG IgMs levels on the in vivo fate of PEGylated liposomes, Int. J. Pharm., 615 (2022) 121539. https://doi.org/10.1016/j.ijpharm.2022.121539.
- [123] G. Kozma, T. Shimizu, T. Ishida, J. Szebeni, Anti-PEG antibodies: Properties, formation and role in adverse immune reactions to PEGylated nano-biopharmaceuticals, Adv. Drug Deliv. Rev., 154–155 (2020) 163-175. https://doi.org/10.1016/j.addr.2020.07.024.
- [124] H.F. Smyth, C.P. Carpenter, C.S. Weil, The Chronic Oral Toxicology of the Polyethylene Glycols*, J. Am. Pharm. Assoc., 44 (1955) 27-30. https://doi.org/10.1002/jps.3030440111.
- [125] D.E. Prentice, S.K. Majeed, Oral toxicity of polyethylene glycol (PEG 200) in monkeys and rats, Toxicol. Lett., 2 (1978) 119-122. https://doi.org/10.1016/0378-4274(78)90084-X.
- [126] W. Thiele, L. Kyjacova, A. Köhler, J.P. Sleeman, A cautionary note: Toxicity of polyethylene glycol 200 injected intraperitoneally into mice, Lab. Anim., 54 (2020) 391-396. 10.1177/0023677219873684.
- [127] G. Liu, Y. Li, L. Yang, Y. Wei, X. Wang, Z. Wang, L. Tao, Cytotoxicity study of polyethylene glycol derivatives, RSC adv., 7 (2017) 18252-18259.
- [128] K. Shiraishi, M. Yokoyama, Toxicity and immunogenicity concerns related to PEGylated-micelle carrier systems: a review, Sci. Technol. Adv. Mater., 20 (2019) 324-336. 10.1080/14686996.2019.1590126.
- [129] M.C. Dispenza, Classification of hypersensitivity reactions, Allergy Asthma Proc., 40 (2019) 4274. https://doi.org/10.2500/aap.2019.40.4274.
- [130] P. Lieberman, R.A. Nicklas, C. Randolph, J. Oppenheimer, D. Bernstein, J. Bernstein, A. Ellis, D.B. Golden, P. Greenberger, S. Kemp, Anaphylaxis—a practice parameter update 2015, Ann. Allergy Asthma Immunol., 115 (2015) 341-384. https://doi.org/10.1016/j.anai.2015.07.019.
- [131] A. Uzzaman, S.H. Cho, Classification of hypersensitivity reactions, in: Allergy and Asthma Proceedings, OceanSide Publications, Inc, 2012, pp. S96-S99.https://doi.org/10.2500/aap.2012.33.3561.
- [132] W.J. Pichler, Delayed drug hypersensitivity reactions, Ann. Intern. Med., 139 (2003) 683-693. https://doi.org/10.7326/0003-4819-139-8-200310210-00012.
- [133] J. Szebeni, Complement activation-related pseudoallergy: a stress reaction in blood triggered by nanomedicines and biologicals, Mol. Immunol., 61 (2014) 163-173. https://doi.org/10.1016/j.molimm.2014.06.038.
- [134] G.T. Kozma, T. Mészáros, I. Vashegyi, T. Fülöp, E. Örfi, L. Dézsi, L. Rosivall, Y. Bavli, R. Urbanics, T.E. Mollnes, Y. Barenholz, J. Szebeni, Pseudo-anaphylaxis to Polyethylene Glycol (PEG)-Coated Liposomes: Roles of Anti-PEG IgM and Complement Activation in a Porcine Model of Human Infusion Reactions, ACS Nano, 13 (2019) 9315-9324. https://doi.org/10.1021/acsnano.9b03942.

- [135] P. Bedocs, J. Capacchione, L. Potts, R. Chugani, Z. Weiszhar, J. Szebeni, C.C. Buckenmaier, Hypersensitivity reactions to intravenous lipid emulsion in swine: relevance for lipid resuscitation studies, Anesth. Analg., 119 (2014) 1094-1101. https://doi.org/10.1213/ANE.00000000000396.
- [136] J. Szebeni, Complement activation-related pseudoallergy caused by liposomes, micellar carriers of intravenous drugs, and radiocontrast agents, Crit. Rev. Ther. Drug Carr. Syst., 18 (2001) 40. https://doi.org/10.1615/CritRevTherDrugCarrierSyst.v18.i6.50.
- [137] J.H. Lang, E.C. Lasser, W.P. Kolb, Activation of serum complement by contrast media, Investig. Radiol., 11 (1976) 303-308. https://doi.org/10.1097/00004424-197607000-00007
- [138] J. Szebeni, Hypersensitivity reactions to radiocontrast media: the role of complement activation, Curr. Allergy Asthma Rep., 4 (2004) 25-30. https://doi.org/10.1007/s11882-004-0038-9.
- [139] A. Borderé, A. Stockman, B. Boone, A.S. Franki, M.J. Coppens, H. Lapeere, J. Lambert, A case of anaphylaxis caused by macrogol 3350 after injection of a corticosteroid, Contact Derm., 67 (2012) 376-378. https://doi.org/10.1111/j.1600-0536.2012.02104.x.
- [140] D. Gachoka, Polyethylene glycol (PEG)-induced anaphylactic reaction during bowel preparation, ACG Case Rep. J., 2 (2015) 216. https://doi.org/10.14309/crj.2015.63.
- [141] H. Hyry, A. Vuorio, E. Varjonen, J. Skyttä, S. Mäkinen-Kiljunen, Two cases of anaphylaxis to macrogol 6000 after ingestion of drug tablets, Allergy, 61 (2006) 1021-1023. https://doi.org/10.1111/j.1398-9995.2006.01083.x.
- [142] J. Szebeni, Complement activation-related pseudoallergy caused by amphiphilic drug carriers: the role of lipoproteins, Curr. Drug Deliv., 2 (2005) 443-449. https://doi.org/10.2174/156720105774370212.
- [143] J.J. Verhoef, J.F. Carpenter, T.J. Anchordoquy, H. Schellekens, Potential induction of anti-PEG antibodies and complement activation toward PEGylated therapeutics, Drug Discov., 19 (2014) 1945-1952. https://doi.org/10.1016/j.drudis.2014.08.015.
- [144] J. Szebeni, G. Storm, Complement activation as a bioequivalence issue relevant to the development of generic liposomes and other nanoparticulate drugs, Biochem. Biophys. Res. Commun., 468 (2015) 490-497. https://doi.org/10.1016/j.bbrc.2015.06.177.
- [145] P.L. Turecek, J. Siekmann, PEG-protein conjugates: nonclinical and clinical toxicity considerations, in: Polymer-Protein Conjugates, Elsevier, 2020, pp. 61-101.https://doi.org/10.1016/B978-0-444-64081-9.00004-8.
- [146] D. Shi, D. Beasock, A. Fessler, J. Szebeni, J.Y. Ljubimova, K.A. Afonin, M.A. Dobrovolskaia, To PEGylate or not to PEGylate: immunological properties of nanomedicine's most popular component, poly (ethylene) glycol and its alternatives, Adv. Drug Deliv. Rev., 180 (2021) 114079. https://doi.org/10.1016/j.addr.2021.114079.
- [147] N.J. Ganson, S.J. Kelly, E. Scarlett, J.S. Sundy, M.S. Hershfield, Control of hyperuricemia in subjects with refractory gout, and induction of antibody against poly (ethylene glycol)(PEG), in a phase I trial of subcutaneous PEGylated urate oxidase, Arthritis Res. Ther., 8 (2005) 1-10. https://doi.org/10.1186/ar1861.
- [148] H. Hasan, O.M. Shaikh, S.R. Rassekh, A.F. Howard, K. Goddard, cancer, Comparison of hypersensitivity rates to intravenous and intramuscular PEG asparaginase in children with acute
- lymphoblastic leukemia: a meta analysis and systematic review, Pediatr. Blood Cancer, 64 (2017) 81-88. https://doi.org/10.1002/pbc.26200.
- [149] E.K. Browne, C. Moore, A. Sykes, Z. Lu, S. Jeha, B.N. Mandrell, Clinical characteristics of intravenous PEG-asparaginase hypersensitivity reactions in patients undergoing treatment for acute lymphoblastic leukemia, J. Pediatr. Oncol. Nurs., 35 (2018) 103-109. https://doi.org/10.1177/1043454217741868.
- [150] S. Gupta, K. Lau, C.O. Harding, G. Shepherd, R. Boyer, J.P. Atkinson, V. Knight, J. Olbertz, K. Larimore, Z. Gu, Association of immune response with efficacy and safety outcomes in adults with phenylketonuria administered pegvaliase in phase 3 clinical trials, eBioMedicine, 37 (2018) 366-373. https://doi.org/10.1016/j.ebiom.2018.10.038.

- [151] T.J. Povsic, M.G. Lawrence, A.M. Lincoff, R. Mehran, C.P. Rusconi, S.L. Zelenkofske, Z. Huang, J. Sailstad, P.W. Armstrong, P.G. Steg, Pre-existing anti-PEG antibodies are associated with severe immediate allergic reactions to pegnivacogin, a PEGylated aptamer, J. Allergy Clin. Immunol. Pract., 138 (2016) 1712-1715. https://doi.org/10.1016/j.jaci.2016.04.058.
- [152] G. Ferrandina, M. Ludovisi, D. Lorusso, S. Pignata, E. Breda, A. Savarese, P. Del Medico, L. Scaltriti, D. Katsaros, D. Priolo, Phase III trial of gemcitabine compared with pegylated liposomal doxorubicin in progressive or recurrent ovarian cancer, J. Clin. Oncol., 26 (2008) 890-896. https://doi.org/10.1200/JCO.2007.13.6606.
- [153] D.A. Solimando Jr, J.P. Wilson, Doxorubicin-induced hypersensitivity reactions, Drug Intell. Clin. Pharm., 18 (1984) 808-811. https://doi.org/10.1177/106002808401801007.
- [154] L. Sharma, A. Subedi, B. Shah, Anaphylaxis to pegylated liposomal Doxorubicin: a case report, West Indian Med. J., 63 (2014) 376. https://doi.org/10.7727/wimj.2013.270
- [155] S.C. Semple, T.O. Harasym, K.A. Clow, S.M. Ansell, S.K. Klimuk, M.J. Hope, Immunogenicity and rapid blood clearance of liposomes containing polyethylene glycol-lipid conjugates and nucleic acid, J. Pharmacol. Exp. Ther., 312 (2005) 1020-1026. https://doi.org/10.1124/jpet.104.078113
- [156] A. Judge, K. McClintock, J.R. Phelps, I. MacLachlan, Hypersensitivity and loss of disease site targeting caused by antibody responses to PEGylated liposomes, Mol. Ther., 13 (2006) 328-337. https://doi.org/10.1016/j.ymthe.2005.09.014.
- [157] K.L. Swingle, A.G. Hamilton, M.J. Mitchell, Lipid nanoparticle-mediated delivery of mRNA therapeutics and vaccines, Trends Mol. Med., 27 (2021) 616-617. https://doi.org/10.1038/s41392-020-0164-4.
- [158] D.V. Parums, First full regulatory approval of a COVID-19 vaccine, the BNT162b2 Pfizer-BioNTech vaccine, and the real-world implications for public health policy, Med. Sci. Monit., 27 (2021) e934625-934621. https://doi.org/10.12659/MSM.934625
- [159] E. Mahase, Covid-19: Moderna applies for US and EU approval as vaccine trial reports 94.1% efficacy, in: British Medical Journal 2020.https://doi.org/10.1136/bmj.m4709
- [160] E. Mahase, Covid-19: UK approves Moderna vaccine to be given as two doses 28 days apart, in: British Medical Journal British Medical Journal Publishing Group, 2021.https://doi.org/10.1136/bmj.n74.
- [161] B. Cabanillas, C.A. Akdis, N. Novak, Allergic reactions to the first COVID-19 vaccine: A potential role of polyethylene glycol?, Allergy, 76 (2021) 1617-1618. https://doi.org/10.1111/all.14711.
- [162] J. Kleine-Tebbe, L. Klimek, E. Hamelmann, O. Pfaar, C. Taube, M. Wagenmann, T. Werfel, M. Worm, Severe allergic reactions to the COVID-19 vaccine statement and practical consequences, Allergol. Select, 5 (2021) 26-28. https://doi.org/10.5414/alx02215e.
- [163] CDC, The Vaccine Adverse Event Reporting System (VAERS). http://wonder.cdc.gov/vaers.html Accessed 4 Jan, 2022., in,
- [164] A.R. Wolfson, L.B. Robinson, L. Li, A.E. McMahon, A.S. Cogan, X. Fu, P. Wickner, U. Samarakoon, R.R. Saff, K.G. Blumenthal, A. Banerji, First-Dose mRNA COVID-19 Vaccine Allergic Reactions: Limited Role for Excipient Skin Testing, J. Allergy Clin. Immunol. Pract., 9 (2021) 3308-3320.e3303. https://doi.org/10.1016/j.jaip.2021.06.010.
- [165] P. Sellaturay, S. Nasser, P. Ewan, Polyethylene Glycol-Induced Systemic Allergic Reactions (Anaphylaxis), J Allergy Clin. Immunol. Pract., 9 (2021) 670-675. https://doi.org/10.1016/j.jaip.2020.09.029.
- [166] M.M. Frank, L.F. Fries, The role of complement in inflammation and phagocytosis, Immunol., 12 (1991) 322-326. https://doi.org/10.1016/0167-5699(91)90009-I.
- [167] M. Noris, G. Remuzzi, Overview of complement activation and regulation, Semin. Nephrol., 33 (2013) 479-492. https://doi.org/10.1016/j.semnephrol.2013.08.001.
- [168] J. Szebeni, F. Muggia, A. Gabizon, Y. Barenholz, Activation of complement by therapeutic liposomes and other lipid excipient-based therapeutic products: prediction and prevention, Adv. Drug Deliv. Rev., 63 (2011) 1020-1030. https://doi.org/10.1016/j.addr.2011.06.017.

- [169] T.E. Hugh, Structure and function of the anaphylatoxins, Semin. Immunopathol., 7 (1984) 193-219. https://doi.org/10.1007/BF01893020.
- [170] I. Hamad, A. Hunter, J. Szebeni, S.M. Moghimi, Poly (ethylene glycol) s generate complement activation products in human serum through increased alternative pathway turnover and a MASP-2-dependent process, Mol. Immunol., 46 (2008) 225-232. https://doi.org/10.1016/j.molimm.2008.08.276.
- [171] S. Shah, T. Prematta, N.F. Adkinson, F.T. Ishmael, Hypersensitivity to polyethylene glycols, J. Clin. Pharmacol., 53 (2013) 352-355. https://doi.org/10.1177/0091270012447122.
- [172] L. Bommarito, S. Mietta, F. Nebiolo, M. Geuna, G. Rolla, Macrogol hypersensitivity in multiple drug allergy, Ann. Allergy Asthma Immunol., 107 (2011) 542-543. https://doi.org/10.1016/j.anai.2011.08.008.
- [173] E. Wenande, M. Kroigaard, H. Mosbech, L.H. Garvey, Polyethylene glycols (PEG) and related structures: overlooked allergens in the perioperative setting, Case Rep., 4 (2015) 61-64. https://doi.org/10.1213/XAA.00000000000126.
- [174] E.C. Wenande, P.S. Skov, H. Mosbech, L.K. Poulsen, L.H. Garvey, Inhibition of polyethylene glycol—induced histamine release by monomeric ethylene and diethylene glycol: A case of probable polyethylene glycol allergy, J. Allergy Clin. Immunol., 131 (2013) 1425-1427. https://doi.org/10.1016/j.jaci.2012.09.037.
- [175] C. Sohy, O. Vandenplas, Y. Sibille, Usefulness of oral macrogol challenge in anaphylaxis after intra-articular injection of corticosteroid preparation, Allergy, 63 (2008) 478-479. https://doi.org/10.1111/j.1398-9995.2007.01610.x.
- [176] J. Szebeni, P. Bedőcs, Z. Rozsnyay, Z. Weiszhár, R. Urbanics, L. Rosivall, R. Cohen, O. Garbuzenko, G. Báthori, M. Tóth, Liposome-induced complement activation and related cardiopulmonary distress in pigs: factors promoting reactogenicity of Doxil and AmBisome, Nanomed.: Nanotechnol. Biol. Med., 8 (2012) 176-184. https://doi.org/10.1016/j.nano.2011.06.003.
- [177] M.B. Pedersen, X. Zhou, E.K.U. Larsen, U.S. Sørensen, J. Kjems, J.V. Nygaard, J.R. Nyengaard, R.L. Meyer, T. Boesen, T. Vorup-Jensen, Curvature of synthetic and natural surfaces is an important target feature in classical pathway complement activation, J. Immunol., 184 (2010) 1931-1945. https://doi.org/10.4049/jimmunol.0902214
- [178] L. Dézsi, T. Fülöp, T. Mészáros, G. Szénási, R. Urbanics, C. Vázsonyi, E. Őrfi, L. Rosivall, R. Nemes, R.J. Kok, Features of complement activation-related pseudoallergy to liposomes with different surface charge and PEGylation: comparison of the porcine and rat responses, J. Control. Release, 195 (2014) 2-10. https://doi.org/10.1016/j.jconrel.2014.08.009.
- [179] H. Gao, Q. He, The interaction of nanoparticles with plasma proteins and the consequent influence on nanoparticles behavior, Expert Opin. Drug Deliv., 11 (2014) 409-420. https://doi.org/10.1517/17425247.2014.877442.
- [180] M.M. Yallapu, M.C. Ebeling, N. Chauhan, M. Jaggi, S.C. Chauhan, Interaction of curcumin nanoformulations with human plasma proteins and erythrocytes, Int. J. Nanomed., 6 (2011) 2779-2790. https://doi.org/10.2147/IJN.S25534.
- [181] A. Alinaghi, M. Rouini, F.J. Daha, H. Moghimi, The influence of lipid composition and surface charge on biodistribution of intact liposomes releasing from hydrogel-embedded vesicles, Int. J. Pharm., 459 (2014) 30-39. https://doi.org/10.1016/j.ijpharm.2013.11.011.
- [182] J. Szebeni, C.R. Alving, L. Rosivall, R. Bünger, L. Baranyi, P. Bedöcs, M. Tóth, Y. Barenholz, Animal models of complement-mediated hypersensitivity reactions to liposomes and other lipid-based nanoparticles, J. Liposome Res., 17 (2007) 107-117. https://doi.org/10.1080/08982100701375118.
- [183] J. Szebeni, L. Baranyi, S. Savay, M. Bodo, D.S. Morse, M. Basta, G.L. Stahl, R. Bünger, C.R. Alving, Liposome-induced pulmonary hypertension: properties and mechanism of a complement-mediated pseudoallergic reaction, Am. J. Physiol. Heart Circ. Physiol., 279 (2000) H1319-H1328. https://doi.org/10.1152/ajpheart.2000.279.3.H1319.

- [184] R. Oksjoki, P.T. Kovanen, M.O. Pentikäinen, Role of complement activation in atherosclerosis, Curr. Opin. Lipidol., 14 (2003) 477-482. https://doi.org/10.1097/01.mol.0000092627.86399.7b.
- [185] G. Caracciolo, D. Pozzi, A.L. Capriotti, C. Cavaliere, A. Laganà, Effect of DOPE and cholesterol on the protein adsorption onto lipid nanoparticles, J Nanopart. Res., 15 (2013) 1-11. https://doi.org/10.1007/s11051-013-1498-4.
- [186] M. Kulkarni, A. Flašker, M. Lokar, K. Mrak-Poljšak, A. Mazare, A. Artenjak, S. Čučnik, S. Kralj, A. Velikonja, P. Schmuki, Binding of plasma proteins to titanium dioxide nanotubes with different diameters, Int. J. Nanomed., 10 (2015) 1359-1373. https://doi.org/10.2147/IJN.S77492.
- [187] S.C. Semple, A. Chonn, P.R. Cullis, Interactions of liposomes and lipid-based carrier systems with blood proteins: Relation to clearance behaviour in vivo, Adv. Drug Deliv. Rev., 32 (1998) 3-17. https://doi.org/10.1016/S0169-409X(97)00128-2.
- [188] H. Harashima, T. Huong, T. Ishida, Y. Manabe, H. Matsuo, H. Kiwada, Synergistic effect between size and cholesterol content in the enhanced hepatic uptake clearance of liposomes through complement activation in rats, Pharm. Res., 13 (1996) 1704-1709. https://doi.org/10.1023/A:1016401025747.
- [189] C. Zhang, K. Fan, X. Ma, D. Wei, Impact of large aggregated uricases and PEG diol on accelerated blood clearance of PEGylated canine uricase, PLOS One 7(2012) e39659. https://doi.org/10.1371/journal.pone.0039659.
- [190] I. Badiu, G. Guida, E. Heffler, G. Rolla, Multiple drug allergy due to hypersensitivity to polyethylene glycols of various molecular weights, (2015).
- [191] B.D. Jakubovic, C. Saperia, G.L. Sussman, Anaphylaxis following a transvaginal ultrasound, Allergy Asthma Clin. Immunol., 12 (2016) 1-4. https://doi.org/10.1186/s13223-015-0106-9.
- [192] A.A. Fisher, Immediate and delayed allergic contact reactions to polyethylene glycol, Contact Derm., 4 (1978) 135-138. https://doi.org/10.1111/j.1600-0536.1978.tb03759.x.
- [193] A. Klos, A.J. Tenner, K.-O. Johswich, R.R. Ager, E.S. Reis, J. Köhl, The role of the anaphylatoxins in health and disease, Mol. Immunol., 46 (2009) 2753-2766. https://doi.org/10.1016/j.molimm.2009.04.027.
- [194] H.J. Lenz, Management and preparedness for infusion and hypersensitivity reactions, Oncol. J., 12 (2007) 601-609. https://doi.org/10.1634/theoncologist.12-5-601.
- [195] A. Fisher, Contact urticaria due to polyethylene glycol, Cutis, 19 (1977) 409-412.
- [196] S. Almer, L. Franzen, G. Olaison, K. Smedh, M. Ström, Increased absorption of polyethylene glycol 600 deposited in the colon in active ulcerative colitis, GUT, 34 (1993) 509-513. http://dx.doi.org/10.1136/gut.34.4.509.
- [197] Y.N. Lamb, BNT162b2 mRNA COVID-19 vaccine: first approval, Drugs, 81 (2021) 495-501. https://doi.org/10.1007/s40265-021-01480-7.
- [198] K.S. Corbett, D.K. Edwards, S.R. Leist, O.M. Abiona, S. Boyoglu-Barnum, R.A. Gillespie, S. Himansu, A. Schäfer, C.T. Ziwawo, A.T. DiPiazza, SARS-CoV-2 mRNA vaccine design enabled by prototype pathogen preparedness, Nature, 586 (2020) 567-571. https://doi.org/10.1038/s41586-020-2622-0.
- [199] A. Dadla, S. Tannenbaum, B. Yates, L. Holle, Delayed hypersensitivity reaction related to the use of pegfilgrastim, J. Oncol. Pharm. Pract., 21 (2015) 474-477. https://doi.org/10.1177/1078155214542493.
- [200] E. McCabe, V. Tormey, J.P. Doran, Polyethylene glycol: an underrecognized compound in certolizumab pegol and Movicol that may cause anaphylaxis, Rheumatology, 59 (2020) 908-910. https://doi.org/10.1093/rheumatology/kez469.
- [201] L.T. Henriksen, A. Harila Saari, E. Ruud, J. Abrahamsson, K. Pruunsild, G. Vaitkeviciene, Ó.G.
- Jónsson, K. Schmiegelow, M. Heyman, H. Schrøder, PEG asparaginase allergy in children with acute lymphoblastic leukemia in the NOPHO ALL2008 protocol, Pediatr. Blood Cancer, 62 (2015) 427-433. https://doi.org/10.1002/pbc.25319.

[202] M. Patrawala, M. Kuruvilla, H. Li, Successful desensitization of Pegvaliase (Palynziq®) in a patient with phenylketonuria, Mol. Genet. Metab. Rep., 23 (2020) 100575. https://doi.org/10.1016/j.ymgmr.2020.100575.

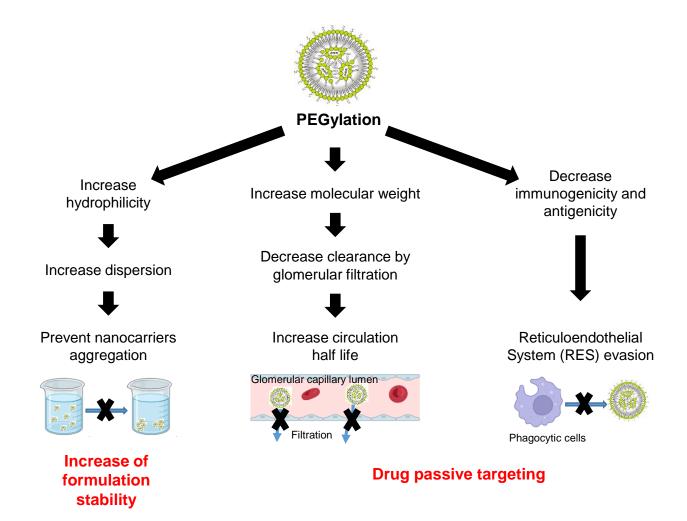
[203] L.R. Sharma, A. Subedi, B.K. Shah, Anaphylaxis to pegylated liposomal Doxorubicin: a case report, West Indian Med. J., 63 (2014) 376. https://doi.org/0.7727/wimj.2013.270.

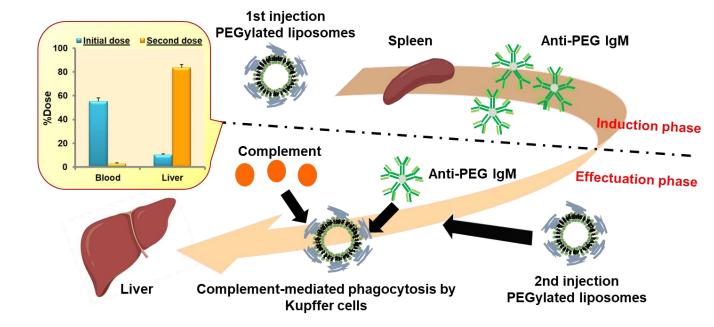
[204] A.C.G. Steffensmeier, A.E. Azar, J.J. Fuller, B.A. Muller, S.R. Russell, Vitreous Injections of Pegaptanib Sodium Triggering Allergic Reactions, Am. J. Ophthalmol., 143 (2007) 512-513. https://doi.org/10.1016/j.ajo.2006.10.007.

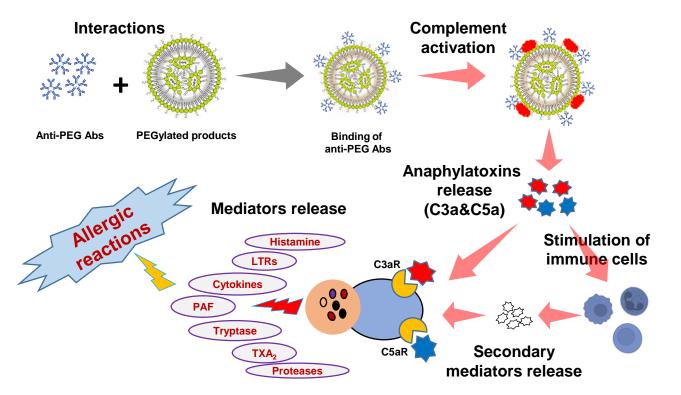
[205] T.J. Povsic, M.G. Lawrence, A.M. Lincoff, R. Mehran, C.P. Rusconi, S.L. Zelenkofske, Z. Huang, J. Sailstad, P.W. Armstrong, P.G. Steg, Pre-existing anti-PEG antibodies are associated with severe immediate allergic reactions to pegnivacogin, a PEGylated aptamer, J. Allergy Clin. Immunol., 138 (2016) 1712-1715. https://doi.org/10.1016/j.jaci.2016.04.058.

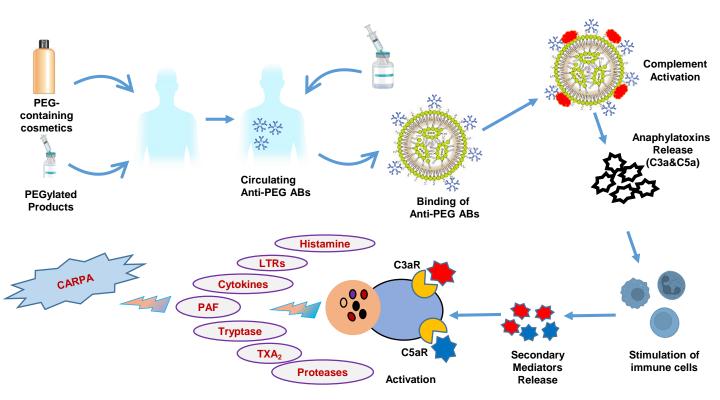
Figures Legends

- Figure 1. Chemical formula for Polyethylene glycol
- Figure 2. The effect of PEGylation on the formulation's stability and drug passive targeting
- Figure 3. Mechanism for the ABC phenomenon in PEGylated liposomes
- Figure 4. Mechanism for PEG-induced anaphylactic reactions









Abbreviations: ABs, Antibodies; C3a and C5a, complement fragments; C3a R and C5a R, complement fragments receptors; LTRs, Leukotrienes; PAF, Platelet activating factor; TXA2, Thromboxane A2; CARPA, complement activation-related pseudoallergy.